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### Acetyl-L-Carnitine in Parkinson's Disease

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#### 1. Introduction

Parkinson's disease (PD) is the most common neurodegenerative movement disorder that is estimated to affect approximately 1% of the population older than 65 years of age (deRijk et al., 2006; Saunders, 2000). PD was first described in 1818 by the British physician J. Parkinson. Before that date, no one had ever described the symptoms of this disease; so many researchers theorize that this pathology is the product of the English Industrial Revolution (Parris, 2000; Perlmutter, 2000). Some authors speculate that new neurotoxic contaminants produced by the industries can have been the cause of this chronic and progressive disease. PD is characterized by the progressive depletion of pigmented dopamine-containing neurons in the region known as the substantia nigra pars compacta and by the presence of intraneuronal aggregates called Lewy bodies (LBs), which are enriched in filamentous  $\alpha$ -synuclein and other proteins that are often ubiquinated (Lee & Trojanowsky, 2006). Approximately 80% of dopaminergic neurons in the substantia nigra are already irreversibly destroyed when the symptoms of PD becomes significantly visible. Depletion of dopamine causes dysregulation of the motor circuits that project throughout the basal ganglia (BG), resulting in the cardinal clinical manifestations of PD: bradykinesia (extreme slowness), tremor, rigidity, and postural instability. Consequently, patients experience increasing difficulty in daily living functions along the course of the disease. Additional neuronal fields and neurotransmitter systems are also involved in PD, including the locus coeruleus, the dorsal motor nucleus, the autonomic nervous system and the cerebral cortex. Consequently, noradrenergic, serotoninergic, and cholinergic neurons are also lost. These widespread neuronal changes led to complex and variable progressive nonmotor symptoms such as cognitive decline, sleep abnormalities, and depression which dominate the later stages of PD (Braak, 2003). In any case, PD is primarily a sporadic disorder and its etiopathogenesis is still not fully understood, but the recent discovery of genes associated with rare monogenic forms of the disease, together with earlier studies and new experimental animal models, has provided important and novel insight into the molecular pathways involved in disease pathogenesis (Wood-Kaczamar, 2006). Increasing evidence indicates that deficits in mitochondrial function, oxidative and nitrosative stress, accumulation of aberrant or misfolded proteins, and ubiquitin-proteasome system dysfunction can represent the principal molecular pathways or events that commonly underlie the pathogenesis of sporadic and familial forms of PD (Schapira, 2008). However it is possible that multiple factors contribute to the cascade of events leading to cells death in patients with PD and that different factors might be more important in different individuals (Olanow, 2009) (Fig. 1).

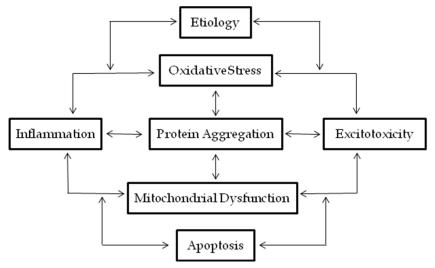


Fig. 1. Schematic illustration of factors that might be involved in the pathogenesis of cells death in PD. (Adapted from Olanow, 2007).

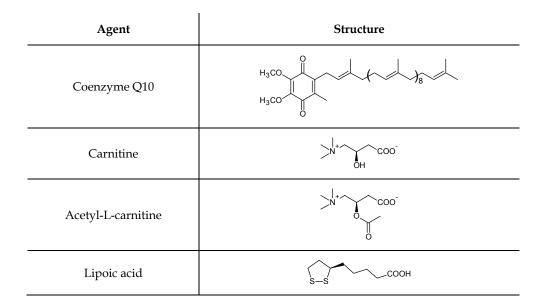
Several pharmacological agents are currently available for the management of PD (Table 1). These drugs can provide symptomatic relief but no agents capable to halt the progression of the neurodegenerative process or reverse the neuronal degeneration have been developed yet. Moreover, the neuroprotective effects suggested for many of the approved drugs have not been convincingly demonstrated in PD patients. Furthermore, although PD also involves degeneration of non-dopaminergic neurons, the treatment of the resulting predominantly non-motor features remains a challenge. The leading therapeutic strategy pursued in PD management is the so called dopamine replacement therapy (DRT), which employs drugs acting on dopamine (DA) circuits to restore the deficient dopaminergic tone existing in this pathology. Pharmacological agents have also been developed which can indirectly boost DA transmission, based on the functional interactions existing between DA and other neurotransmitters in the BG. L-Dopa is the key compound in the treatment of PD, acting as a precursor of DA. It has a long clinical record as the most effective antiparkinsonian drug, and it is still considered the "gold standard" in pharmacological treatment of PD (Mercuri & Bernardi, 2005). However, besides offering only symptomatic relief for patients, motor and non-motor untoward effects are often observed in the course of L-dopa therapy, which can be severe and limit its therapeutic potential (Encarnation & Hauser, 2008; Fox & Lang, 2008). Furthermore, exposure of patients to L-dopa, results in fluctuations of motor responses in approximately 30-50% of patients exposed to therapy for as little as 5 or more years. The most common fluctuation experienced is the so-called "on-

Drug	Mechanism of action			
HO HO L-dop	NH <sub>2</sub>	Precursor of DA		
$\begin{array}{c c} & & & & & & \\ & & & & & & \\ & & & & & $	Agonist at D2-like receptors			
Ropinirole Pramipexc	$S_{N} \rightarrow NH_{2}$ $O$			
Selegiline	Rasagiline	MAO-B inhibitors		
HO + CN + CN + O + O + O + O + O + O + O + O + O +	HO HO NO <sub>2</sub> Tolcapone	COMT inhibitors		
Amantad		NMDA glutamate receptor antagonist		
он Biperiden Triexypheni	v dyl Metixene	Muscarinic receptor antagonists		

Table 1. Currently available pharmacological therapies for PD treatment.

off" phenomenon that results in an unpredictable transient loss of therapeutic effect. Apart from L-dopa, drugs that are currently prescribed for the management of PD include DA receptor agonists, selective monoaminooxidase type B inhibitors (MAO-B), catecol-Omethyltransferase (COMT) inhibitors, amantadine (an antiviral agent that also bears action as an antiglutamatergic agent), and anticholinergics. DA receptor agonists counteract PDassociated motor impairment chiefly stimulating the D<sub>2</sub>-like receptors, though some of them can also bind non-dopaminergic receptors. They may be used alone to delay the need for Ldopa or as multiple-medication therapy (MMT) with L-dopa to increase its effectiveness (Cavalli et al. 2008). Neuroprotective properties have been suggested for some dopaminergic agonists (e.g. bromocriptine and pramipexole), although the clinical evidence collected so far does not convincingly support this hypothesis (Schapira, 2003). Certain other available drugs, like MAO-B (Fernandez & Chen, 2007) and COMT inhibitors (Canesi et al., 2008; Schrag, 2005), are used mainly as MMT with L-dopa, since they alter the in vivo metabolism of DA by increasing its plasma half-life. Functional interactions between glutamate and DA receptors exist in BG, and evidence suggests that the loss of DA in PD may lead to glutamatergic hyperactivity, which participates in the manifestation of motor impairment accompanying the disease (Chase & Oh, 2003). On this basis, glutamatergic antagonists have been extensively investigated as potential antiparkinsonian drugs (Johnson et al. 2009). Among these, amantadine is the best characterized antiglutamatergic agent used in PD management. In addition to the blockade of ionotropic N-methyl-D-aspartate (NMDA) receptors, amantadine posses other mechanisms of action which contribute to its effects: anticholinergic activity, stimulation of DA release, modulation of the affinity of postsynaptic DAergic receptors for DA (Metman et al., 1998, Peeters et al., 2003). Anticholinergic compounds were the first, and for a long time the only, pharmacological agents available to treat motor deficits accompanying PD (Brocks, 1999). They were intend to correct the imbalance between DA and acetylcholine levels that take place in the BG, where a reduction of cholinergic tone may amplify DA-mediated signal (Cragg, 2006). Although these drugs produce some beneficial effects on PD symptoms, they are associated with adverse cognitive effects (Cancelli et al., 2009). All the anticholinergics used against PD bind to the central muscarinic receptors, having no affinity for the nicotinic ones, although they also block peripheral muscarinic receptors, and this triggers many of their adverse effects, which include nausea, constipation and urinary retention (Lees, 2005). Nevertheless, currently available pharmacological therapies are unable to arrest or to reverse the progression of this relentlessly progressive and severely debilitating condition. PD is currently an incurable disease, and the number of subjects afflicted with this disease is constantly increasing due to the increasing global geriatric population. Therefore, the need for newer and more effective agents is receiving a great deal of attention and, consequently, being subjected to extensive research. The vast amount of information gained regarding the pathogenesis of PD has fuelled numerous developments and vast range of investigated agents have demonstrated immense potential for preventing and eventually providing cure for this condition. Clinical and biochemical evidences suggest that PD involves multifactorial, oxidative neurodegeneration and that L-dopa therapy aggravates the oxidative burden. Strong evidence now exists to support an aberrant role for mitochondrial functions, as well as increased oxidative stress, in the pathogenesis of PD. If mitochondrial defects and oxidative damage play a role in the pathogenesis of PD, then one would suspect that

agents that may improve mitochondrial function or exert antioxidative effects could be neuroprotective. There are several agents that are currently under investigation for their potential neuroprotective effects based on their capacity to modify mitochondrial dysfunction. These include creatine, coenzyme Q10, nicotinamide, lipoic acid and acetyl-L-carnitine, etc. (Table 2). These agents are therefore promising candidates for neuroprotective drugs against PD (Beal, 2003). Acetyl-L-carnitine, (3R)-3-(acetyloxy)-4-(trimethylammonio)butanoate (table 2), is an ester of the trimethylated amino acid, Lcarnitine, and is synthesized in the brain, liver and kidney by the enzyme acetyl-Lcarnitine transferase. Acetyl-L-carnitine facilitates the transport of fatty acids and other moieties across the membranes of mitochondria, thereby participating in the production of energy and mitochondrial function within the brain. Acetyl-L-carnitine has been proposed to have beneficial effects in preventing the loss of brain functions which typically occur during aging and neurodegenerative disorders. The main mechanism of action of acetyl-L-carnitine is the improvement of mitochondrial respiration which allows the neuron to produce ATP necessary to maintain the normal membrane potential. However, acetyl-L-carnitine has been shown to be neuroprotective through a variety of other effects such as the increase in protein kinase C (PKC) activity (McDaniel, et al. 2003). Moreover acetyl-L-carnitine has also been reported to attenuate the occurrence of parkinsonian symptoms associated with 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP) in vivo, and protects in vitro against the toxicity of neurotoxic 1-methyl-4phenylpyridinium (MPP<sup>+</sup>), a metabolite of MPTP (Hongyu et. al., 2010). Therefore, acetyl-L-carnitine with its well known antioxidant energizing protective activities and with its trophic effects, might be an effective and safe prevention strategy for PD.



