



*Int J Mol Sci.* 2013 Jul; 14(7): 14575–14593.

Published online 2013 Jul 12. doi: [10.3390/ijms140714575](https://doi.org/10.3390/ijms140714575)

PMCID: PMC3742260

PMID: [23857055](#)

## Melatonin in Alzheimer's Disease

Li Lin,<sup>1,2,†</sup> Qiong-Xia Huang,<sup>3,†</sup> Shu-Sheng Yang,<sup>2</sup> Jiang Chu,<sup>1</sup> Jian-Zhi Wang,<sup>1,\*</sup> and Qing Tian<sup>1,\*</sup>

<sup>1</sup>Key Laboratory of Neurological Disease of National Education Ministry and Hubei Province, Department of Pathology and Pathophysiology, Tongji Medical College, Huazhong University of Science and Technology, Wuhan 430030, China; E-Mail: [chujiang@hust.edu.cn](mailto:chujiang@hust.edu.cn)

<sup>2</sup>Department of Pathology and Pathophysiology, College of Medical Science, Jishou University, 120 People Road, Jishou 436100, China; E-Mails: [linli@jsu.edu.cn](mailto:linli@jsu.edu.cn) (L.L.); [yangshusheng@jsu.edu.cn](mailto:yangshusheng@jsu.edu.cn) (S.-S.Y.)

<sup>3</sup>Department of TCM Rationale, College of Basic Medicine, Hubei University of Chinese Medicine, 1 West Road Huangjia Lake, Wuhan 430065, China; E-Mail: [huangqx@hbtcm.edu.cn](mailto:huangqx@hbtcm.edu.cn)

†These authors contributed equally to this work.

\*Authors to whom correspondence should be addressed; E-Mails: [wangjz@mails.tongji.edu.cn](mailto:wangjz@mails.tongji.edu.cn) (J.-Z.W.); [tianq@mail.hust.edu.cn](mailto:tianq@mail.hust.edu.cn) (Q.T.); Tel./Fax: +86-27-8369-3883 (J.-Z.W.); Tel.: +86-27-8369-2625 (Q.T.).

Received 2013 Jan 9; Revised 2013 Jun 21; Accepted 2013 Jul 5.

**Copyright** © 2013 by the authors; licensee MDPI, Basel, Switzerland

This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution license (<http://creativecommons.org/licenses/by/3.0/>).

## Abstract

Alzheimer's disease (AD), an age-related neurodegenerative disorder with progressive cognition deficit, is characterized by extracellular senile plaques (SP) of aggregated  $\beta$ -amyloid (A $\beta$ ) and intracellular neurofibrillary tangles, mainly containing the hyperphosphorylated microtubule-associated protein tau. Multiple factors contribute to the etiology of AD in terms of initiation and progression. Melatonin is an endogenously produced hormone in the brain and decreases during aging and in patients with AD. Data from clinical trials indicate that melatonin supplementation improves sleep, ameliorates sundowning and slows down the progression of cognitive impairment in AD patients. Melatonin efficiently protects neuronal cells from A $\beta$ -mediated toxicity via antioxidant and anti-amyloid properties. It not only inhibits A $\beta$  generation, but also arrests the formation of amyloid fibrils by a structure-dependent interaction with A $\beta$ . Our studies have demonstrated that melatonin efficiently attenuates Alzheimer-like tau hyperphosphorylation. Although the exact mechanism is still not fully understood, a direct regulatory influence of melatonin on the activities of protein kinases and protein phosphatases is proposed. Additionally, melatonin also plays a role in protecting the cholinergic system and in anti-inflammation. The aim of this review is to stimulate interest in melatonin as a potentially useful agent in the prevention and treatment of AD.

**Keywords:** Alzheimer's disease, melatonin, tau hyperphosphorylation, beta amyloid, antioxidation, cholinergic, neuroinflammation

## 1. Introduction

Alzheimer's disease (AD) is an age-associated neurodegenerative disease and characterized by progressive loss of cognition and other neurobehavioral manifestations. Pathological hallmarks of AD include extracellular senile plaques (SP), mainly consisting of  $\beta$ -amyloid ( $A\beta$ ), and intracellular neurofibrillary tangles (NFTs), mainly composed of abnormally hyperphosphorylated tau, a microtubule-associated protein [1]. In spite of a large number of studies undertaken, the etiology of AD is largely unknown. Many mechanisms have been proposed, including genetic predispositions (e.g., expression levels and subforms of presenilins (PS) and Apolipoprotein (Apo) E), inflammatory processes associated with cytokine releasing, oxidative stress and neurotoxicity by metal ions [2–6].

Melatonin (*N*-acetyl-5-methoxytryptamine), a tryptophan metabolite and synthesized mainly in the pineal gland, has a number of physiological functions, including regulating circadian rhythms, clearing free radicals, improving immunity and generally inhibiting the oxidation of biomolecules. Decreased melatonin in serum and cerebrospinal fluid (CSF) and the loss of melatonin diurnal rhythm are observed in AD patients [7–12]. Furthermore, the level of melatonin in CSF decreases with the progression of AD neuropathology, as determined by the Braak stages [12]. Melatonin levels both in CSF and in postmortem human pineal gland are already reduced in preclinical AD subjects, who are cognitively still intact and have only the earliest signs of AD neuropathology [8,12]. A strong correlation exists between pineal content and CSF level of melatonin [8] and between CSF and plasma melatonin levels [7], suggesting that a reduced CSF melatonin level may serve as an early marker for the very first stages of AD. In mammals, melatonin exerts some of its functions through two specific high-affinity membrane receptors, melatonin receptor 1 (MT1) and melatonin receptor 2 (MT2). Decreased MT2 immunoreactivity and increased MT1 immunoreactivity have been reported in the hippocampus of AD patients [13,14]. Although the pineal gland of AD patients has molecular changes, no changes in pineal weight, calcification or total protein content have been observed [8,15]. It is also shown that  $\beta 1$ -adrenergic receptor mRNA disappeared, and the activity and gene expression of monoamine oxidase (MAO) were upregulated in AD patients, suggesting that the dysregulation of noradrenergic innervations and the depletion of serotonin, the precursor of melatonin, might be responsible for the loss of melatonin rhythm and reduced melatonin levels in AD [16]. Melatonin supplementation has been suggested to improve circadian rhythmicity, for example, decreasing agitated behavior, confusion and “sundowning”, and to produce beneficial effects on memory in AD patients [17–21]. Therefore, melatonin supplementation, with its marked low toxicity [22–24], may be one of the possible strategies for symptomatic treatment.

In AD,  $A\beta$  is generally believed to play an important role in promoting neuronal degeneration by rendering neurons more vulnerable to age-related increases in levels of oxidative stress and impairments in cellular energy metabolism [25]. As the major microtubule-associated protein, tau promotes microtubule assembly and stabilizes microtubules. Hyperphosphorylation will obviously reduce the abilities of tau, which leads to cytoskeletal arrangement disruption [26,27]. The extent of neurofibrillary pathology, and particularly the number of cortical NFT, correlates positively with the severity of dementia [28]. As melatonin is able to improve some of the clinical symptoms of AD, and the level of melatonin decreases dramatically during AD, studies on the relationship between melatonin and AD pathology will be helpful to assess its potential in the prevention or treatment of AD. In this review, we will address the role of melatonin in tau hyperphosphorylation and  $A\beta$  toxicity. As cholinergic deficit and inflammation are involved in AD pathogenesis, the protection of melatonin on the cholinergic system and inflammation are also introduced. Each part is described, from phenomenon observation to mechanism investigation and speculation.

## 2. Melatonin in Tau Hyperphosphorylation

*Int J Mol Sci*

Hyperphosphorylated tau has been identified in more than a dozen of neurodegenerative disorders, termed tauopathies, including AD, Niemann-Pick type C disease, and so on [29–31]. Among these tau-related disorders, AD is the most common and the best-studied tauopathy. In AD brains, the level of the hyperphosphorylated tau is 3–4-fold higher than that of tau from normal adult brains [32,33]. There are 79 putative serine or threonine phosphorylation sites in the longest human brain tau isoform, and more than 30 phosphorylation sites have been identified in AD brain, including Thr39, Ser46, Thr50, Thr69, Thr153, Thr175, Thr181, Ser184, Ser185, Ser195, Ser198, Ser199, Ser202, Thr205, Ser208, Ser210, Thr212, Ser214, Thr217, Thr231, Ser235, Ser237, Ser238, Ser245, Ser258, Ser262, Ser285, Ser293, Ser305, Ser320, Ser324, Ser352, Ser356, Thr377, Ser396, Ser400, Thr403, Ser404, Ser409, Ser412, Ser413, Ser416 and Ser422 [34–39]. The spreading of tau pathology in the brain is the hallmark of AD pathogenesis, and the number of NFTs is positively correlated with the clinical cognitive deficit of AD patients [40].

Inhibition of tau hyperphosphorylation is one target in AD treatment. Thus, we systematically studied the effect of melatonin on tau hyperphosphorylation and found that melatonin efficiently attenuates tau hyperphosphorylation induced by wortmannin [41], calyculin A (CA) [42–44] and okadaic acid [45] in N2a and SH-SY5Y neuroblastoma cells. It was further demonstrated that melatonin significantly ameliorated tau hyperphosphorylation elicited by wortmannin [46], isoproterenol [47,48], CA [44] and constant light illumination [24] in rats. To elucidate the mechanisms underlying the inhibitory effect of melatonin on tau hyperphosphorylation, alterations of the activities of protein kinases and phosphatases were detected. Melatonin treatment not only inhibited wortmannin-induced glycogen synthase kinase-3 (GSK-3) activation, isoproterenol-induced protein kinase A (PKA) activation and CA-induced protein phosphatase-2A (PP-2A) inactivation, but also antagonized the oxidative stress induced by these agents [46,49,50]. These results from our studies provide evidence for the strong efficacy of melatonin in inhibiting tau hyperphosphorylation.