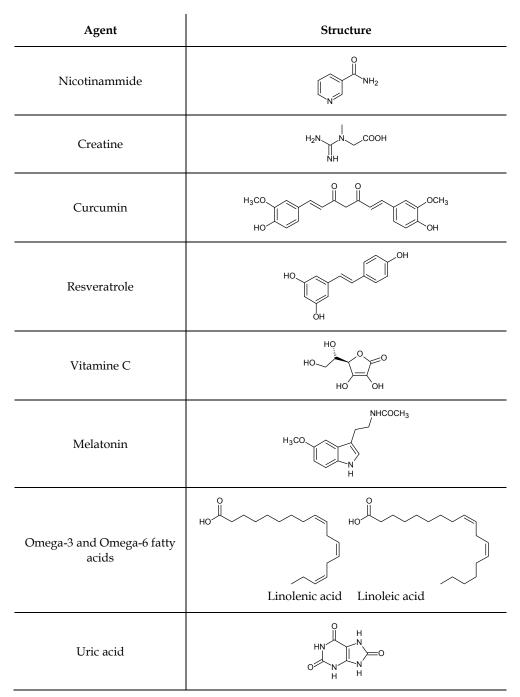


Table 2. Neuroprotective agents effective in PD models.

#### 2. Nutritional and biochemical aspects of acetyl-L-carnitine

Carnitine has been discovered in the bovine muscle in 1905, but its structure was defined only in 1927. In 1958 the role of carnitine has been discovered by I. Frizz, who demonstrated that this substance is important in stimulating the oxidation of long chain fatty acids into the mitochondria. L-Carnitine (-)-3-hydroxy-4-(trimethylammonio)butanoate is a highly polar, water-soluble quaternary amine that exists as a zwitterion under physiological conditions. It was initially called vitamin T, because it is necessary for the growth of the Tenebrio Molitor warm. Although it is structurally similar to an amino acid it is not involved in the formation of proteins, and it is more similar to acetylcholine. Carnitine is synthesized in vivo from the amino acids lysine and methionine, especially in liver, kidney, and muscle, and it is stored mainly in skeletal and cardiac muscles (Marquis & Fritz, 1965). Exogenous carnitine, taken predominantly with the meat of the diet, is about 75% of the body carnitines, while the daily requirement is about 200-300 mg. In vivo synthesis of carnitine, supplemented by carnitine from diet, provides sufficient carnitine to maintain metabolic functions. However, in cases of excessive loss of carnitine (low carnitine intake with the diet, altered carnitine metabolism or disease states such as in neurodegenerative diseases and geriatric depression), supplementation with acetyl-L-carnitine may be beneficial. Tissue levels of L-carnitine in animals and humans decrease with age, due to reduced integrity of the mitochondrial membranes. Acetyl-L-carnitine, an ester of the Lcarnitine, is synthesized in human brain, liver, and kidney by the enzyme acetyl-Lcarnitine transferase. Carnitine, acetyl-L-carnitine and acyl-L-carnitine are responsible for many biological actions. Several authors have suggested that acetyl-L-carnitine has beneficial effects on brain functions during aging and in conditions of neurodegenerative diseases such as Parkinson's and Alzheimer's diseases. It has been demonstrated that acetyl-L-carnitine plays a role in increasing the potency of cholinergic and anti-cholinergic actions, by reacting with the electrophilic or cationic site of the cholinergic receptor (Sinicropi et al., 2010). Carnitine as acyl-L-carnitine is important in the mitochondrial process of  $\beta$ -oxidation of fatty acids (Bremer, 1962; Bremer et al., 1983) and the acetyl moiety can be utilized to maintain acetyl-CoA levels. Acetyl-L-carnitine promotes acetylcholine production and release, and stimulates membrane phospholipid synthesis (Pettegrew et al., 2000). In addition, the acetyl moiety of acetyl-L-carnitine can acetylate -NH<sub>2</sub> and/or -OH functional groups of lysine, serine, threonine, tyrosine and N-terminal amino acids in proteins modifying their structure, function and activity. Moreover acetyl-L-carnitine modulates glucose metabolism and stimulates glycogen synthesis, restores ammonia induced depletion of brain energy stores in sparse-fur mice with elevated ammonia and glutamine levels (Rao et al., 1997) and, with carnitine, maintains progressive spermatozoa motility (Jeulin et al., 1988). This molecule acts also on the mitochondrial redox reactions that allow neurons to produce ATP, required to maintain normal membrane potential (McDaniel et al., 2003). L-carnitine and acetyl-L-carnitine are administrated orally, intravenously and/or intramuscularly. These compounds are absorbed in the jejunum by simple diffusion. Transport into tissues and cells is via an active transport mechanism and acetyl-L-carnitine and carnitine plasma concentrations reach equilibrium via carnitine acetyl-transferase activity. Both intravenous and oral administrations result in a corresponding increase of cerebrospinal fluid (CSF) concentrations of acetyl-L-carnitine, indicating that it readily crosses the blood-brain barrier (Kido et al., 2001; Thal et al., 1996).

#### 3. MPTP and neuroprotective effect of acetyl-L-carnitine in pathogenesis of Parkinson's disease

In the mitochondria of all cells redox reactions produce free radicals. High levels of reactive oxygen species (ROS) and reactive nitrogen species (RNS), especially in Parkinson's disease, can result in damage to phospholipids and polyunsaturated fatty acids, which are both abundant in the brain and therefore susceptible to oxidative damage. Therefore, there are many evidences for increased oxidative damage also to DNA and proteins (Dexter et al., 1994). Many evidences have accumulated implicating mitochondrial defects and oxidative stress damage in the pathogenesis of Parkinson's disease. In this contest, fundamental role is attributed to 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP), a neurotoxin that is able to produce an experimental model of Parkinson's disease (PD) in humans and laboratory animals (such as primates and mice). It replicates most of the clinical features of PD as well as the main biochemical and pathologic hallmarks of the disease. The apparent neurotoxic specificity of MPTP is mediated through its conversion into 1-methyl-4-phenylpyridine (MPP+) (Fig. 2) by the action of the mitochondrial enzyme MAO B (Javitch & Snyder, 1984; Javitch et al., 1985).

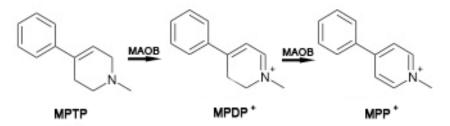


Fig. 2. Conversion of MPTP in MPP+

The neurotoxicity of MPTP was discovered in 1976 when B. Kidstone, a 23 years old student of chemistry in Maryland, synthesized MPTP and injected it himself. He was contaminated by the MPTP and three days later he showed all the symptoms of Parkinson's disease. Studies on MPTP toxicity showed that it is mediated by inhibition of respiratory chain complex I activity (Bloem et al., 1990). There are at least three ways that MPP+ can follow once inside dopaminergic neurons (Przedborski & Vila, 2003). It can: a) take the vesicular pathway, bind to dopamine transporters to be translocated into synaptosomal vesicles (Liu et al., 1992); b) interact with various cytosolic enzymes by remaining into the cytosol (Klaidman et al., 1993); c) be concentrated in the mitochondria (Ramsay & Singer, 1986). MPP+ can passively enter the mitochondria through the transmembrane potential of the mitochondrial membranes and it can accumulate into the mitochondrial matrix. First of all, MPP+, after being entered the mitochondrial matrix, inhibits the Krebs cycle enzyme αketoglutarate dehydrogenase (Mizuno et al., 1987), but the main cause of mitochondrial dysfunction involves the respiratory chain complex I (Ramsay et al., 1991). The MPP+ toxicity is associated with oxidative damage. In fact MPP+ induces superoxide production and increases lipid peroxidation. Important studies of Dexter and colleagues, showing increases in both malondialdehyde and in cholesterol hydroperoxides, led to a direct evidence of oxidative damage in Parkinson's disease (Dexter et al., 1994). Even the concentration of 8-hydroxy-2'-deoxyguanosine has been found three to four times higher in the caudate and substantia nigra of Parkinson's disease subjects (Sancher-Ramos et al., 1994). Shergill and co-authors have found a significant increase of nitrosyl complexes in Parkinson's disease substantia nigra (Shergill, 1996). However, recent studies suggest that MPP<sup>+</sup> toxicity, at least in the initial stages, is primarily due to a decrease in mitochondrial ATP synthesis rather than the formation of free radicals (Fonck & Baudry, 2003). Subsequently, the protective effect of acetyl-L-carnitine in Parkinson's disease, induced by MPTP, has been studied in a group of primates by the research group of Bodis-Wolner (Bodis-Wolner et al., 1991). For their studies Bodis-Wolner and colleagues used three groups of primates, the first of which was treated just with MPTP. To the second group was administered acetyl-L-carnitine before the MPTP, while the third group had a control role. Their results have shown that primates treated only with MPTP developed the classic symptoms of parkinsonism. In the second group, only in a primate a weak form of Parkinson's disease has evolved to signify the protective effect of acetyl-L-carnitine on the disease development. It is conceivable that the effect of MPP+ results in altering the mitochondrial respiration processes linked to NAD (Nicklas et al., 1985, 1987; Heikkila et al, 1985; Ramsey et al., 1986; Mizuno et al., 1988).