To explore whether a decrease of melatonin would induce the alteration of tau phosphorylation, we inhibited melatonin biosynthesis by brain injection of haloperidol, an inhibitor of 5-hydroxyindole-*O*-methyltransferase (one key enzyme in melatonin synthesis) in rats [51]. It was found that inhibition of melatonin biosynthesis not only resulted in spatial memory impairment in rats, but also induced an increase in tau phosphorylation with a concomitant decrease in PP-2A activity. Supplementation with melatonin by prior injection for one week and reinforcement during the haloperidol administration period significantly improved memory retention deficits, arrested tau hyperphosphorylation and oxidative stress and restored PP-2A activity [51]. We also used constant illumination to interrupt melatonin metabolism in rats. Concomitant with decreased serum melatonin, the constantly illuminated rats developed spatial memory deficits, tau hyperphosphorylation at multiple sites, activation of GSK-3 and PKA, as well as suppression of PP-1. Prominent oxidative damage and organelle lesions, demonstrated by increased expression of endoplasmic reticulum (ER) stress-related proteins, including immunoglobulin-binding protein (BiP)/GRP78 and CHOP/GADD153, decreased the number of rough ER and free ribosome, resulted in thinner synapses and increased superoxide dismutase and MAO, which were also observed in the light exposed rats. Simultaneous supplementation of melatonin partially arrested the behavioral and molecular impairments [24]. Although it is unclear whether diminished melatonin concentration is one causative factor in AD pathology or only a secondary process, our results strongly implicate the important role of decreased melatonin in Alzheimer-like spatial memory impairment and tau hyperphosphorylation.

Chemical agents used in our studies, including wortmannin, isoproterenol and CA, not only induced tau phosphorylation, but also initiated oxidative stress, as manifested by an elevated level of malondialdehyde and an altered activity of superoxide dismutase (SOD) [41–43]. Furthermore, melatonin is a potent direct free radical scavenger and indirect antioxidant that acts by augmenting the activity of several important antioxidative enzymes, such as SOD, glutathione peroxidase and

glutathione reductase [52]. Constant illumination not only induced the decreased serum melatonin level, but also the increased levels of SOD and MAO in rats [24]. Oxidative stress is known to influence the phosphorylation state of tau [53–55]. The accumulation of misfolded and aggregated proteins in neurons of AD brain was considered to be related to oxidative stress, along with its molecular structure changes with aging [56]. As an antioxidant and free radical scavenger [57–59], melatonin prevents the overproduction of free radicals and reduces neuronal damage resulting from a variety of pathological processes [60–62]. It is therefore possible that prevention against tau phosphorylation by melatonin is partially due to its antioxidant activity.

Some studies also indicated melatonin may act as an enzyme modulator in a way that is unrelated to its antioxidant properties. Accumulating data provide evidence for the regulation by melatonin of protein kinases, including PKA [63,64], protein kinase C (PKC) [65,66], Ca<sup>2+</sup>/calmodulin-dependent kinase II (CaMKII) [67,68] and the mitogen-activated protein kinase (MAPK) family [69–72]. The documented correlation between melatonin and cAMP indicates that melatonin might inhibit PKA activity through the melatonin receptor coupled inhibition of adenylyl cyclase and reduction of cAMP [63,64]. Although there is no evidence of a direct relationship between melatonin and GSK-3 activity, one of our studies has found that melatonin treatment revised constant light illumination induced GSK-3 activation in the brain of rats [24].

### 3. Melatonin and A $\beta$ Toxicity

A $\beta$ , composing 39–43 amino acid residues derived from its precursor, amyloid precursor protein (APP), plays a pivotal role in the pathogenesis of AD [25,73]. Mature APP is processed proteolytically by distinct  $\alpha$ -secretase or  $\beta$ -secretase pathways [74]. The nonamyloidogenic  $\alpha$ -pathway involves the cleavage of APP within the A $\beta$  sequence by  $\alpha$ -secretase to release an N-terminal APP fragment, which, in turn, is cleaved by  $\gamma$ -secretase. Thus, the cleavage by  $\gamma$ -secretase precludes the formation of A $\beta$ . The amyloidogenic  $\beta$ -secretase pathway, which results in the formation of intact A $\beta$  peptide, is mediated by the sequential cleavage of  $\beta$ -secretase and  $\gamma$ -secretases at the N- and C-terminals of the A $\beta$  sequence, respectively [75]. Melatonin has been found to inhibit normal levels of secretion of soluble APP (sAPP) in different cell lines by interfering with APP full maturation [76]. Additionally, administration of melatonin efficiently reduced A $\beta$  generation and deposition both *in vivo* [77,78] and *in vitro* [76,79–81]. We have demonstrated that melatonin reduces A $\beta$  generation in mouse neuroblastoma N2a cells harboring APP695 [80,82].

However, an *in vivo* study showed that melatonin did not affect the expression of APP holoprotein in transgenic Tg2576 mice [77]. Furthermore, despite achieving high plasma concentrations of melatonin, chronic melatonin therapy in old Tg2576 mice initiated at 14 months of age not only failed to remove existing plaques, but also failed to prevent additional A $\beta$  deposition [83]. This result is in contrast with those of diminished A $\beta$  in melatonin-treated wild-type mice [78] and reduced A $\beta$  and protein nitration in melatonin-treated Tg2576 mice [77]. The initiation time of melatonin treatment might account for the difference between the studies of Matsubara *et al.* [77] and Quinn *et al.* [83], in which the same transgenic Tg2576 mouse model was used. Amyloid plaque pathology typically appears in 10–12-month old Tg2576 mice [84]. Melatonin treatment in the study of Matsubara *et al.* was started when the mice were four months old (prior to the appearance of hippocampal and cortical plaques) [77], an earlier pathological stage compared with 14 months of age in the study of Quinn *et al.* [83]. However, both studies concur in finding little evidence of the potent antioxidant effects of melatonin in the oldest mice. These findings indicate that melatonin has the ability to regulate APP metabolism and prevent A $\beta$  pathology, but fails to exert anti-amyloid or antioxidant effects when initiated after A $\beta$  deposition. Although consistent conclusions were achieved, none of the related studies further explain how melatonin exerts its inhibitory effect on A $\beta$  generation. The proteolytic cleavage of APP by the  $\alpha$ -secretase pathway is regulated by many physiological and pathological stimuli and the PKC-dependent

mechanism is one of the most recognized. Stimuli, such as muscarinic and metabotropic glutamate receptor agonists and phorbol esters, share the capacity to stimulate soluble APP secretion and inhibit A $\beta$  formation through PKC activation [75]. The mechanism whereby PKC activity increases soluble APP secretion is still unknown, but it may involve additional kinase steps and the eventual activation of the secretases that mediated APP cleavage. Recently, the inhibitory regulation by GSK-3 inhibition on A $\beta$  generation has been well-established [85–87]. The mechanism behind this is not clear. It was demonstrated that inhibition of GSK-3 and upregulation of c-Jun N-terminal kinase (JNK) result in elevated matrix metalloprotease activity and increased degradation of A $\beta$  [88]. As phosphorylation of GSK-3 leads to its inactivation, the data suggest that activated GSK-3 may inhibit or reduce JNK activation by certain stimuli [89]. GSK-3 interacts with presenilin-1 (PS1), a cofactor for  $\gamma$ -secretase, implying that GSK-3 may function as a component in the  $\gamma$ -secretase complex [90,91]. Assuming that melatonin can influence PKC and GSK-3 activity as mentioned earlier, it is postulated that melatonin may regulate APP processing through the PKC and GSK-3 pathways. As PKC is an upstream regulator of GSK-3, GSK-3 may be one of the common signal pathways increasing A $\beta$  generation and tau hyperphosphorylation. Regulation of the A $\beta$  fibril formation and the important pathological property of A $\beta$ , such as neurotoxicity and resistance to proteolytic degradation, depend on the ability of peptides to form  $\beta$ -sheet structures and/or amyloid fibrils [92,93].

Intervention in the A $\beta$  aggregation process can be considered an approach to stopping or slowing the progression of AD. Melatonin can interact with A $\beta$ 40 and A $\beta$ 42 and inhibit the progressive formation of  $\beta$ -sheet and/or amyloid fibrils [94–96]. The anti-fibrillogenic effect of melatonin has been demonstrated by different techniques, including circular dichroism (CD) spectroscopy, electron microscopy and nuclear magnetic resonance (NMR) spectroscopy and electrospray ionization mass spectrometry (ESI-MS) [95]. Moreover, the interaction between melatonin and A $\beta$  appears to depend on the structural characteristics of melatonin, rather than on its antioxidant properties, because it could not be reproduced by melatonin analogs or other free radical scavengers [92,94]. Evidence derived from ESI-MS proved that there was a hydrophobic interaction between A $\beta$  and melatonin, and proteolytic investigations suggested that the interaction took place on the 29–40 residues of the A $\beta$  segment [95]. Results from NMR spectroscopy further confirmed a residue-specific interaction between melatonin and any of the three histidine and aspartate residues of A $\beta$  [96]. The imidazole-carboxylate salt bridges formed by the side chains of histidine (His $^+$ ) and aspartate (Asp $^-$ ) residues are critical to the formation of the amyloid  $\beta$ -sheet structures [97], and disruption of these salt bridges promotes fibril dissolution [98].

Melatonin could promote the conversion of  $\beta$ -sheets into random coils by disrupting the imidazole-carboxylate salt bridges and, thus, prevent A $\beta$  fibrillogenesis and aggregation. It is therefore possible that by blocking the formation of the secondary  $\beta$ -sheet conformation, melatonin may not only reduce neurotoxicity, but also facilitate clearance of the peptide via increased proteolytic degradation. It has been demonstrated that melatonin directly interacts with A $\beta$  and prevents its aggregation [99]. A $\beta$  treatment elicits a spectrum of cellular damage, including increases in lipid peroxidation and intracellular free calcium concentration, oxidative damage to mitochondrial DNA and the emergence of apoptotic markers [100]. Mitochondria are not only the primary site of reactive oxygen species (ROS) generation, but also the primary target of attack for ROS. Melatonin was considered to stabilize the fluidity of mitochondrial inner membranes; and binding to mitochondrial membranes was revealed [101]. Oxidative stress acts synergistically with the disturbance of intracellular calcium homeostasis. The free radical-induced membrane damage induces further calcium influx, and the resultant accentuated calcium influx, in turn, will induce the generation of further free radicals. Therefore, oxidative stress plays a central role in A $\beta$ -induced neurotoxicity and even cell death. Aside from A $\beta$  causing oxidative stress, it has been proposed that oxidative damage could exacerbate a vicious cycle, in which amyloidogenic processing of APP would be further facilitated to generate more A $\beta$ , which, in turn, enhances oxidative stress [102]. Accumulating data implies that melatonin efficiently protects

*Int J Mol Sci*

cells against A $\beta$ -induced oxidative damage and cell death *in vitro* [103,104] and *in vivo* [77,105–107]. In A $\beta$ -treated cells and animals, melatonin exerts its protective activity mainly through an antioxidant effect, whereas in APP-transfected cells and transgenic animal models, the underlying mechanism is attributed to not only its antioxidant property, but also its anti-amyloid property, including inhibition of both A $\beta$  generation and formation of  $\beta$ -sheets and/or amyloid fibrils.

Additionally, some findings suggest a role for perturbed melatonin signaling in the sleep disturbances that are common in AD patients [108–110]. By *in vivo* microdialysis in Tg2576 mice, it was found that the amount of brain interstitial fluid (ISF) A $\beta$  correlated with wakefulness. Additionally, the ISF A $\beta$  also significantly increased during acute sleep deprivation, but decreased with infusion of a dual orexin receptor antagonist. Chronic sleep restriction significantly increased A $\beta$  plaque formation [111]. Furthermore, administration of melatonin efficiently reduced A $\beta$  generation and deposition, both *in vivo* and *in vitro* [76–81]. Thus, melatonin may inhibit A $\beta$  generation and loading, but the mechanism needs further investigation.

#### 4. Protection of Melatonin on the Cholinergic System

Cholinergic system deficit is also an early and primary event in the pathogenesis of AD [112]. Neurons in the nucleus basalis of Meynert, a major source of cholinergic innervation of the cerebral cortex and hippocampus, undergo a profound and selective degeneration in AD brain [113–115]. The level of acetylcholine (ACh) is decreased in the early stage of AD, whereas the activities of the synthesizing enzyme, choline acetyltransferase (ChAT), and the hydrolyzing enzyme, acetylcholinesterase (AChE), do not change until the late stage of AD [116–118]. Other biological investigations of tissue from biopsy and autopsy have found a profound decrease of ChAT activity in the neocortex of AD patients, correlating positively with the severity of dementia [102]. Although the mechanism leading to the ACh deficit is still unknown, the inhibitor of AChE has been employed as a treatment and is considered the standard of care for the treatment for mild-to-moderate AD [119].

Melatonin has protective effects on the cholinergic system. A previous study has also demonstrated that melatonin partially prevented peroxynitrite-induced inhibition of choline transport and ChAT activity in several neuronal proteins from synaptosomes and, more readily, from synaptic vesicles [120]. Additionally, it is reported that four-month melatonin treatment significantly ameliorated the neuropathological, behavioral and biochemical changes in eight-month-old APP695 transgenic mice with A $\beta$  deposition, significant learning and memory deficit and a profound reduction in ChAT activity in the frontal cortex and hippocampus [105]. Another study also showed that similar treatment with melatonin antagonized spatial memory deficit and decreased ChAT activity in ovariectomized adult rats [121]. However, in rats infused intracerebroventricularly with A $\beta$  for 14 days, where ChAT activity was significantly reduced, melatonin was unable to restore the activity of this enzyme [122]. Melatonin showed the inhibition only on the lipopolysaccharide (LPS)-induced increase in AChE activity, whereas no changes were observed in CSF treated rats. These results are in support of the inhibitory influence of melatonin on AChE activity in streptozotocin-induced dementia [123].

Compared with placebo, the cholinesterase inhibitors, such as donepezil, tacrine, rivastigmine and galantamine, which can resolve acetylcholine deprivation, are able to stabilize or slow decline in cognition, function, behavior and global change [124]. Melatonin secretion decreases in AD, and this decrease has been postulated as responsible for the circadian disorganization, decrease in sleep efficiency and impaired cognitive function seen in those patients. Melatonin replacement has been shown effective to treat sundowning, mild cognitive impairment (MCI), an etiologically heterogeneous syndrome that precedes dementia, and other sleep wake disorders in AD patients. Besides inhibition on AChE activity, the prospects for melatonin as a treatment for AD are also based on scavenging of reactive oxygen and nitrogen species, resolving sleep disturbance, decreasing A $\beta$  toxicity and loading [125]. There is no clinical evidence to show which is better for AD patients regarding the AChE

inhibitor and melatonin. However, the combination of these two drugs may have much better effects. Recently tacrine-melatonin hybrids were designed and synthesized as new multifunctional drug candidates for AD [126,127]. The compounds show improved cholinergic and antioxidant properties, being more potent and selective inhibitors of human AChE than tacrine and capturing free radicals better than melatonin. They exhibit low toxicity and may be able to penetrate the central nervous system [126]. Direct intracerebral administration of one of these hybrids, *N*-(2-(1H-indol-3-yl)ethyl)-7-(1,2,3,4-tetrahydroacridin-9-ylamino) heptanamide, decreased A $\beta$ -induced cell death and amyloid burden in the brain parenchyma of APP/PS1 mice. Furthermore, the reduction in A $\beta$  pathology was accompanied by a recovery in cognitive function [127].

## 5. Role of Melatonin in Neuroinflammation of AD

A common factor in AD pathogeny is the over activation of microglia with the consequent over expression of proinflammatory cytokines [128–130]. The accumulation of A $\beta$  in plaques, as well as A $\beta$  oligomers may produce sequential inflammatory/oxidative events and excitotoxicity, causing neurodegeneration and cognitive impairment [131]. Furthermore, the epidemiological studies have shown that non-steroidal anti-inflammatory drug (NSAID) use decreases the incidence of AD [132]. A $\beta$  has been shown to act as a proinflammatory agent, activating many inflammatory components, and SP, surrounded by microglia and astrocytes, coexist with cytokines and chemokines [133]. The A $\beta$ -induced activation of microglia is thought to be one of the major sources of the inflammatory response [134]. It has been reported that melatonin attenuates kainic acid induced microglial and astrogliat responses, as determined by immunohistochemical detection of isolectin-B4 and glial fibrillary acidic protein (GFAP), the specific markers for microglia and astroglia, respectively [135]. Oral melatonin administration also attenuated A $\beta$ -induced proinflammatory cytokines, nuclear factor- $\kappa$ B (NF- $\kappa$ B) and nitric oxide (NO) in the rat brain [107].

It is reported that melatonin significantly reduced the proinflammatory response, decreasing by nearly 50% the A $\beta$ -induced levels of proinflammatory cytokines, Interleukin-1- $\beta$  (IL1- $\beta$ ), Interleukin-6 (IL6) and tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ), *in vivo* [107]. Furthermore, NF- $\kappa$ B DNA binding activity was inhibited by melatonin [136,137]. More recently, it has been demonstrated that melatonin reduces NF- $\kappa$ B-induced IL-6 in a concentration-dependent manner in A $\beta$ -treated brain slices [138]. Melatonin administration is also reported to reduce A $\beta$ -induced learning and memory impairment in rats, along with a significant decrease in positive glial cells expressing NF- $\kappa$ B-induced IL-1 $\beta$  in addition to complement 1q (C1q) in hippocampus [139].

## 6. Conclusions

Melatonin is one of the most powerful antioxidants acting at various levels, and the level of melatonin reduces during aging and in AD patients [24,40,140–142]. Additionally, its indirect antioxidant effects and anti-amyloid effects are based on the support of appropriate circadian phasing and anti-excitotoxic actions [68,143]. Thus, it is not surprising that melatonin is protective in numerous experimental systems and has been proposed as a treatment for AD. Recent studies from APP transgenic mice have indicated that early, long-term melatonin supplementation produces anti-amyloid and antioxidant effects, but no such effect is produced when melatonin treatment is initiated after the age of amyloid formation [76,77,79,80]. Extensive clinical trials and studies with transgenic models are necessary to confirm the role of melatonin at the late pathological stage of AD. If melatonin has no effect at the late stage of AD, studies on melatonin should be limited to the prevention of AD, rather than treatment.