#### 4. Toxic and antioxidant compounds in Parkinson's disease

Oxidants, as hydrogen peroxide and superoxide radicals, are produced as by-products of oxidative phosphorylation into mitochondria, making these organelles the main site of ROS generation within the cells. In fact, mitochondria are a major source of ROS, with up to 2-3 % of all oxygen consumed by mitochondria being converted to hydrogen peroxide (Boveris et al., 1972). This is the normal condition and basal levels of ROS can be limited by the presence of efficient antioxidant defence systems, including the enzymatic antioxidants (superoxide dismutase, catalase, peroxidases, and heme oxygenase) and the non-enzymatic redox-regulating antioxidants (glutathione and vitamin C). However, in pathological neurodegenerative conditions, like in Parkinson's disease, where mitochondrial respiratory defects occur, the amount of ROS produced by the electron transport chain dramatically increases, abolishing the antioxidant protection systems (Parker et al., 1989). In studies on brain tissue in patients with PD, the activity of complex I is reduced in the substantia nigra, without any decrease in other brain regions. As shown by Haas et al. (Hass et al., 1995), the activity of complex I is reduced also in PD platelets of un-medicated patients. A decrease of coenzyme Q10 levels in platelets mitochondria, which is correlated with reduced complex I of respiratory chain, is reported in Shults et al. (Shults et al., 1997). In parkinsonian subjects platelets mitochondria were found to have lower levels of coenzyme Q10 than mitochondria from age/sex-matched controls. As shown in certain clinical studies, coenzyme Q10 appears to slow the progressive deterioration of function in PD (Shults et al., 2002). Coenzyme Q10 is necessary for the normal activity of the respiratory chain and transfers electrons from complexes I and II to III. It can be worthwhile to use coenzyme Q10 to restore the functions of the respiratory chain and scavenge ROS. Nevertheless, coenzyme Q10 protects primary dopaminergic neurons in vitro against cell death induced by MPTP (Gille et al., 2004), and seems that at least partially restores the function of complex I in the tissues of patients with PD (Shults et al., 1998; Storch et al., 2007). Moreover, it has antioxidant properties; it has been also shown to prevent peroxidation of membrane lipids and protect mitochondrial DNA from free oxygen radicals. Important results on the defects of complex I activity in the pathogenesis of PD are derived from studies with the toxin rotenone. Rotenone (Table 3) is a

natural compound extracted from the roots of certain plants and has been used as an insecticide for vegetables. Rotenone rapidly crosses the blood-brain barrier due to its lipophilic structure, and rapidly crosses the biological membranes of mitochondria into the cells, in which the toxin reduces the activity of oxidative phosphorylation, by binding to PSST subunit of respiratory chain complex I (Schuler & Casida, 2001). It is known that rotenone is a highly specific inhibitor of complex I of the electrons transport chain. The possibility that rotenone and other pesticides are involved in the pathogenesis of PD stems from epidemiological studies (Gorell et al., 1998; Seidler et al., 1996). In fact, an atypical Parkinson's syndrome has developed in populations of the French West Indies, taking fruit and herbal tea containing insecticides (Caparros-Lefebvre & Elbaz, 1999). Pesticides and herbicides become highly suspect as potential PD triggers. A connection was long suspected between PD and rural living, including the drinking of contaminated well water or exposure to pesticides and herbicides (Hancock et al., 2008; Stephenson, 2000). Recently it was observed that high blood levels of homocysteine (Table 3) are present in patients with PD receiving L-dopa.

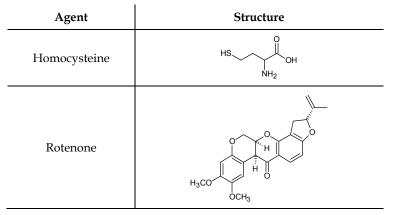


Table 3. Neurotoxic agents in Parkinsons's disease

On the other hand it is known that the increase in homocysteine is a risk factor for atherosclerosis, stroke, vascular disease, and dementia. There are many proposed mechanisms for toxicity of homocysteine in promoting neurodegenerative diseases such as of Parkinson's and Alzheimer's diseases (Postuma & Lang, 2004; Seshadri et al., 2002): free radicals formation, induction of inflammation, and altered vulnerability of complex I mitochondrial respiratory chain. The formation of homocysteine occurs from methionine which is converted to S-adenosylmethionine and then demethylated to Sadenosylhomocysteine, which forms homocysteine. Homocysteine itself is reused to form methionine by the action of two enzymes rate-limiting methylenetetrahydrofolate reductase (MTHFR) and betaine homocysteine methyltransferase (BHMT). Homocysteine can be metabolized to cysteine due to the cystathionine-betasintethase (CBS). The MTHFR requires cofactors such as vitamin B12 and folate, while the CBS requires vitamin B6. The administration of L-dopa urges COMT activities causing methylation of L-dopa to 3-Omethyl-dopa and, at the same time, the demethylation of S-adenosylmethionine to Sadenosylhomocysteine, which rapidly forms homocysteine. Therefore, since the

demethylation of S-adenosylmethionine results in an increase of homocysteine, it is easy to understand why Parkinson's patients treated with L-dopa have higher levels of homocysteine. Consequently, any substance that can reduce blood levels of homocysteine should be administered to Parkinson's patients who require L-dopa: COMT inhibitors, vitamin B6, folate and vitamin B12. The toxicity of these compounds can be prevented with the administration of antioxidants. Complex I of respiratory chain is genetically coded for the ring-shaped mitochondrial DNA (mtDNA). A line of evidence, implicating mitochondria and mitochondrial genome (mtDNA) in PD pathogenesis, comes from "cybrid cells". While many proteins and enzymes of all electron transport chain complexes are coded from nuclear genes, 13 of them are coded in the small circular double-stranded mtDNA, located within the mitochondrial matrix. The human mitochondrial genome contains 37 genes (16,560 base pairs), including 13 that encode subunits of proteins of respiratory chain, and in particular 7 subunits of complex I. Mitochondrial genes exhibit a much higher mutation rate compared to nuclear genes and mtDNA is exposed to ROS generated during respiration. It is believed, therefore, that the oxidative damage to mitochondrial DNA and its mutation can play a role for mitochondrial dysfunction in PD. To determine if complex I is genetically abnormal, Swerdlow and colleagues (Swerdlow et al., 1996) devised an experiment with cybrid cells, generated to uncouple potential effects of a damaged mtDNA from effects of the nuclear DNA. These Swerdlow cybrid cells are hybrid cells which combine the nuclear genome from neuroblastoma cells with the mitochondrial genome from platelets of PD patients or healthy control subjects. Using these cybrid cells, Swerdlow and colleagues confirmed that PD mitochondria were less efficient in complex I activity (- 20%) associated with increased free radical production and apoptotic cell death (Gu et al., 1998; Swerdlow et al., 1996). Since only mtDNA is derived from the donor platelets, the Swerdlow experiment can be interpreted as a suggestion for mtDNA transmission of the mitochondrial defect. Besides, many authors suggest that alterations in processes of ubiquitination and degradation of proteins by the 26S proteasome can play a primary role in the PD pathogenesis (McNaught & Ienner, 2001; McNaught et al., 2001). Products of oxidative damage can contribute to substantia nigra degeneration in PD. The oxidized proteins can not be adequately ubiquitinated and recognized by the proteasomes and accumulated within the cells. The accumulation of ubiquitinated proteins and the loss of proteasomal activity can induce mitochondrial dependent apoptotic death of dopaminergic neurons in a manner similar to that occurring in the substantia nigra in PD. Although rare, some genetic cases of Parkinson's disease are linked to mutations in a synaptic protein called  $\alpha$ -synuclein that was originally identified from smaller peptides isolated in amyloid-containing fractions of Alzheimer disease brains (Hong, 2005). The  $\alpha$ synuclein protein is another aggregating, fibril-forming protein that is a major component of the Lewy body lesions characteristic of PD as well as certain cases of Alzheimer (AD) and several other neurodegenerative conditions. a-Synuclein aggregates show evidence of nitration-based oxidative damage that might play a critical role in aggregate formation (Giasson et al., 2000). Recent studies have shown that the polyphenol curcumin (the active principle of turmeric Curcuma longa) can reduce the aggregation of  $\alpha$ -synuclein, and its administration to cultured cells with  $\alpha$ -synuclein aggregate formation results in fewer aggregates (Ono & Yamada, 2006; Pandey & Galvin, 2005). Also the pesticide rotenone leads to the presence of Lewy bodies with aggregation of  $\alpha$ -synuclein (Sherer et al., 2003). The evidence that rotenone, a inhibitor of mitochondrial respiratory chain complex I, causes

aggregation of Lewy bodies may means that mitochondrial dysfunction has a role in the development of these pathologic fibril-forming proteins in Parkinson's disease (Dawson & Dawson, 2003a). Rajeswary has shown that curcumin protects mouse brain from MPTPinduced neurotoxicity by virtue of its scavenger activity (Rajeswary, 2006). Moreover, curcumin has been shown to protect PC12 cells from MPP+ by inducing bcl-2, a antiapoptotic protein, preventing the dissipation of membrane potential of mitochondria and reducing then ROS and iNOS levels (Chen et al., 2006). The importance of mitochondria in the neuroprotective effect of curcumin has been also emphasized by Mythri et al. (2007), who demonstrated that curcumin inhibits the formation of peroxynitrite responsible for the damage of respiratory chain complex I. Studies with humans and rodents have demonstrated that after oral administration curcumin is transformed to curcumin glucuronide and curcumin sulphate, not only in the liver (Rahaman et al., 2006) but also in the intestinal tract (Ireson et al., 2002). In these organs, curcumin is also reduced into (DHC), tetrahydrocurcumin hexahydrocurcumin, dihydrocurcumin (THC), octahydrocurcumin and hexahydrocurcuminol (Ireson et al., 2002; Rahaman et al., 2006); curcumin, DHC and THC can be further converted in glucuronide conjugates. It is important to note that curcumin and THC have anti-inflammatory activity; in humans and rodents curcumin inhibits the activity of cytochrome P450 enzymes, glutathione-transferase and UDP-glucuronosyl transferases (Basu et al., 2004; Hayeshi et al., 2007; Thapliyal & Maru, 2001). Moreover, it has been shown that a large number of polyphenolic antioxidants have a protective effect against the degeneration induced by high levels of ROS and RNS in cases of mitochondrial dysfunction. It has been proven that green tea polyphenols have a protective effect against 6-hydroxydopamine toxicity in SH-SY5Y cells (Guo et al., 2005). 6-Hydroxydopamine is a hydroxylated analogue of dopamine, extensively used in rodents. 6-Hydroxydopamine possess a high affinity for many membrane transporters of catecholamines and norepinephrine, allowing the drugs to freely enter both dopaminergic and noradrenergic neurons (Bovè et al., 2005). The efficacy of the green tea component epigallocatechin 3-gallate has been demonstrated in the MPTP mouse model of Parkinson's disease. It has been shown that in these treated rats there is both loss of dopaminergic neurons and attenuation of striatal dopamine levels (Choi et al., 2002). Choi et al. (2002) suggest that this protection is mediated by inhibition of NOS expression. Epidemiological evidence shows that two caps a day of green tea have a protective effect against the Parkinson's disease development (Chan et al., 1998). Another polyphenol used in the fight against Parkinson's disease is the oxyresveratrol, found in large amounts in mulberry wood, which has shown potent scavenger activity against ROS and RNS in glial cells exposed to hydrogen peroxide (Lorenz et al., 2003). In addition, in a study on 6-hydroxydopaminetreated neuroblastoma SH-SY5Y cells has been found that the oxyresveratrol drastically reduces the production of ROS and reduces also the apoptotic activity of caspase-3 caused by damaged mitochondria (Chao et al., 2008). Other important antioxidant is uric acid (Ames et al., 1981). Recent studies and epidemiological researches have shown a correlation between high levels of uric acid in serum and a lower incidence of Parkinson's disease (Annanmaki et al., 2007, Weisskopf et al., 2007; Winquist et al., 2010). It was also seen that the uric acid protects against the damage caused by free radicals on the mtDNA (Anderson & Harris, 2003), helping to maintain the integrity of the mitochondrial genome and prevent possible mutations. In addition, uric acid prevents the death of dopaminergic cells treated with rotenone and homocysteine; treatments that increase the production of ROS and act on mitochondrial membrane depolarization (Duan et al., 2002). Most likely uric acid neutralizes ROS through the Fenton reaction, thus providing dopaminergic neuroprotection. But we have to balance the benefits of dietary supplementation of uric acid on parkinsonism and the possible risk of developing gout and cardiovascular problems.

# 5. Acetyl-L-carnitine and other nutrients in age-dependent neurodegenerative diseases

A broad spectrum of both genetic and environmental factors has been suggested as contributing to the initiation and progression of PD. Among these, an important risk factor for the disease is the aging (Parris, 2000). It contributes to PD progression, perhaps because of accumulative oxidative damage and decrease of antioxidant capacity. Many evidences support the validity of the oxidative stress hypothesis, which suggests that lowered functional capacity in aged organisms is the result of an increased generation of reactive species. The increased levels of ROS and RNS can cause damage to intracellular macromolecules, as DNA, proteins and lipids and consequently impairing the function of vulnerable tissues and leading to the accumulation of altered gene products (Calabrese et al., 2006a). In addition, protein, lipid or glucose oxidation disrupts redox homeostasis and leads to accumulation of unfolded or misfolded proteins in the aging brain. For this reason Parkinson's and Alzheimer's diseases, having a common denominator, production of abnormal proteins, mitochondrial dysfunction and oxidative stress, are called "protein conformational diseases" (Calabrese et al., 2008). In particular, an unfolded protein response conformational disease is condition that arise from dysfunctional aggregation of proteins in non-native conformations. This is often associated with multiple metabolic derangements that result in the excessive production of ROS and oxidative stress (Zhang et al., 2006). Genetic studies have also revealed that aging can be controlled by changes in intracellular NAD/NADH ratio regulating sirtuins, a group of proteins linked to aging, metabolism and stress tolerance in several organisms. Consistently, the neuroprotective roles of dietary antioxidants including for example, curcumin, carnosine, resveratrol and acetyl-L-carnitine have been demonstrated through the activation of these redox-sensitive intracellular pathways. In particular, acetyl-L-carnitine has been proposed to have beneficial effects in preventing the loss of brain function which typically occurs during aging and neurodegenerative disorders. In fact, acetyl-L-carnitine treatment has been shown to prevent age-related changes in mitochondrial respiration and decrease oxidative stress biomarkers thorough the up-regulation of HO-1 (heme oxygenase-1), Hsp70 (heat shock protein 70) and superoxide dismutase-2 in senescent rats (Calabrese et al, 2006c). It acts through the activaction of transcription factor Nrf2, which after binding to the ARE (antioxidant responsive element) in the HO-1 gene, up-regulates both HO-1 and thioredoxin reductase (TrxR), thus counteracting pro-oxidant conditions. Heme oxygenase-1 is, in fact, a key enzyme in the prevention of brain damage (Calabrese et al., 2006; Maines, 1997; Mancuso, 2004). The neuroprotective effects of over-expressed HO-1 has been attributed to several factors such as: a) increased level of both cGMP and bcl-2 in neurons; b) inactivation of the pro-apoptotic transcription factor p53; c) increase in antioxidant sources, i.e. the iron sequestering protein, as ferritin (Panahian, 1999). Hsp70 is, instead, a member of the stress protein. Hsc70 (heat shock cognate, the constitutive form), Hsp70 (the inducible form, also referred to as Hsp72) and GRP-75 (a constitutively expressed glucose-regulated protein) are included in this family (Calabrese et al., 2006b; Yenari et al., 1999). Recently it has been demonstrated that overproduction of Hsp70 leads to protection in several models of nervous system injury (Fig. 3). Oxidative stress, which has been suggested to be involved in the pathogenesis of PD, may originate in glial cells (Jenner, 2003). This is supported by postmortem studies demonstrating the capacity of oxidative stress and oxidizing toxin to induce nigral cell degeneration (Olanow et al., 1998). There is evidence to support that there are high levels of basal oxidative stress in the substantia nigra pars compacta in the normal brain and that this is increased in PD. The brain is therefore particularly sensitive to oxidative stress. This is due to several factors: a) the neurons are particularly enriched in polyunsaturated fatty acids, prime targets for oxidative attack (Kidd & Levine, 1985); b) the brain consumes a high share of the body's oxygen intake and consequently results in the oxyradical formation; c) the activity of the antioxidant enzymes catalase and peroxidase is low in the brain, instead the superoxide dismutase is active. They acquire superoxide oxyradical and convert it in hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>).

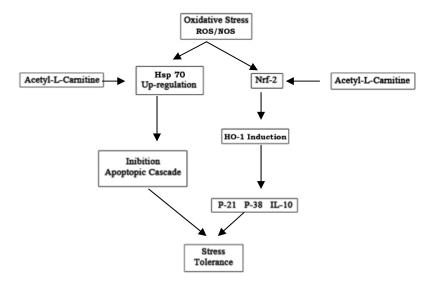


Fig. 3. The role of acetyl-L-carnitine in cell stress tolerance

In the absence of catalase and peroxidase, which normally would detoxify these peroxide products, that are done by glutathione peroxidase enzyme. This enzyme uses glutathione (GSH) as its essential cofactor, and when it is stimulated the brain's GSH reserves are more sensitive to depletion from oxidative attack (Kidd, 1997; Levine & Kidd 1985). As mentioned, the substantia nigra is particularly susceptible to oxidative stress and this is due to varied and many biochemical process that occur in it. It has a high content of dopamine and its metabolism and its "auto-oxidation" could be responsible for the high basal levels of oxidative stress. DA has a strong tendency to "auto-oxidation", generating reactive autometabolites, as 6-hydroxydopamine quinone and dopamine aminochrome, the formation of which can be accelerated by free (ionized) iron or by other redox-reactive elements such as copper, zinc or manganese (Pezzella, 1997; Youdim, 1989). Then, the degradation of dopamine by monoamine oxidase to produce  $H_2O_2$  could increase the formation of oxidized glutathione (GS-SG), suggesting the presence of oxidative stress and impairment of the

antioxidant system (Spina & Cohen, 1988). The H<sub>2</sub>O<sub>2</sub> generated is also converted (by Fenton reaction), in the presence of the high levels of iron, in toxic hydroxyl radical which can damage DNA and other biomolecules (Youdim et al., 1989). It should be noted that an extremely high content of iron is concentrated in the substantia nigra zona compacta, and various iron-mediated reactions in substantia nigra would potentiate oxidative stress. For example ionized iron, or copper and zinc catalyze transformation of the protein  $\alpha$ -synuclein into aggregated form, prominent component of the Lewy aggregates that develop in the SN of Parkinson's patients (Braak & Braak 2000; Paik et al., 2000). Moreover, important role of the iron is for dopamine-melanin, macromolecular material formed from the autoxidation of dopamine and normally scavenger of free radicals. When it is infiltrated with high levels of ionized iron it can drive Fenton reaction converting endogenous hydrogen peroxide to hydroxyl radical. The population of melanin-enriched, dopaminergic neurons found in the SN's zona compacta is the worst affected in PD. In the substantia nigra there are high levels of melanin and it could act as support matrix upon which ionized iron would catalyze oxyradical generation from available hydrogen peroxide or from neuromelanin itself (Youdim, et al., 1990).