Adverse reactions of melatonin may occur, such as fever on the first day of melatonin treatment, hyperkinesia or complaints of restless legs, menorrhagia, pigmentation on arms and legs, headache and abdominal reactions, thrombosis and drowsiness [16,17,144,145]. Apart from these adverse reactions, early and long-term application of melatonin may at least slow down the development of AD. Besides

the positive effects in experimental systems concerning antagonism of cholinergic deficit, inflammation, fibrillogenesis and tangle formation, the sleep-promoting effects and the suppression of sundowning are important results justifying the use melatonin. Although there is evidence to postulate melatonin as a useful and therapeutic tool in MCI and AD, larger double-blind multicenter studies are urgently needed to further explore and investigate the potential and usefulness of melatonin. As decreased MT2 immunoreactivity and increased MT1 immunoreactivity have been reported in the hippocampus of AD patients [13,14], specific melatonin receptor regulators and new melatonin derivatives are expected urgently.

## Acknowledgments

This work was supported by grants from the Natural Science Foundation of China (30971204, 81271404).

## Conflict of Interest

The authors declare no conflict of interest.

## References

1. Lars M.I., Jürgen G. Amyloid-β and tau—A toxic pas de deux in Alzheimer's disease. *Nat. Rev. Neurosci.* 2011;12:67–72. [\[PubMed\]](#) [\[Google Scholar\]](#)
2. Mustapic M., Popovic Hadzija M., Pavlovic M., Pavkovic P., Presecki P., Mrazovac D., Mimica N., Korolija M., Pivac N., Muck-Seler D. Alzheimer's disease and type 2 diabetes: The association study of polymorphisms in tumor necrosis factor-alpha and apolipoprotein E genes. *Metab. Brain Dis.* 2012;27:507–512. [\[PubMed\]](#) [\[Google Scholar\]](#)
3. Ajala T., Rafi J., Wray R., Whitehead M.W., Zaidi J. There may be a link between intrahepatic cholestasis of pregnancy and familial combined hyperlipidaemia: A case report. *Cases J.* 2009;2 doi: 10.4076/1757-1626-2-8679. [\[PMC free article\]](#) [\[PubMed\]](#) [\[CrossRef\]](#) [\[Google Scholar\]](#)
4. Leszek J., Sochocka M., Gasiorowski K. Vascular factors and epigenetic modifications in the pathogenesis of Alzheimer's disease. *J. Neurol. Sci.* 2012;323:25–32. [\[PubMed\]](#) [\[Google Scholar\]](#)
5. Rocchi A., Valensin D., Aldinucci C., Giani G., Barbucci R., Gaggelli E., Kozlowski H., Valensin G. NMR metabolomic investigation of astrocytes interacted with Abeta(42) or its complexes with either copper(II) or zinc(II). *J. Inorg. Biochem.* 2012;117:326–333. [\[PubMed\]](#) [\[Google Scholar\]](#)
6. Rukhsana S., Butterfield D.A. Role of oxidative stress in the progression of Alzheimer's disease. *J. Alzheimers Dis.* 2010;19:341–353. [\[PubMed\]](#) [\[Google Scholar\]](#)
7. Wu Y.H., Swaab D.F. The human pineal gland and melatonin in aging and Alzheimer's disease. *J. Pineal Res.* 2005;38:145–152. [\[PubMed\]](#) [\[Google Scholar\]](#)
8. Wu Y.H., Feenstra M.G., Zhou J.N., Liu R.Y., Torano J.S., van Kan H.J., Fischer D.F., Ravid R., Swaab D.F. Molecular changes underlying reduced pineal melatonin levels in Alzheimer's disease: Alterations in preclinical and clinical stages. *J. Clin. Endocr. Metab.* 2003;88:5898–5906. [\[PubMed\]](#) [\[Google Scholar\]](#)
9. Ferrari E., Arcaini A., Gornati R., Pelanconi L., Cravello L., Fioravanti M., Solerte S.B., Magri F. Pineal and pituitary-adrenocortical function in physiological aging and in senile dementia. *Exp. Gerontol.* 2000;35:1239–1250. [\[PubMed\]](#) [\[Google Scholar\]](#)

10. Ohashi Y., Okamoto N., Uchida K., Iyo M., Mori N., Morita Y. Daily rhythm of serum melatonin levels and effect of light exposure in patients with dementia of the Alzheimer's type. *Biol. Psychiat.* 1999;45:1646–1652. [[PubMed](#)] [[Google Scholar](#)]
  11. Liu R.Y., Zhou J.N., van Heerikhuijze J., Hofman M.A., Swaab D.F. Decreased melatonin levels in postmortem cerebrospinal fluid in relation to aging, Alzheimer's disease, and apolipoprotein E-epsilon4/4 genotype. *J. Clin. Endocr. Metab.* 1999;84:323–327. [[PubMed](#)] [[Google Scholar](#)]
  12. Zhou J.N., Liu R.Y., Kamphorst W., Hofman M.A., Swaab D.F. Early neuropathological Alzheimer's changes in aged individuals are accompanied by decreased cerebrospinal fluid melatonin levels. *J. Pineal Res.* 2003;35:125–130. [[PubMed](#)] [[Google Scholar](#)]
  13. Savaskan E., Olivieri G., Meier F., Brydon L., Jockers R., Ravid R., Wirz-Justice A., Muller-Spahn F. Increased melatonin 1a-receptor immunoreactivity in the hippocampus of Alzheimer's disease patients. *J. Pineal Res.* 2002;32:59–62. [[PubMed](#)] [[Google Scholar](#)]
  14. Savaskan E., Ayoub M.A., Ravid R., Angeloni D., Fraschini F., Meier F., Eckert A., Muller-Spahn F., Jockers R. Reduced hippocampal MT2 melatonin receptor expression in Alzheimer's disease. *J. Pineal Res.* 2005;38:10–16. [[PubMed](#)] [[Google Scholar](#)]
  15. Friedland R.P., Luxenberg J.S., Koss E. A quantitative study of intracranial calcification in dementia of the Alzheimer's type. *Int. Psychogeriatr.* 1990;2:36–43. [[PubMed](#)] [[Google Scholar](#)]
  16. Wu Y.H., Fischer D.F., Swaab D.F. A promoter polymorphism in the monoamine oxidase A gene is associated with the pineal MAOA activity in Alzheimer's disease patients. *Brain Res.* 2007;1167:13–19. [[PubMed](#)] [[Google Scholar](#)]
  17. Cohen-Mansfield J., Garfinkel D., Lipson S. Melatonin for treatment of sundowning in elderly persons with dementia—A preliminary study. *Arch. Gerontol. Geriatr.* 2000;31:65–76. [[PubMed](#)] [[Google Scholar](#)]
  18. Brusco L.I., Marquez M., Cardinali D.P. Melatonin treatment stabilizes chronobiologic and cognitive symptoms in Alzheimer's disease. *Neuro Endocrinol. Lett.* 2000;21:39–42. [[PubMed](#)] [[Google Scholar](#)]
  19. Brusco L.I., Marquez M., Cardinali D.P. Monozygotic twins with Alzheimer's disease treated with melatonin: Case report. *J. Pineal Res.* 1998;25:260–263. [[PubMed](#)] [[Google Scholar](#)]
  20. Cardinali D.P., Brusco L.I., Perez Lloret S., Furio A.M. Melatonin in sleep disorders and jet-lag. *Neuro Endocrinol. Lett.* 2002;23:9–13. [[PubMed](#)] [[Google Scholar](#)]
  21. Cardinali D.P., Brusco L.I., Liberczuk C., Furio A.M. The use of melatonin in Alzheimer's disease. *Neuro Endocrinol. Lett.* 2002;23:20–23. [[PubMed](#)] [[Google Scholar](#)]
  22. Karasek M., Reiter R.J., Cardinali D.P., Pawlikowski M. Future of melatonin as a therapeutic agent. *Neuro Endocrinol. Lett.* 2002;23:118–121. [[PubMed](#)] [[Google Scholar](#)]
  23. Singer C., Tractenberg R.E., Kaye J., Schafer K., Gamst A., Grundman M., Thomas R., Thal L.J. Alzheimer's disease cooperative, SA multicenter, placebo-controlled trial of melatonin for sleep disturbance in Alzheimer's disease. *Sleep.* 2003;26:893–901. [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)]
  24. Ling Z.Q., Tian Q., Wang L., Fu Z.Q., Wang X.C., Wang Q., Wang J.Z. Constant illumination induces Alzheimer-like damages with endoplasmic reticulum involvement and the protection of melatonin. *J. Alzheimers Dis.* 2009;16:287–300. [[PubMed](#)] [[Google Scholar](#)]