#### 6. Conclusion

Although the selective loss of DA neurons and the accumulation of  $\alpha$ -synuclein are crucial in the development of PD, many evidences indicate that oxidative stress, and mitochondrial and proteasome dysfunctions have central role in this pathogenesis. Environmental factors, such as exposure to toxins, are also important in late-onset of the disease, whereas in earlyonset PD, genetic factors assume predominant importance. Recently, the identification of several genes causing early-onset PD (such as  $\alpha$ -synuclein; UCHLI, a ubiquitin carboxyterminal hydrolase L1; parkin; DJ1, a parkin-associated protein involved with oxidative stress; PINK1, a putative serine threonine kinase) has yielded crucial insights into the possible pathogenic mechanisms (Dawson & Dawson, 2003b). The use of several neurotoxins to produce the clinical symptoms of PD both *in vitro* and *in vivo* has allowed to understand the molecular mechanism of disease. The functions of mitochondria make these subcellular organelles susceptible to oxidative damage, resulting in cell death by apoptosis and mtDNA mutations. In this context, the mitochondria represent, therefore, a highly promising target for the development of disease biomarkers by use of genetic and biochemical approaches.

The mitochondrial antioxidant/nutrient acetyl-L-carnitine, with its antioxidant energizing protective activities and with its trophic effects, at optimal doses, can be an effective and safe prevention strategy for PD, offering the possibility of new and innovative therapeutic strategies for this neurodegenerative disease. Acetyl-L-carnitine is a highly bioavailable molecule, it is thought to penetrate the brain barrier better than carnitine, and it is readily converted to carnitine as needed. Experimental evidences suggest that acetyl-L-carnitine boosts mitochondrial ATP production and helps to protect mitochondria against oxidative attack. This molecule is therefore of great interest for its wide clinical application in various neurological disorders, it has beneficial effects in preventing the loss of brain function which typically occurs during aging, and its neuroprotective benefits have been observed in the hippocampus, prefrontal cortex, substantia nigra and muscarinic receptor portions of the brain. These include antioxidant activity, improved mitochondrial energetics, stabilization of intracellular membranes and cholinergic neurotransmission. In particular, the most

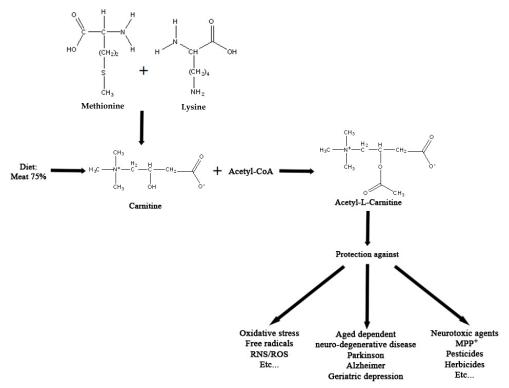


Fig. 4. Biosynthesis and physiological role of acetyl-L-carnitine

common function of acetyl-L-carnitine is the transport of fatty acids across the inner mitochondrial membrane, thereby being involved in the production of energy within the brain and in the maintenance of neuron and repairing of damages. Moreover, it has a variety of other neuronal effects. It increases protein kinase C (PKC) activity and reverse the agerelated decline in the number of N-methyl-D-aspartate (NMDA) receptors on neuronal membranes. In addition, it is thought that it influences the cholinergic system acting as a cholinergic receptor agonist; it can also promote the synthesis and the release of acetylcholine and stimulates proteins and membrane phospholipids synthesis (Calabrese et al., 2005). Acetyl-L-carnitine can also increase the levels of neurotrophins such as nerve growth factor (NGF) and can reduce the energetic deficits in brain and phospholipids metabolism, probably because it aids mitochondrial functions (Mark et al., 2003). In fact, it increases mitochondrial biogenesis and decreases ROS production through the upregulation of the PGC-1, as a possible underlying mechanism. In animal models, it partially protects the substantia nigra against 1-methyl-4-phenyl-pyridinium (MPP+, active metabolite of MPTP) attack, by strengthening the dopaminergic transmission (Bodis-Wollner et al. 1991; Hongyu et. al., 2010; Sinicropi et al. 2010). In fact, brain histology reveals that acetyl-L-carnitine protects neurons in the substantia nigra, which otherwise have been devastated by MPTP attack. It has been well observed that long-term acetyl-L-carnitine administration in rats increases longevity and improves spatial learning, avoidance learning in aged rats, and long-term memory performance (Barnes et al., 1990; Ghiraldi et al., 1989;

Markowska et al., 1990). In summary, the protection provided by acetyl-L-carnitine offers the possibility of new therapeutic strategies for neurodegenerative diseases (including PD) which can share the same final neurotoxic pathway in mitochondria (Fig. 4).

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#### 8. References

- Albertini, P., Amenta, F., Bissoni, G., Cavallotti, C., Felici, L., Ferrante, F. & Garivaghi, G. (1989). Effect of acetyl-L-carnitine treatment on the density of muscarinic receptor in the brain of methylazomethanol-microencephalic rats. *Drugs Under Experimental* & Clinical Research, Vol.15, pp. 421-427, ISSN 0378-6501.
- Anderson, R.F. & Harris, T.A. (2003). Dopamine and uric acid act as antioxidant in the repair of DNA radicals: implications in Parkinson's disease. *Free Radical Research*, Vol.37, No.10, (September 2003), pp. 1131-1136, ISSN 1029-2470.
- Annanmaki, T., Muuronen A. & Murros, K. (2007). Low plasma uric acid level in Parkinson's disease. *Movement Disorders*, Vol.22, No.8, (June 2007), pp. 1133-1137, ISSN 0885-3185.
- Barnes, C.A., Markowska, A.L., Ingram, D.K., Kametani, H., Spangler, E.L., Lemken, V.J. & Olton, D.S. (1990). Acetyl-L-carnitine 2: effects on learning and memory performance and aged rats in simple and complex mazes. *Neurobiology of Aging*, Vol.11, No.5, (September-October 1990), pp. 499-506, ISSN 0197-4580.
- Basu, N.K., Ciotti, M., Hwang, M.S., Kole, L., Mitra, P.S., Cho, J.W. & Owens, I.S. (2004). Differential and special properties of the major human UGT1-encoded gastrointestinal UDP-glucuronosyltransferases enhance potential to control chemical uptake. *The Journal of Biological Chemistry*, Vol.279, No.4, (January 2004), pp. 1429-1414, ISSN 1083-351X.
- Beal, M.F. (2003). Bioenergetic approachs for neuroprotection in Parkinson's disease. Annals of Neurology, Vol.53, No.S3, pp. S39-S48, ISSN 0364-5134.
- Bloem, B.R., Irwin, I., Buruma, O.J.S., Haan, J., Roos, R.A.C., Tetrud, J.W. & Langston, J.W. (1990). The MPTP model: versatile contributions to the treatment of idiopathic Parkinson's disease. *Journal of Neurological Sciences*, Vol.97, No.2-3, (July 1990), pp. 273-293, ISSN 0022-510X.
- Bodis-Wollner, I., Chung, E., Ghilardi, M.F., Glover, A., Onofrj, M., Pasik, P. & Samson, Y. (1991). Acetyl-levo-carnitine protects against MPTP-induced parkinsonism in primates. *Journal of Neural Transmission-Parkinson's Disease and Dementia Section*, Vol.3, No.1, pp. 63-72, ISSN 0936-3076.
- Bovè, J., Prou, D., Perier, C. & Przedborski, S. (2005). Toxin-induced model of Parkinson's disease. *NeuroRx*, Vol.2, (July 2005), pp. 484-494, ISSN 1545-5351.
- Boveris, A.; Oshino, N. & Chance, B. (1972). The cellular production of hydrogen peroxide. *Biochemical Journal*, Vol.128, No.3, pp. 617-630, ISSN 1470-8728.
- Braak, H. & Braak, E. (2000) Pathoanatomy of Parkinson's disease. *Journal of Neurology*, Vol.247, No.S6 (June 2000), pp. 3-10, ISSN 0340-5354.

- Braak, H., Del Tredici, K., Rub, U., de Vos, R.A., Jansen Steur, E.N. & Braak, E. (2003). Staging of brain pathology related to sporadic Parkinson's disease. *Neurobiology of Aging*, Vol.24, No.2, (March-April 2003), pp. 197-211, ISSN 0197-4580.
- Bremer, J. (1962). Carnitine in Intermediary Metabolism: The metabolism of fatty acid esters of carnitine by mitochondria. *The Journal of Biological Chemistry*, Vol.237, No.12, (December 1962), pp. 3628-3632, ISSN 1083-351X.
- Broocks, D.R. (1999). Anticholinergic drugs used in Parkinson's disease. An overlooked class of drugs from a pharmacokinetic perspective. *Journal of Pharmacy and Pharmaceutical Sciences*, Vol.2, No.2, (May-August 1999), pp. 39-46, ISSN 1482-1826.
- Calabrese, V., Ravagna, A., Colombrita, C., Scapagnini, G., Guagliano, E., Calvani, M., Butterfield, D.A. & Giuffrida Stella, A.M. (2005). Acetylcarnitine induces heme oxygenase in rat astrocytes and protects agonist oxidative stress: involvement of the transcription factor Nrf2. *Journal of Neuroscience Research*, Vol.79, No.4, (February 2005), pp. 509-521, ISSN 1097-4547.
- Calabrese, V., Giuffrida Stella, A.M., Calvani, M. & Butterfield, D.A. (2006a) Acetylcarnitine and cellular stress response: roles in nutritional redox homeostasis and regulation of longevity genes. *Journal of Nutritional Biochemistry*, Vol.17, No.2, (February 2006), pp. 73-88, ISSN 0955-2863.
- Calabrese, V., Butterfield, D.A., Scapagnini, G., Giuffrida Stella, A.M. & Maines, M.D. (2006b). Redox regulation of heat shock protein expression by signaling involving nitric oxide and carbon monoxide: relevance to brain aging, neurodegenerative disorders, and longevity. *Antioxidants & Redox Signaling*, Vol.8, No.3-4, (March-April 2006), pp. 444-477, ISSN 1557-7716.
- Calabrese, V., Colombrita, C., Sultana, R., Scapagnani, G., Calvani, M., Butterfield, D.A. & Giuffrida Stella, A.M. (2006c). Redox modulation of heat shock protein expression by acetylcarnitine in aging brain: relationship to antioxidant status and mitochondrial function. *Antioxidants & Redox Signaling*, Vol.8, No.3-4, (March-April 2006), pp. 404-416, ISSN 1557-7716.
- Calabrese, V., Cornelius, C., Mancuso, C., Pennisi, G., Calafato. S., Bellia, F., Bates, T.E., Giuffrida Stella, A.M., Schapira, T., Dinkova Kostova, A. & Rizzarelli, E. (2008). Cellular stress response: a novel target for chemoprevention and nutritional neuroprotection in aging, neurodegenerative disorders and longevity. *Neurochemical Research*, Vol.33, No. 12, (December 2008), pp. 2444-2471, ISSN 1573-6903.
- Cancelli, I., Beltrame, M., Gigli, G.L. & Valente, M. (2009). Drugs with anticholinergic properties: Cognitive and neuropsychiatric side-effects in elderly patients. *Neurological Sciences*, Vol.30, No.2, (April 2009), pp. 87-92, ISSN 1590-3478.
- Canesi, M., Zecchinelli, A.L., Pezzoli, G. & Antonini, A. (2008). Clinical experience of tolcapone in advances Parkinson's disease. *Neurological Sciences*, Vol.29, No.5, (December 2008) pp. S380–382, ISSN 1590-3478.
- Caparros-Lefebvre, D. & Elbaz, A. (1999) Possible relation of atypical parkinsonism in the French West Indies with consumption of tropical plants: a case control study. Caribbean Parkinsonism Study Group. *The Lancet*, Vol.354, No.9175, (July 24, 1999), pp. 281-286, ISSN 0140-6736.

- Casal, J., Elizan, T.S. & Yahr, M.D. (1998). Postencephalitic parkinsonism a review. *Journal of Neural Transmission*, Vol.105, No. 6-7, (September 1998), pp. 645-676, ISSN 0300-9564.
- Cavalli, A., Bolognesi, M.L., Minarini, A., Rosini, M., Tumiatti, V., Recanatini, M. & Melchiorre, C. (2008). Multi-target-Direct Ligands to Combat Neurodegenerative Disease. *Journal of Medicinal Chemistry*, Vol.51, No.3, (February 2008), pp. 347-372, ISSN 0022-2623.
- Chan, D.K., Woo, J., Ho, S.C., Pang, C.P., Law, L.K., Ng, P.W., Hung, W.T., Kwok, T., Hui, E., Orr, K., Leung, M.F. & Kay, R. (1998). Genetic and environmental risk factors for Parkinson's disease in a Chinese population. *Journal of Neurolology, Neurosurgery, & Psychiatry*, Vol.65, No.5, (November 1998), pp. 781-784, ISSN 1468-330X.
- Chao, J., Yu, M.S., Ho, Y.S., Wang, M. & Chahg, R.C.C. (2008) Dietary oxyresveratrol prevents parkinsonian mimetic 6-hydroxydopamine neurotoxicity. *Free Radical Biology and Medicine*, Vol.45, No.7, (October 2008), pp. 1019-1026, ISSN 0891-5849.
- Chase, T.N. & Oh, J.D. (2000). Striatal dopamine- and glutamate-mediate dysregulation in experimental parkinsonism. *Trends in Neuroscience*, Vol.23, No.S10, (October 2000), pp. S86-S91, ISSN 0166-2236.
- Chen, J., Tang, X.P, Zhi, J.L., Cui, Y., Yu, H.M., Tang, E.H., Sun, S.N., Feng, J.Q. & Chen, P.X. (2006) Curcumin protects PC12 cells against 1-methyl-4-phenylpyridinium ioninduced apoptosis by bcl-2 mitochondria-ROS-iNOS pathway. *Apoptosis*, Vol.11, No.6, (June 2006), pp.943-953, ISSN 1573-675X.
- Choi J.Y., Park C.S., Kim D.J., Cho M.H., Jin B.K., Pie J.E. & Chung W.G. 2002. Prevention of nitric oxide-mediated 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine-induced Parkinson's disease in mice by tea phenolic epigallocatechin 3-gallate. *Neurotoxicology*, Vol.23, No.3, (September 2002), p. 367-374, ISSN 0161-813X.
- Crag, S.J. (2006). Meaningful silence: How dopamine listens to the Ach pause. *Trends in Neuroscience*, Vol.29, No.3, (March 2006), pp. 125-131, ISSN 0166-2236.
- Dawson, T.M., & Dawson V.L. (2003a) Molecular pathways of neurodegeneration in Parkinson's disease. Science, Vol.302, No.5646, (October 2003), pp. 819-822, ISSN 1095-9203.
- Dawson, T.M., & Dawson, V.L. (2003b). Rare genetic mutation shed light on the pathogenesis of Parkinson disease. *The Journal of Clinical Investigation*, Vol.111, No.2, (January 2003), pp. 145-151, ISSN 0021-9738.
- deRijk, M.C., Launer, L.J., Berger, K., Breteler, M.M., Dartigues, J.F., Baldereschi, M., Fratiglioni, L., Lobo, A., Martinez-Lage, J., Trenkwalder, C., & Hofman, A. (2000). Prevalence of Parkinson's disease in Europe: a collaborative study of populationbased cohorts. *Neurology*, Vol.54, No.11, pp. S21–S23, ISSN 0028-3878.
- Dexter, D.T., Holley, A.E., Flitter, W.D., Slater, TF, Wells, F.R., Daniel, S.E., Lees, A.J., Jenner, P. & Marsden, C.D. (1994). Increased levels of lipid hydroperoxides in the Parkinsonian substantia nigra: an HPLC and ESR study. *Movement Disorders*, Vol.9, No.1, pp. 92-97, ISSN 0885-3185.
- Di Monte, D.A.; Lavasani, M. & Manning-Bog, A.B. (2002). Environmental factors in Parkinson's disease. *Neurotoxicology*, Vol.23, No.4-5, (October 2003), pp. 487-502, ISSN 0161-813X.
- Duan, W., Ladenheim, B., Cutler, R.G., Kruman III, I.I., Cadet, J.L. & Mattson, M.P. (2002). Dietary folate deficiency and elevated homocysteine levels endanger dopaminergic

neurons in models of Parkinson's disease. *Journal of Neurochemistry*, Vol.80, No.1, (January 2002), pp. 101-110, ISSN 1471-4159.

- Encarnation, E.V. & Hauser, R.A. (2008). Levodopa-induced dyskinesias in Parkinson's disease: Etiology, impact on quality of life, and treatments. *European Neurology*, Vol.60, No.2, (July 2008), pp. 57-66, ISSN 0014-3022.
- Fernandez, H.H. & Chen, J.J. (2007). Monoamine oxidase inhibitors: current and emerging agents for Parkinson disease. *Clinical Neuropharmacology*, Vol.30, No.3, (May-June 2007), pp. 150-168, ISSN 0362-5664.
- Fonck, C. & Baudry, M. (2003) Rapid reduction of ATP synthesis and lack of free radical formation in rat brain synaptosomes and mitochondria. *Brain Research*, Vol.975, No.1-2, (June 2003), pp. 214-221, ISSN 0006-8993.
- Fox, S.H. & Lang, A.E. (2008). Levodopa-related motor complications-phenomenology. *Movement disorders*, Vol.23 No.3, pp. S509-S514, ISSN 0885-3185.
- Ghiraldi, O., Milano S., Ramanacci, M.T. & Angelucci, L. (1989). Long-term acetyl-Lcarnitine preserves spatial learning in the senescent rat. *Progress in Neuro-Psychopharmacology and Biological Psychiatry*, Vol.13, No.1-2, pp. 237-245, ISSN 0278-5846.
- Giasson, B.I, Duda, J.E., Murray, I.V., Chen, Q., Souza, J.M., Hurtig, H.I., Ischiropoulos, H., Trojanowski, J.Q. & Lee, V.M. (2000). Oxidative damage linked to neurodegeneration by selective alpha-synuclein nitration in synucleinopathy lesions. *Science*, Vol.290, No.5493, (November 2000), pp. 985–989, ISSN 1095-9203.
- Gille, G., Hung, S.T., Reichmann, H. & Rausch, W.D. (2004). Oxidative stress to dopaminergic neurons as models of Parkinson's disease. *Annals of the New York Academy of Science*, Vol.1018, (June 2004), pp. 533-540, ISSN 00778923.
- Gorell, J.M., Johnson, C.C., Rybicki, B.A., Peterson, E.L. & Richerdson, R.J. (1998) The risk of Parkinson's disease with exposure to pesticides, farming, well water, and rural living. *Neurology*, Vol.50, No.5, (May 1998), pp. 1346-1350, ISSN 0028-3878.
- Gu, M., Cooper, J.M-, Taanmann, J.W. & Schapira A.H.V. (1998). Mitochondrial DNA transmission of the mitochondrial defect in Parkinson's disease. *Annals of Neurology*, Vol.44, No.2, (August 1998), pp. 177-186, ISSN 0364-5134.
- Guo, S., Bezard, E & Zhao, B. (2005). Protective effect of green tea polyphenols on the SH-SY5Y cells against 6-OHDA induced apoptosis through ROS-NOS pathway. *Free Radical Biology and Medicine*, Vol.39, No.5, (September 2005), pp. 682-695, ISSN 0891-5849.
- Haas, R.H., Nasirian, F., Nakano, K., Ward, D., Pay, M., Trojanowski, J.Q. & Lee, V.M. (1995). Low platelet mitochondrial complex I and complex II/III activity in early untreated Parkinson's disease. *Annals of Neurology*, Vol.37, No.6, (June 1995), pp. 714-722, ISSN 0364-5134.
- Halliwell, B. (2006) Oxidative stress and neurodegeneration: where are we now? *Journal of Neurochemistry*, Vol.97, No.6, (June 2006), pp. 1634-1658, ISSN 1471-4159.
- Hancock, D.B., Martin, E.R., Mayhew, G.M., Stajich, J.M., Jewett, R., Stacy, M.A., Scott, B.L., Vance, J.M., & Scott W.R., (2008). Pesticide exposure and risk of Parkinson's disease. A family –based case- control study. *BMC Neurology*, Vol.8, No.6, (March 2008), pp. 6-17, ISSN 1471-2377.
- Hayeshi, R., Mutingwende, I., Mavengere, W., Masiyanise, V. & Mukanganyama, S. (2007). The inhibition of human glutathione S-transferase activity by plant polyphenolic