25. Selkoe D.J. Cell biology of protein misfolding: The examples of Alzheimer's and Parkinson's diseases. *Nat. Cell Biol.* 2004;6:1054–1061. [[PubMed](#)] [[Google Scholar](#)]
26. Brion J.P., Anderton B.H., Authelet M., Dayanandan R., Leroy K., Lovestone S., Octave J.N., Pradier L., Touchet N., Tremp G. Neurofibrillary tangles and tau phosphorylation. *Biochem. Soc. Symp.* 2001;67:81–88. [[PubMed](#)] [[Google Scholar](#)]
27. Billingsley M.L., Kincaid R.L. Regulated phosphorylation and dephosphorylation of tau protein: Effects on microtubule interaction, intracellular trafficking and neurodegeneration. *Biochem. J.* 1997;323:577–591. [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)]
28. Braak E., Braak H., Mandelkow E.M. A sequence of cytoskeleton changes related to the formation of neurofibrillary tangles and neuropil threads. *Acta Neuropathol.* 1994;87:554–567. [[PubMed](#)] [[Google Scholar](#)]
29. Avila J., Perez M., Lucas J.J., Gomez-Ramos A., Santa Maria I., Moreno F., Smith M., Perry G., Hernandez F. Assembly *in vitro* of tau protein and its implications in Alzheimer's disease. *Curr. Alzheimer's Res.* 2004;1:97–101. [[PubMed](#)] [[Google Scholar](#)]
30. Sahara N., DeTure M., Ren Y., Ebrahim A.S., Kang D., Knight J., Volbracht C., Pedersen J.T., Dickson D.W., Yen S.H., et al. Characteristics of TBS-extractable hyperphosphorylated Tau species: Aggregation intermediates in rTg4510 mouse brain. *J. Alzheimers Dis.* 2013;33:249–263. [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)]
31. Lei P., Ayton S., Finkelstein D.I., Spoerri L., Ciccotosto G.D., Wright D.K., Wong B.X., Adlard P.A., Cherny R.A., Lam L.Q., et al. Tau deficiency induces parkinsonism with dementia by impairing APP-mediated iron export. *Nat. Med.* 2012;18:291–295. [[PubMed](#)] [[Google Scholar](#)]
32. Khatoon S., Grundke-Iqbali I., Iqbal K. Brain levels of microtubule-associated protein tau are elevated in Alzheimer's disease: A radioimmuno-slot-blot assay for nanograms of the protein. *J. Neurochem.* 1992;59:750–753. [[PubMed](#)] [[Google Scholar](#)]
33. Khatoon S., Grundke-Iqbali I., Iqbal K. Levels of normal and abnormally phosphorylated tau in different cellular and regional compartments of Alzheimer's disease and control brains. *FEBS Lett.* 1994;351:80–84. [[PubMed](#)] [[Google Scholar](#)]
34. Hanger D.P., Betts J.C., Loviny T.L., Blackstock W.P., Anderton B.H. New phosphorylation sites identified in hyperphosphorylated tau (paired helical filament-tau) from Alzheimer's disease brain using nanoelectrospray mass spectrometry. *J. Neurochem.* 1998;71:2465–2476. [[PubMed](#)] [[Google Scholar](#)]
35. Hanger D.P., Byers H.L., Wray S., Leung K.Y., Saxton M.J., Seereeram A., Reynolds C.H., Ward M.A., Anderton B.H. Novel phosphorylation sites in tau from Alzheimer's brain support a role for casein kinase 1 in disease pathogenesis. *J. Biol. Chem.* 2007;282:23645–23654. [[PubMed](#)] [[Google Scholar](#)]
36. Hasegawa M., Morishima-Kawashima M., Takio K., Suzuki M., Titani K., Ihara Y. Protein sequence and mass spectrometric analyses of tau in the Alzheimer's disease brain. *J. Biol. Chem.* 1992;267:17047–1754. [[PubMed](#)] [[Google Scholar](#)]
37. Morishima-Kawashima M., Hasegawa M., Takio K., Suzuki M., Yoshida H., Watanabe A., Titani K., Ihara Y. Hyperphosphorylation of tau in PHF. *Neurobiol. Aging.* 1995;16:365–371. [[PubMed](#)] [[Google Scholar](#)]

38. Wang X.F., Dong C.F., Zhang J., Wan Y.Z., Li F., Huang Y.X., Han L., Shan B., Gao C., Han J., et al. Human tau protein forms complex with PrP and some GSS- and fCJD-related PrP mutants possess stronger binding activities with tau *in vitro*. *Mol. Cell. Biochem.* 2008;310:49–55. [[PubMed](#)] [[Google Scholar](#)]
39. Peng C.X., Hu J., Liu D., Hong X.P., Wu Y.Y., Zhu L.Q., Wang J.Z. Disease-modified glycogen synthase kinase-3beta intervention by melatonin arrests the pathology and memory deficits in an Alzheimer's animal model. *Neurobiol. Aging*. 2013;34:1555–1563. [[PubMed](#)] [[Google Scholar](#)]
40. Peter T.N., Irina A., Eileen H.B., Constantin B., Heiko B., Nigel J.C., Rudolph J.C., Barbara J.C., Peter D., Kelly D.T., et al. Correlation of Alzheimer's disease neuropathologic changes with cognitive status: A review of the literature. *J. Neuropathol. Exp. Neurol.* 2012;71:362–381. [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)]
41. Deng Y.Q., Xu G.G., Duan P., Zhang Q., Wang J.Z. Effects of melatonin on wortmannin-induced tau hyperphosphorylation. *Acta Pharmacol. Sin.* 2005;26:519–526. [[PubMed](#)] [[Google Scholar](#)]
42. Li X.C., Wang Z.F., Zhang J.X., Wang Q., Wang J.Z. Effect of melatonin on calyculin A-induced tau hyperphosphorylation. *Eur. J. Pharmacol.* 2005;510:25–30. [[PubMed](#)] [[Google Scholar](#)]
43. Li S.P., Deng Y.Q., Wang X.C., Wang Y.P., Wang J.Z. Melatonin protects SH-SY5Y neuroblastoma cells from calyculin A-induced neurofilament impairment and neurotoxicity. *J. Pineal Res.* 2004;36:186–191. [[PubMed](#)] [[Google Scholar](#)]
44. Yang X., Yang Y., Fu Z., Li Y., Feng J., Luo J., Zhang Q., Wang Q., Tian Q. Melatonin ameliorates Alzheimer-like pathological changes and spatial memory retention impairment induced by calyculin A. *J. Psychopharmacol.* 2011;25:1118–1125. [[PubMed](#)] [[Google Scholar](#)]
45. Wang Y.P., Li X.T., Liu S.J., Zhou X.W., Wang X.C., Wang J.Z. Melatonin ameliorated okadaic-acid induced Alzheimer-like lesions. *Acta Pharmacol. Sin.* 2004;25:276–280. [[PubMed](#)] [[Google Scholar](#)]
46. Liu S.J., Wang J.Z. Alzheimer-like tau phosphorylation induced by wortmannin *in vivo* and its attenuation by melatonin. *Acta Pharmacol. Sin.* 2002;23:183–187. [[PubMed](#)] [[Google Scholar](#)]
47. Wang D.L., Ling Z.Q., Cao F.Y., Zhu L.Q., Wang J.Z. Melatonin attenuates isoproterenol-induced protein kinase A overactivation and tau hyperphosphorylation in rat brain. *J. Pineal Res.* 2004;37:11–16. [[PubMed](#)] [[Google Scholar](#)]
48. Wang X.C., Zhang J., Yu X., Han L., Zhou Z.T., Zhang Y., Wang J.Z. Prevention of isoproterenol-induced tau hyperphosphorylation by melatonin in the rat. *Acta Pharmacol. Sin.* 2005;26:7–12. [[PubMed](#)] [[Google Scholar](#)]
49. Avila J. Tau aggregation into fibrillar polymers: Taupathies. *FEBS Lett.* 2000;476:89–92. [[PubMed](#)] [[Google Scholar](#)]
50. Gong C.X., Liu F., Grundke-Iqbali I., Iqbal K. Post-translational modifications of tau protein in Alzheimer's disease. *J. Neural Transm.* 2005;112:813–838. [[PubMed](#)] [[Google Scholar](#)]
51. Zhu L.Q., Wang S.H., Ling Z.Q., Wang D.L., Wang J.Z. Effect of inhibiting melatonin biosynthesis on spatial memory retention and tau phosphorylation in rat. *J. Pineal Res.* 2004;37:71–77. [[PubMed](#)] [[Google Scholar](#)]
52. Reiter R.J., Acuna-Castroviejo D., Tan D.X., Burkhardt S. Free radical-mediated molecular damage. Mechanisms for the protective actions of melatonin in the central nervous system. *Ann. N. Y. Acad. Sci.* 2001;939:200–215. [[PubMed](#)] [[Google Scholar](#)]

53. Zhu X., Rottkamp C.A., Boux H., Takeda A., Perry G., Smith M.A. Activation of p38 kinase links tau phosphorylation, oxidative stress, and cell cycle-related events in Alzheimer's disease. *J. Neuropath. Exp. Neur.* 2000;59:880–888. [PubMed] [Google Scholar]

54. Gomez-Ramos A., Diaz-Nido J., Smith M.A., Perry G., Avila J. Effect of the lipid peroxidation product acrolein on tau phosphorylation in neural cells. *J. Neurosci. Res.* 2003;71:863–870. [PubMed] [Google Scholar]

55. Lovell M.A., Xiong S., Xie C., Davies P., Markesberry W.R. Induction of hyperphosphorylated tau in primary rat cortical neuron cultures mediated by oxidative stress and glycogen synthase kinase-3. *J. Alzheimers Dis.* 2004;6:659–671. [PubMed] [Google Scholar]

56. Kenyon C.J. The genetics of ageing. *Nature*. 2010;464:504–512. [PubMed] [Google Scholar]

57. Paradies G., Petrosillo G., Paradies V., Reiter R.J., Ruggiero F.M. Melatonin, cardiolipin and mitochondrial bioenergetics in health and disease. *J. Pineal Res.* 2010;48:297–310. [PubMed] [Google Scholar]

58. Romero A., Egea J., Garcia A.G., Lopez M.G. Synergistic neuroprotective effect of combined low concentrations of galantamine and melatonin against oxidative stress in SH-SY5Y neuroblastoma cells. *J. Pineal Res.* 2010;49:141–148. [PubMed] [Google Scholar]

59. Hardeland R., Tan D.X., Reiter R.J. Kynuramines, metabolites of melatonin and other indoles: The resurrection of an almost forgotten class of biogenic amines. *J. Pineal Res.* 2009;47:109–126. [PubMed] [Google Scholar]

60. Jou M.J., Peng T.I., Hsu L.F., Jou S.B., Reiter R.J., Yang C.M., Chiao C.C., Lin Y.F., Chen C.C. Visualization of melatonin's multiple mitochondrial levels of protection against mitochondrial Ca(2+)-mediated permeability transition and beyond in rat brain astrocytes. *J. Pineal Res.* 2010;48:20–38. [PubMed] [Google Scholar]

61. Hong Y., Palaksha K.J., Park K., Park S., Kim H.D., Reiter R.J., Chang K.T. Melatonin plus exercise-based neurorehabilitative therapy for spinal cord injury. *J. Pineal Res.* 2010;49:201–209. [PubMed] [Google Scholar]

62. Das A., McDowell M., Pava M.J., Smith J.A., Reiter R.J., Woodward J.J., Varma A.K., Ray S.K., Banik N.L. The inhibition of apoptosis by melatonin in VSC4.1 motoneurons exposed to oxidative stress, glutamate excitotoxicity, or TNF-alpha toxicity involves membrane melatonin receptors. *J. Pineal Res.* 2010;48:157–169. [PMC free article] [PubMed] [Google Scholar]

63. Schuster C., Williams L.M., Morris A., Morgan P.J., Barrett P. The human MT1 melatonin receptor stimulates cAMP production in the human neuroblastoma cell line SH-SY5Y cells via a calcium-calmodulin signal transduction pathway. *J. Neuroendocrinol.* 2005;17:170–178. [PubMed] [Google Scholar]

64. Peschke E., Muhlbauer E., Musshoff U., Csernus V.J., Chankiewitz E., Peschke D. Receptor (MT(1)) mediated influence of melatonin on cAMP concentration and insulin secretion of rat insulinoma cells INS-1. *J. Pineal Res.* 2002;33:63–71. [PubMed] [Google Scholar]

65. Witt-Enderby P.A., MacKenzie R.S., McKeon R.M., Carroll E.A., Bordt S.L., Melan M.A. Melatonin induction of filamentous structures in non-neuronal cells that is dependent on expression of the human mt1 melatonin receptor. *Cell Motil. Cytoskeleton*. 2000;46:28–42. [PubMed] [Google Scholar]

- Int J Mol Sci | <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3742260/> | 13/18
66. Rivera-Bermudez M.A., Gerdin M.J., Earnest D.J., Dubocovich M.L. Regulation of basal rhythmicity in protein kinase C activity by melatonin in immortalized rat suprachiasmatic nucleus cells. *Neurosci. Lett.* 2003;346:37–40. [\[PubMed\]](#) [\[Google Scholar\]](#)
67. Benitez-King G., Rios A., Martinez A., Anton-Tay F. *In vitro* inhibition of  $\text{Ca}^{2+}$ /calmodulin-dependent kinase II activity by melatonin. *Biochim. Biophys. Acta.* 1996;1290:191–196. [\[PubMed\]](#) [\[Google Scholar\]](#)
68. Chen S., Xu Y., Xu B., Guo M., Zhang Z., Liu L., Ma H., Chen Z., Luo Y., Huang S., Chen L. CaMKII is involved in cadmium activation of MAPK and mTOR pathways leading to neuronal cell death. *J. Neurochem.* 2011;119:1108–1118. [\[PMC free article\]](#) [\[PubMed\]](#) [\[Google Scholar\]](#)
69. Chan A.S., Lai F.P., Lo R.K., Voyno-Yasenetskaya T.A., Stanbridge E.J., Wong Y.H. Melatonin m1 and MT2 receptors stimulate c-Jun N-terminal kinase via pertussis toxin-sensitive and -insensitive G proteins. *Cell. Signal.* 2002;14:249–257. [\[PubMed\]](#) [\[Google Scholar\]](#)
70. Chen L., Xu B., Liu L., Luo Y., Yin J., Zhou H., Chen W., Shen T., Han X., Huang S. Hydrogen peroxide inhibits mTOR signaling by activation of AMPKalpha leading to apoptosis of neuronal cells. *Lab Invest.* 2010;90:762–773. [\[PMC free article\]](#) [\[PubMed\]](#) [\[Google Scholar\]](#)
71. Chen L., Xu B., Liu L., Luo Y., Zhou H., Chen W., Shen T., Han X., Kontos C.D., Huang S. Cadmium induction of reactive oxygen species activates the mTOR pathway, leading to neuronal cell death. *Free Radic. Biol. Med.* 2011;50:624–632. [\[PMC free article\]](#) [\[PubMed\]](#) [\[Google Scholar\]](#)
72. Xu B., Chen S., Luo Y., Chen Z., Liu L., Zhou H., Chen W., Shen T., Han X., Chen L., Huang S. Calcium signaling is involved in cadmium-induced neuronal apoptosis via induction of reactive oxygen species and activation of MAPK/mTOR network. *PLoS One.* 2011;6:e19052. [\[PMC free article\]](#) [\[PubMed\]](#) [\[Google Scholar\]](#)
73. Selkoe D.J. Alzheimer's disease is a synaptic failure. *Science.* 2002;298:789–791. [\[PubMed\]](#) [\[Google Scholar\]](#)
74. Selkoe D.J. The cell biology of beta-amyloid precursor protein and presenilin in Alzheimer's disease. *Trends Cell Biol.* 1998;8:447–453. [\[PubMed\]](#) [\[Google Scholar\]](#)
75. Fisher A., Pittel Z., Haring R., Bar-Ner N., Kliger-Spatz M., Natan N., Egozi I., Sonego H., Marcovitch I., Brandeis R. M1 muscarinic agonists can modulate some of the hallmarks in Alzheimer's disease: Implications in future therapy. *J. Mol. Neurosci.* 2003;20:349–356. [\[PubMed\]](#) [\[Google Scholar\]](#)
76. Lahiri D.K. Melatonin affects the metabolism of the beta-amyloid precursor protein in different cell types. *J. Pineal Res.* 1999;26:137–146. [\[PubMed\]](#) [\[Google Scholar\]](#)
77. Matsubara E., Bryant-Thomas T., Pacheco Quinto J., Henry T.L., Poeggeler B., Herbert D., Cruz-Sanchez F., Chyan Y.J., Smith M.A., Perry G., et al. Melatonin increases survival and inhibits oxidative and amyloid pathology in a transgenic model of Alzheimer's disease. *J. Neurochem.* 2003;85:1101–1108. [\[PubMed\]](#) [\[Google Scholar\]](#)
78. Lahiri D.K., Chen D., Ge Y.W., Bondy S.C., Sharman E.H. Dietary supplementation with melatonin reduces levels of amyloid beta-peptides in the murine cerebral cortex. *J. Pineal Res.* 2004;36:224–231. [\[PubMed\]](#) [\[Google Scholar\]](#)
79. Song W., Lahiri D.K. Melatonin alters the metabolism of the beta-amyloid precursor protein in the neuroendocrine cell line PC12. *J. Mol. Neurosci.* 1997;9:75–92. [\[PubMed\]](#) [\[Google Scholar\]](#)