compounds ellagic acid and curcumin. *Food and Chemical Toxicolology*, Vol.45, No.2, (February 2007), pp. 286-295, ISSN 0278-6915.

- Hong, J.S. (2005). Role of inflammation in the pathogenesis of Parkinson's disease: Models, mechanisms, and therapeutic interventions. *Annals of the New York Academy of Science*, Vol.1053, (August 2005), pp. 151–152, ISSN 00778923.
- Hongyu, Z., Haiqun, J., Jianghai, L., Ni, A., Bing,Y., Weili, S., Xuemin, W., Xin, L., Cheng, L. & Jiankang, L. (2010) Combined R-α-lipoic acid and acetyl-L-carnitine exerts efficient preventative effects in a cellular model of Parkinson's disease. *Journal of Cellular and Molecular Medicine*, Vol.14, No.1-2, (January-February 2010), pp. 215-225, ISSN 1582-1838.
- Ireson, C.R., Jones, D.J., Orr, S., Coughtric, M.W., Boocock, D.J., Williams, M.L., Farmer, P.B., Steward, W.P. & Gescher, A.J. (2002). Metabolism of the cancer chemopreventive agent curcumin in human and rat intestine. *Cancer Epidemiology, Biomarkers & Prevention*, Vol.11, No.1, (January 2002), pp. 105-111, ISSN 1538-7755.
- Jenner, P. (2003). Oxidative stress in Parkinson's disease. Annals of Neurology, Vol.53, No.3, pp. S26-S38, ISSN 0364-5134.
- Jeulin , C., Soufir, J.C., Marson, J., Paquignon, M. & Dacheux, J.L. (1988). Acetylcarnitine and spermatozoa: relationship with epididymal maturation and motility in the boar and man. *Reproduction Nutrition Development*, Vol.28, No.5, pp. 1317-1327, ISSN 1297-9708.
- Johnson, K.A., Conn, P.J. & Niswender, C.M. (2009), Glutamate receptors as therapeutic targets for Parkinson's disease. CNS and Neurological Disorders- Drug Targets, Vol.8, No.6, (December 2009), pp. 475-491, ISSN 1871-5273.
- Kidd P.M. (1997). Gluthatione: systemic protectant against oxidative and free radical damage. *Alternative Medicine Review*, Vol.2, pp. 155-176, ISSN 1089-5159.
- Kidd, P.M. & Levine, S.A. (1985). The biochemistry of free radicals, In: Antioxidant Adaptation: Its Role in Free Radical Pathology, S.A. Levine, P.M. Kidd, (Ed.) El Cerrito, ISBN 0961463007, CA: PMK
- Kido, J., Tamai, I., Ohnari, A., Sai, Y., Kagami, T., Nezu, J., Nikaido, H., Hashimoto, N., Asano, M. & Tsuji, A. (2001). Functional relevance of carnitine transporter OCTN2 to brain distribution of L-carnitine and acetyl-L-carnitine across the blood-brain barrier. *Journal of Neurochemistry*, Vol.79, No. 5, (November 2001), pp. 959-969, ISSN 1471-4159.
- Klaidman, L.K., Adams Jr., J.D., Leung, A.C., Kim, S.S. & Cadenas, E. (1993). Redox cycling of MPP<sup>+</sup>: evidence for a new mechanism involving hydride transfer with xanthine oxidase, aldehyde dehydrogenase, and lipoamide dehydrogenase. *Free Radical Biology and Medicine*, Vol.15, No.1, (July 1993), pp. 69-179, ISSN 0891-5849.
- Lee, V. M. & Trojanowsky, J.Q. (2006). Mechanisms of Parkinson's disease linked to pathological alpha-synuclein: new targets for drug discovery. *Neuron*, Vol.52, No.1, (October 2006), pp. 33–38, ISSN 0896-6273.
- Lees, A. (2005). Alternatives to levodopa in the initial treatment of early Parkinson's disease. *Drugs & Aging*, Vol.22, No.9, pp. 731-740, ISSN 1170-229X.
- Levine, S.A. & Kidd, P.M. Antioxidant Adaptation: Its Role in Free Radical Pathology. San Leandro C.A. Biocurrents/Allergy Research group,1985
- Liu, Y., Roghani, A. & Edwards R.H. (1992). Gene transfer of a reserpine-sensitive mechanism of resistance to N-methyl-4-phenylpyridinium. *Proceedings of the*

National Academy of Sciences of the United States of America. Vol.89, No.19, (October 1992), 9074-9078, ISSN 0027-8424.

- Lorenz, P., Roychowdhury, M., Engelmann, M., Wolf, G. & Horn, T.F.W. (2003). Oxyresveratrol and resveratrol are potent antioxidant and free radical scavengers: effect on nitrosative and oxidative stress derived from microglial cells. *Nitric oxide – Biology and Chemistry*. Vol.9, No.2, (September), pp. 64-76, ISSN 1089-8603.
- Maines, M.D. (1997). The heme oxygenase system: a regulator of second messenger gases. *Annuual Review of Pharmacolology and Toxicology*, Vol.37, (April 1997), pp. 517-554, ISSN 0362-1642.
- Mancuso, C. (2004). Heme oxygenase and its products in the nervous system. *Antioxidants and Redox Signling*, Vol.6, No.5, (October 2004), pp. 878-887, ISSN 1523-0864.
- Markowska, A.L., Ingram, D.K., Barnes, C.A., Spangler, E.L., Lemken, V.J., Kametani, H., Yee, W. & Olton, D.S. (1990). Acetyl-L-carnitine 1: effects on mortality; pathology and sensory-motor performance in aging rats. *Neurobiology of Aging*, Vol.11, No.5, (September-October 1990), pp. 491-498, ISSN 0197-4580.
- Marquis, N.R. & Fritz, I.B. (1965). The distribution of carnitine, acetylcarnitine and acetyl transferase in rat tissues. *Journal of Biological Chemistry*, Vol.240, No.5, (May 1965), pp. 2193-2196, ISSN 1083-351X.
- McDaniel, M.A., Maier, S.F., & Einstein, G.O. (2003). "Brain-Specific" Nutrients: A Memory Cure? *Nutrition*, Vol.19, No.11 (November 2003), pp. 957-975, ISSN 0899-9007.
- McNaught, K.S. & Jenner, P. (2001). Proteasomal function is impaired in substantia nigra in Parkinson's disease. *Neuroscience Letters*, Vol.297, No.3, (January 2001), pp. 191-194, ISSN 0304-3940.
- McNaught, K.S., Olanow, C.W., Halliwell, B., Isacson, O. & Jenner, P. (2001). Failure of the ubiquitin-proteasome system in Parkinson's disease. *Nature Review Neuroscience*, Vol.2, No.8, (August 2001), pp. 589-594, ISSN 1471-0048.
- Metman, V.L., Del Dotto, P., van den Munckhog, P., Fang, J., Mouradian, M.M. & Chase, T.N. (1998). Amantadine as treatment for dyskinesias and motor fluctuations in Parkinson's disease. *Neurology*, Vol.50, No. 5, (May 1998), pp. 1323-1326 ISSN 0028-3878.
- Mercuri, N.B. & Bernardi, G. (2005). The 'magic' of L-dopa: Why is the gold standard Parkinson's disease therapy? *Trends in Pharmacological Sciences*, Vol.26, No.7, (July 2005), pp. 341–344, ISSN 0165-6147.
- Mizuno, Y., Saitoh, T. & Sone, N. (1987). Inhibition of mitochondrial alpha-ketoglutarate dehydrogenase by 1-methyl-4-phenylpyridinium ion. *Biochemical and Biophysical Research Communications*, Vol.143, No. 3, (March 1987), pp. 971-976, ISSN 0006-291X.
- Mythri, R.B., Jagatha, B., Pradhan, N., Andersen, J. & Bharath, M.M. (2007). Mitochondrial complex I inhibition in Parkinson's disease; how can curcumin protect mitochondria? *Antioxidant and Redox Signaling*, Vol.9, No.3, (March 2007), pp. 399-408, ISSN 1523-0864.
- Olanow C.W., Jenner P., Tatton N.A. & Tatton W.G. (1998). Neurodegeneration and Parkinson's disease, In *Parkinson's disease and movement disorders*, J. Jankovic, E. Tolosa, (Ed.), 67-103. Lippincott Williams & Wilkins, ISBN 0781778816, Baltimore.
- Olanow, C.W., Jenner, P. & Youdim, M. Neurodegeneration and neuroprotection in Parkinson's disease. New York, NY: Academic Press: 1996.