80. Zhang Y.C., Wang Z.F., Wang Q., Wang Y.P., Wang J.Z. Melatonin attenuates beta-amyloidinduced inhibition of neurofilament expression. *Acta Pharmacol. Sin.* 2004;25:447–451. [PubMed] [Google Scholar]
81. Olivieri G., Hess C., Savaskan E., Ly C., Meier F., Baysang G., Brockhaus M., Muller-Spahn F. Melatonin protects SHSY5Y neuroblastoma cells from cobalt-induced oxidative stress, neurotoxicity and increased beta-amyloid secretion. *J. Pineal Res.* 2001;31:320–325. [PubMed] [Google Scholar]
82. Wang X.C., Zhang Y.C., Chatterjie N., Grundke-Iqbali I., Iqbal K., Wang J.Z. Effect of melatonin and melatonylvalpromide on beta-amyloid and neurofilaments in N2a cells. *Neurochem. Res.* 2008;33:1138–1144. [PubMed] [Google Scholar]
83. Quinn J., Kulhanek D., Nowlin J., Jones R., Pratico D., Rokach J., Stackman R. Chronic melatonin therapy fails to alter amyloid burden or oxidative damage in old Tg2576 mice: Implications for clinical trials. *Brain Res.* 2005;1037:209–213. [PubMed] [Google Scholar]
84. Hsiao K., Chapman P., Nilsen S., Eckman C., Harigaya Y., Younkin S., Yang F., Cole G. Correlative memory deficits, Abeta elevation, and amyloid plaques in transgenic mice. *Science.* 1996;274:99–102. [PubMed] [Google Scholar]
85. Su Y., Ryder J., Li B., Wu X., Fox N., Solenberg P., Brune K., Paul S., Zhou Y., Liu F., et al. Lithium, a common drug for bipolar disorder treatment, regulates amyloid-beta precursor protein processing. *Biochemistry.* 2004;43:6899–6908. [PubMed] [Google Scholar]
86. Ryder J., Su Y., Liu F., Li B., Zhou Y., Ni B. Divergent roles of GSK-3 and CDK5 in APP processing. *Biochem. Biophys. Res. Commun.* 2003;312:922–929. [PubMed] [Google Scholar]
87. Phiel C.J., Wilson C.A., Lee V.M., Klein P.S. GSK-3alpha regulates production of Alzheimer's disease amyloid-beta peptides. *Nature.* 2003;423:435–439. [PubMed] [Google Scholar]
88. Donnelly P.S., Caragounis A., Du T., Laughton K.M., Volitakis I., Cherny R.A., Sharples R.A., Hill A.F., Li Q.X., Masters C.L., et al. Selective intracellular release of copper and zinc ions from bis (thiosemicarbazone) complexes reduces levels of Alzheimer's disease amyloid-beta peptide. *J. Biol. Chem.* 2008;283:4568–4577. [PubMed] [Google Scholar]
89. White A.R., Du T., Laughton K.M., Volitakis I., Sharples R.A., Xilinas M.E., Hoke D.E., Holsinger R.M., Evin G., Cherny R.A., et al. Degradation of the Alzheimer's disease amyloid beta-peptide by metal-dependent up-regulation of metalloprotease activity. *J. Biol. Chem.* 2006;281:17670–17680. [PubMed] [Google Scholar]
90. Tesco G., Tanzi R.E. GSK-3 beta forms a tetrameric complex with endogenous PS1-CTF/NTF and beta-catenin. Effects of the D257/D385A and FAD-linked mutations. *Ann. N. Y. Acad. Sci.* 2000;920:227–232. [PubMed] [Google Scholar]
91. Takashima A., Murayama M., Murayama O., Kohno T., Honda T., Yasutake K., Nihonmatsu N., Mercken M., Yamaguchi H., Sugihara S., et al. Presenilin 1 associates with glycogen synthase kinase-3beta and its substrate tau. *Proc. Natl. Acad. Sci. USA.* 1998;95:9637–9641. [PMC free article] [PubMed] [Google Scholar]
92. Simmons L.K., May P.C., Tomaselli K.J., Rydel R.E., Fuson K.S., Brigham E.F., Wright S., Lieberburg I., Becker G.W., Brems D.N., et al. Secondary structure of amyloid beta peptide correlates with neurotoxic activity *in vitro*. *Mol. Pharmacol.* 1994;45:373–379. [PubMed] [Google Scholar]
93. Soto C., Castano E.M. The conformation of Alzheimer's beta peptide determines the rate of amyloid formation and its resistance to proteolysis. *Biochem. J.* 1996;314:701–707. [PMC free article] [PubMed] [Google Scholar]

94. Poeggeler B., Miravalle L., Zagorski M.G., Wisniewski T., Chyan Y.J., Zhang Y., Shao H., Bryant-Thomas T., Vidal R., Frangione B., et al. Melatonin reverses the profibrillogenic activity of apolipoprotein E4 on the Alzheimer's amyloid Abeta peptide. *Biochemistry*. 2001;40:14995–5001. [\[PubMed\]](#) [\[Google Scholar\]](#)
95. Skribanek Z., Balaspiri L., Mak M. Interaction between synthetic amyloid-beta-peptide (1–40) and its aggregation inhibitors studied by electrospray ionization mass spectrometry. *J. Mass Spectrom.* 2001;36:1226–1229. [\[PubMed\]](#) [\[Google Scholar\]](#)
96. Pappolla M., Bozner P., Soto C., Shao H., Robakis N.K., Zagorski M., Frangione B., Ghiso J. Inhibition of Alzheimer's beta-fibrillogenesis by melatonin. *J. Biol. Chem.* 1998;273:7185–7188. [\[PubMed\]](#) [\[Google Scholar\]](#)
97. Huang T.H., Fraser P.E., Chakrabarty A. Fibrillogenesis of Alzheimer's Abeta peptides studied by fluorescence energy transfer. *J. Mol. Biol.* 1997;269:214–224. [\[PubMed\]](#) [\[Google Scholar\]](#)
98. Fraser P.E., Nguyen J.T., Surewicz W.K., Kirschner D.A. pH-dependent structural transitions of Alzheimer's amyloid peptides. *Biophys. J.* 1991;60:1190–1201. [\[PMC free article\]](#) [\[PubMed\]](#) [\[Google Scholar\]](#)
99. Masilamoni J.G., Jesudason E.P., Dhandayuthapani S., Ashok B.S., Vignesh S., Jebaraj W.C., Paul S.F., Jayakumar R. The neuroprotective role of melatonin against amyloid beta peptide injected mice. *Free Radic. Res.* 2008;42:661–673. [\[PubMed\]](#) [\[Google Scholar\]](#)
100. Pacchierotti C., Iapichino S., Bossini L., Pieraccini F., Castrogiovanni P. Melatonin in psychiatric disorders: A review on the melatonin involvement in psychiatry. *Front. Neuroendocrin.* 2001;22:18–32. [\[PubMed\]](#) [\[Google Scholar\]](#)
101. Yuan H., Pang S.F. [<sup>125</sup>I]Iodomelatonin-binding sites in the pigeon brain: Binding characteristics, regional distribution and diurnal variation. *J. Endocrinol.* 1991;128:475–482. [\[PubMed\]](#) [\[Google Scholar\]](#)
102. Bieschke J., Zhang Q., Powers E.T., Lerner R.A., Kelly J.W. Oxidative metabolites accelerate Alzheimer's amyloidogenesis by a two-step mechanism, eliminating the requirement for nucleation. *Biochemistry*. 2005;44:4977–4983. [\[PubMed\]](#) [\[Google Scholar\]](#)
103. Feng Z., Zhang J.T. Protective effect of melatonin on beta-amyloid-induced apoptosis in rat astrogloma C6 cells and its mechanism. *Free Radic. Biol. Med.* 2004;37:1790–1801. [\[PubMed\]](#) [\[Google Scholar\]](#)
104. Zatta P., Tognon G., Carampin P. Melatonin prevents free radical formation due to the interaction between beta-amyloid peptides and metalions [Al(III), Zn(II), Cu(II), Mn(II), Fe(II)]. *J. Pineal Res.* 2003;35:98–103. [\[PubMed\]](#) [\[Google Scholar\]](#)
105. Feng Z., Chang Y., Cheng Y., Zhang B.L., Qu Z.W., Qin C., Zhang J.T. Melatonin alleviates behavioral deficits associated with apoptosis and cholinergic system dysfunction in the APP 695 transgenic mouse model of Alzheimer's disease. *J. Pineal Res.* 2004;37:129–136. [\[PubMed\]](#) [\[Google Scholar\]](#)
106. Shen Y.X., Xu S.Y., Wei W., Sun X.X., Liu L.H., Yang J., Dong C. The protective effects of melatonin from oxidative damage induced by amyloid beta-peptide 25–35 in middle-aged rats. *J. Pineal Res.* 2002;32:85–89. [\[PubMed\]](#) [\[Google Scholar\]](#)
107. Rosales-Corral S., Tan D.X., Reiter R.J., Valdivia-Velazquez M., Martinez-Barboza G., Acosta-Martinez J.P., Ortiz G.G. Orally administered melatonin reduces oxidative stress and proinflammatory cytokines induced by amyloid-beta peptide in rat brain: a comparative, *in vivo* study *versus* vitamin C

- and *E. J. Pineal Res.* 2003;35:80–84. [[PubMed](#)] [[Google Scholar](#)]
108. Slats D., Claassen J.A., Verbeek M.M., Overeem S. Reciprocal interactions between sleep, circadian rhythms and Alzheimer's disease: Focus on the role of hypocretin and melatonin. *Ageing Res. Rev.* 2013;12:188–200. [[PubMed](#)] [[Google Scholar](#)]
109. Rothman S.M., Mattson M.P. Sleep disturbances in Alzheimer's and Parkinson's diseases. *Neuromolecular Med.* 2012;14:194–204. [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)]
110. Cecon E., Markus R.P. Relevance of the chronobiological and non-chronobiological actions of melatonin for enhancing therapeutic efficacy in neurodegenerative disorders. *Recent Pat. Endocr. Metab. Immune Drug Discov.* 2011;5:91–99. [[PubMed](#)] [[Google Scholar](#)]
111. Kang J.E., Lim M.M., Bateman R.J., Lee J.J., Smyth L.P., Cirrito J.R., Fujiki N., Nishino S., Holtzman D.M. Amyloid-beta dynamics are regulated by orexin and the sleep-wake cycle. *Science.* 2009;326:1005–1007. [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)]
112. Struble R.G., Cork L.C., Whitehouse P.J., Price D.L. Cholinergic innervation in neuritic plaques. *Science.* 1982;216:413–415. [[PubMed](#)] [[Google Scholar](#)]
113. Coyle J.T., Price D.L., DeLong M.R. Alzheimer's disease: A disorder of cortical cholinergic innervation. *Science.* 1983;219:1184–1190. [[PubMed](#)] [[Google Scholar](#)]
114. Rasool C.G., Svendsen C.N., Selkoe D.J. Neurofibrillary degeneration of cholinergic and noncholinergic neurons of the basal forebrain in Alzheimer's disease. *Ann. Neurol.* 1986;20:482–488. [[PubMed](#)] [[Google Scholar](#)]
115. Samuel W., Masliah E., Hill L.R., Butters N., Terry R. Hippocampal connectivity and Alzheimer's dementia: Effects of synapse loss and tangle frequency in a two-component model. *Neurology.* 1994;44:2081–2088. [[PubMed](#)] [[Google Scholar](#)]
116. Davis K.L., Mohs R.C., Marin D., Purohit D.P., Perl D.P., Lantz M., Austin G., Haroutunian V. Cholinergic markers in elderly patients with early signs of Alzheimer's disease. *J. Am. Med. Assoc.* 1999;281:1401–1406. [[PubMed](#)] [[Google Scholar](#)]
117. Rinne J.O., Laine M., Hiltunen J., Erkinjuntti T. Semantic decision making in early probable AD: A PET activation study. *Cogn. Brain Res.* 2003;18:89–96. [[PubMed](#)] [[Google Scholar](#)]
118. Terry A.V., Jr, Buccafusco J.J. The cholinergic hypothesis of age and Alzheimer's disease-related cognitive deficits: Recent challenges and their implications for novel drug development. *J. Pharmacol. Exp. Ther.* 2003;306:821–827. [[PubMed](#)] [[Google Scholar](#)]
119. Spencer J.P., Middleton L.J., Davies C.H. Investigation into the efficacy of the acetylcholinesterase inhibitor, donepezil, and novel procognitive agents to induce gamma oscillations in rat hippocampal slices. *Neuropharmacology.* 2010;59:437–443. [[PubMed](#)] [[Google Scholar](#)]
120. Guermonprez L., Ducrocq C., Gaudry-Talarmain Y.M. Inhibition of acetylcholine synthesis and tyrosine nitration induced by peroxynitrite are differentially prevented by antioxidants. *Mol. Pharmacol.* 2001;60:838–846. [[PubMed](#)] [[Google Scholar](#)]
121. Feng Z., Cheng Y., Zhang J.T. Long-term effects of melatonin or 17 beta-estradiol on improving spatial memory performance in cognitively impaired, ovariectomized adult rats. *J. Pineal Res.* 2004;37:198–206. [[PubMed](#)] [[Google Scholar](#)]
122. Tang F., Nag S., Shiu S.Y., Pang S.F. The effects of melatonin and Ginkgo biloba extract on memory loss and choline acetyltransferase activities in the brain of rats infused intracerebroventricularly with beta-amyloid 1–40. *Life Sci.* 2002;71:2625–2631. [[PubMed](#)]

[\[Google Scholar\]](#)

123. Agrawal R., Tyagi E., Shukla R., Nath C. A study of brain insulin receptors, AChE activity and oxidative stress in rat model of ICV STZ induced dementia. *Neuropharmacology*. 2009;56:779–787.

[\[PubMed\]](#) [\[Google Scholar\]](#)

124. Hansen R.A., Gartlehner G., Webb A.P., Morgan L.C., Moore C.G., Jonas D.E. Efficacy and safety of donepezil, galantamine, and rivastigmine for the treatment of Alzheimer's disease: A systematic review and meta-analysis. *Clin. Interv. Aging*. 2008;3:211–225. [\[PMC free article\]](#)

[\[PubMed\]](#) [\[Google Scholar\]](#)

125. Cardinali D.P., Furio A.M., Brusco L.I. Clinical aspects of melatonin intervention in Alzheimer's disease progression. *Curr. Neuropharmacol.* 2010;8:218–227. [\[PMC free article\]](#) [\[PubMed\]](#)

[\[Google Scholar\]](#)

126. Fernandez-Bachiller M.I., Perez C., Campillo N.E., Paez J.A., Gonzalez-Munoz G.C., Usan P., Garcia-Palomero E., Lopez M.G., Villarroya M., Garcia A.G., et al. Tacrine-melatonin hybrids as multifunctional agents for Alzheimer's disease, with cholinergic, antioxidant, and neuroprotective properties. *ChemMedChem*. 2009;4:828–841. [\[PubMed\]](#) [\[Google Scholar\]](#)

127. Spuch C., Antequera D., Isabel Fernandez-Bachiller M., Isabel Rodriguez-Franco M., Carro E. A new tacrine-melatonin hybrid reduces amyloid burden and behavioral deficits in a mouse model of Alzheimer's disease. *Neurotox. Res.* 2010;17:421–431. [\[PubMed\]](#) [\[Google Scholar\]](#)

128. Arends Y.M., Duyckaerts C., Rozemuller J.M., Eikelenboom P., Hauw J.J. Microglia, amyloid and dementia in Alzheimer's disease. A correlative study. *Neurobiol. Aging*. 2000;21:39–47. [\[PubMed\]](#) [\[Google Scholar\]](#)

129. Combadiere C., Feumi C., Raoul W., Keller N., Rodero M., Pezard A., Lavalette S., Houssier M., Jonet L., Picard E., et al. CX3CR1-dependent subretinal microglia cell accumulation is associated with cardinal features of age-related macular degeneration. *J. Clin. Invest.* 2007;117:2920–2928.

[\[PMC free article\]](#) [\[PubMed\]](#) [\[Google Scholar\]](#)

130. Streit W.J., Mrak R.E., Griffin W.S. Microglia and neuroinflammation: A pathological perspective. *J. Neuroinflamm.* 2004;1 doi: 10.1186/1742-2094-1-14. [\[PMC free article\]](#) [\[PubMed\]](#) [\[CrossRef\]](#) [\[Google Scholar\]](#)

131. Hardy J.A., Higgins G.A. Alzheimer's disease: The amyloid cascade hypothesis. *Science*. 1992;256:184–185. [\[PubMed\]](#) [\[Google Scholar\]](#)

132. Stuchbury G., Munch G. Alzheimer's associated inflammation, potential drug targets and future therapies. *J. Neural. Transm.* 2005;112:429–453. [\[PubMed\]](#) [\[Google Scholar\]](#)

133. Tuppo E.E., Arias H.R. The role of inflammation in Alzheimer's disease. *Int. J. Biochem. Cell B.* 2005;37:289–305. [\[PubMed\]](#) [\[Google Scholar\]](#)

134. Park S.Y., Jin M.L., Kim Y.H., Kim Y., Lee S.J. Anti-inflammatory effects of aromatic-turmerone through blocking of NF-kappaB, JNK, and p38 MAPK signaling pathways in amyloid beta-stimulated microglia. *Int. Immunopharmacol.* 2012;14:13–20. [\[PubMed\]](#) [\[Google Scholar\]](#)

135. Chung S.Y., Han S.H. Melatonin attenuates kainic acid-induced hippocampal neurodegeneration and oxidative stress through microglial inhibition. *J. Pineal Res.* 2003;34:95–102. [\[PubMed\]](#) [\[Google Scholar\]](#)

136. Mohan N., Sadeghi K., Reiter R.J., Meltz M.L. The neurohormone melatonin inhibits cytokine, mitogen and ionizing radiation induced NF-kappa B. *Biochem. Mol. Biol. Int.* 1995;37:1063–1070. [\[PubMed\]](#) [\[Google Scholar\]](#)

137. Chuang J.I., Mohan N., Meltz M.L., Reiter R.J. Effect of melatonin on NF-kappa-B DNA-binding activity in the rat spleen. *Cell Biol. Int.* 1996;20:687–692. [PubMed] [Google Scholar]
138. Lau W.W., Ng J.K., Lee M.M., Chan A.S., Wong Y.H. Interleukin-6 autocrine signaling mediates melatonin MT(1/2) receptor-induced STAT3 Tyr(705) phosphorylation. *J. Pineal Res.* 2012;52:477–489. [PubMed] [Google Scholar]
139. Shen Y., Zhang G., Liu L., Xu S. Suppressive effects of melatonin on amyloid-beta-induced glial activation in rat hippocampus. *Arch. Med. Res.* 2007;38:284–290. [PubMed] [Google Scholar]
140. Grases F., Costa-Bauza A., Prieto R.M. A potential role for crystallization inhibitors in treatment of Alzheimer's disease. *Med. Hypoth.* 2010;74:118–119. [PubMed] [Google Scholar]
141. Wollen K.A. Alzheimer's disease: The pros and cons of pharmaceutical, nutritional, botanical, and stimulatory therapies, with a discussion of treatment strategies from the perspective of patients and practitioners. *J. Clin. Ther.* 2010;15:223–244. [PubMed] [Google Scholar]
142. Dowling G.A., Burr R.L., van Someren E.J., Hubbard E.M., Luxenberg J.S., Mastick J., Cooper B.A. Melatonin and bright-light treatment for rest-activity disruption in institutionalized patients with Alzheimer's disease. *J. Am. Geriatr. Soc.* 2008;56:239–246. [PMC free article] [PubMed] [Google Scholar]
143. Pappolla M.A., Chyan Y.J., Poeggeler B., Frangione B., Wilson G., Ghiso J., Reiter R.J. An assessment of the antioxidant and the antiamyloidogenic properties of melatonin: Implications for Alzheimer's disease. *J. Neural. Transm.* 2000;107:203–231. [PubMed] [Google Scholar]
144. Nagtegaal J., Smits M., Van Der Meer Y., Fischer-Steenvoorden M. Melatonin: A survey of suspected adverse drug reactions. *Sleep-Wake Res. Netherl.* 1996;7:115–118. [Google Scholar]
145. Avery D., Lenz M., Landis C. Guidelines for prescribing melatonin. *Ann. Med.* 1998;30:122–130. [PubMed] [Google Scholar]

---

Articles from International Journal of Molecular Sciences are provided here courtesy of **Multidisciplinary Digital Publishing Institute (MDPI)**