- Olanow, C.W. (2007). Pathogenesis of cell death in Parkinson's disease. *Movement Disorders*, Vol.22, No.S17, pp. S335-S342, ISSN 0885-3185.
- Olanow, C.W., Stern, M.B. & Sethi, K. (2009). The scientific and clinical basis for the treatment of Parkinson disease. *Neurology*, Vol.72, No.21, (May 2009), pp. S1–S136, ISSN 0028-3878.
- Ono, K. & Yamada, M. (2006). Antioxidant compounds have potent anti-fibrillogenic and fibril-destabilizing effects for alpha-synuclein fibrils *in vitro*. *Journal of Neurochemistry*, Vol.97, No.1, (April 2006), pp. 105-115, ISSN 1471-4159.
- Paik, S.R., Shin, H.J. & Lee, J.H. (2000). Metal-catalyzed oxidation of alpha-synuclein in the presence of copper (II) and hydrogen peroxide. *Archives of Biochemistry and Biophysics*, Vol.378, pp. 269-277.
- Panahian, N., Yoshiura, M. & Maines, M.D. (1999) Overexpression of heme oxygenase-1 is neuroprotective in a model of permanent middle cerebral artery occlusion in transgenic mice. *Journal of Neurochemistry*, Vol.72, No.3, (March 1999), pp. 1187-1203, ISSN 1471-4159.
- Pandey, N. & Galvin, J.E. (2005) Curcumin prevents aggregation of alpha-synuclein. *Society for Neuroscience,* Vol.31, abs 1007.9.
- Parker Jr., W.D., Boyson, S.J. & Parks, J.K. (1989). Abnormalities of the electron transport chain in idiopathic Parkinson's disease. *Annals of Neurology*, Vol.26, No.6, (December 1989), pp. 719-723, ISSN 0364-5134.
- Parris, M.K. (2000). Parkinson's disease as multifactorial oxidative neurodegeneration: implications for integrative management. *Alternative Medicine Review*, Vol.5, pp. 502-545, ISSN 1089-5159.
- Peeters, M., Maloteaux, J.M. & Hermans, E. (2003). Distinct effects of amantadine and memantine on dopaminergic transmission in the rat striatum. *Neuroscience Letters*, Vol.343, No.3, (June 2003), pp. 205-209, ISSN 0304-3940.
- Perlmutter, D. BrainRecovery.com: Powerful therapy for challenging brain disorders. Naples, FL: The Perlmutter Health Center; www.brainrecovery.com; 2000.
- Pettegrew, J.W., Levine, J. & McClure, R.J. (2000) Acetyl-L-carnitine physical-chemical, metabolic, and therapeutic properties: relevance for its mode of action in Alzheimer's disease and geriatric depression. *Molecular Psychiatry*, Vol.5, No.6, (November), pp. 616-632, ISSN 1476-5578.
- Pezzella, A., d'Ischia, M., Napolitano, A., Misuraca, G. & Prota, G. (1997). Iron-mediated generation of the neurotoxin 6-hydroxydopamine quinone by reaction of fatty acid hydroperoxides with dopamine: as possible contributory mechanism for neuronal degeneration in Parkinson's disease. Journal of Medicinal Chemistry, Vol.40, No.14, (July 1997), pp. 2211-2216, ISSN 0022-2623.
- Postuma, R.B. & Lang, A.E. (2004). Homocysteine and levodopa: should Parkinson disease patients receive preventive therapy? *Neurology*, Vol.63, No.5, (September 2004), pp. 886-891, ISSN 0028-3878.
- Przedborski, S. & Vila, M. (2003). The 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine mouse model: a tool to explore the pathogenesis of Parkinson's disease. *Annals of the New York Academy of Sciences*, Vol.991, (June 2003), pp. 189-198 ISSN 00778923.
- Rahaman, I., Biswas, S.K. & Kirkam, P.A. (2006). Regulation of inflammation and redox signalling by dietary polyphenols. *Biochemical Pharmacology*, Vol.72, No.11, (November 2006), pp. 1439-1452, ISSN 0006-2952.

- Rajeswary, A. (2006). Curcumin protects mouse brain from oxidative stress caused by 1methyl-4-phenyl-1,2,3,6-tetrahydropyridine. *European Review for Medical and Pharmacological Sciences*, Vol.10, No.4, pp. 157-161, ISSN 1128-3602.
- Ramsay, R.R. & Singer, T.P. (1986) Energy-dependent uptake of N-methyl-4phenylpyridinium, the neurotoxin metabolite of 1-methyl-4-phenyl-1,2,3,6tetrahydropyridine, by mitochondria. *Journal of Biological Chemistry*, Vol.261, No.16, (June 1896), pp. 7585-7587, ISSN 1083-351X.
- Ramsay, R.R., Krueger, M.J., Youngster, S.K., Gluck, M.R., Casida, J.E. & Singer T.P. (1991). Interaction of 1-methyl-4-phenylpyridinium ion (MPP) and its analogs with the rotenone/piericidin binding site of NADH dehydrogenase. *Journal of Neurochemistry*, Vol.56, No.4, (April 1991), pp. 1184-1190, ISSN 1471-4159.
- Rao, K.V., Mawal, Y.R. & Qureshi I.A. (1997). Progressive decrease of cerebral cytochrome C oxidase activity in sparse-fur mice: role of acetyl-L-carnitine in restoring the ammonia-induced cerebral energy depletion. *Neuroscience Letters*, Vol.224, No.2, (March 1997), pp. 83-86, ISSN 0304-3940.
- Sancher-Ramos, T.R., Overvik, E. & Ames, B.N. (1994). A marker of oxyradical-mediated DNA damage (8-hydroxy-2'-deoxyguanosine) is increased in nigro-striatum of Parkinson's disease brain. *Neurodegeneration*, Vol.3, pp. 197-201, ISSN 1068-719X.
- Saunders CD. Parkinson's disease: a new hope. Boston MA: Harvard Health Publication; 2000.
- Schapira, A.H.V. (2003). Neuroprotection in PD a role for dopamine agonists? *Neurology*, Vol.61, No.6, (September 2003), pp. S34–S42, ISSN 0028-3878.
- Schapira, A.H.V. (2008). Mitochondria in the aetiology and pathogenesis of Parkinson's disease. *The Lancet Neurology*, Vol.7, No.1, (January 2008), pp. 97-109, ISSN 1474-4422.
- Schrag, A. (2005). Entacapone in the treatment of Parkinson's disease. *The Lancet Neurology*, Vol.4, No.6, (June 2005), pp. 366–370, ISSN 1474-4422.
- Sculer, F. & Casida, J.E. (2001) Functional coupling of PSST and ND1 subunits in NADH: ubiquinone oxidoreductase ewstablished by photoaffinity labelling. *Biochimica and Biophysica Acta* Vol.1506, pp. 79-87, ISSN:0006-3002.
- Seidler, A., Hellenbrand, W., Robra, B.P., Vieregge, P., Nischan, P., Joerg, J., Oertel, H., Ulm, G. & Schneider, E. (1996). Possible environmental, occupational, and other etiologic factors for Parkinson's disease: a case-control study in Germany. *Neurology*, Vol.46, No.5 (May 1996), pp. 1275-1284, ISSN 0028-3878.
- Sherer, T.B., Kim, J.H., Betarber, R. & Greenamyre, J.T. (2003) Subcutaneous rotenone exposure causes highly selective dopaminergic degeneration and α-synuclein aggregation. *Experimental Neurology*, Vol.179, No.1, (January 2003), pp. 9-16, ISSN 0014-4886.
- Shergill, J.K., Cammack, R., Cooper, C.E., Cooper, J.M., Mann, V.M. & Schapira, A.H.V. (1996). Detection of nitrosyl complexes in human substantia nigra in relation to Parkinson's disease. *Biochemical and Biophysical Research Communications*, Vol.228, No.2, (November 1996), pp. 298-303, ISSN 0006-291X.
- Sesadri, S., Beiser, A., Selhub, J., Jacques, P.E., Rosenberg, I.H., D'Agostino, R.B., Wilson, P.W.F. & Wolf, P.A. (2002). Plasma homocysteine as a risk factor for dementia and Alzheimer disease. *New England Journal of Medicine*, Vol.346, No.7, (February 2002), pp. 476-483, ISSN 1533-4406.

- Shults, C.W., Haas, R.H., Passov, D. & Beal, M.F. (1997). Coenzyme Q10 is reduced in mitochondria from parkinsonian patients. *Annals of Neurolology*, Vol.42, No.2, (August 1997), pp. 261-264, ISSN 0364-5134.
- Shults, C.W., Beal, M.F., Fontaine, D., Nakano, K. & Haas, R.H. (1998). Absorption, tolerability, and effects on mitochondrial activity of oral coenzyme Q10 in parkinsonian patients. *Neurology*, Vol.50, No.3, (March 1998), pp. 793-795, ISSN 0028-3878.
- Shults, C.W., Oakes, D., Kieburtz, K., Beal, M.F., Haas, R., Plumb, S., Juncos, J.L., Nutt, J., Shoulson, I., Carter, J., Kompoliti, K., Perlmutter, J.S., Reich, S., Stern, M., Watts, R.L., Kurlan, R., Molho, E, Harrison, M. & Lew, M. (2002). The Parkinson Study Group. Effects of coenzyme Q10 in early Parkinson's disease: evidence of slowing of the functional decline. *Archives of Neurolology*, Vol.59, No.10, (October 2002), pp. 1541-1550, ISSN 0375-8540.
- Singh, N., Pillay, V. & Choonara, Y. E. (2007). Advances in the treatment of Parkinson's disease. *Progress in Neurobiology*, Vol.81, No.1, (January 2007), pp. 29–44, ISSN 0301-0082.
- Sinicropi, M.S., Leone, F., Rovito, N. & Genchi, G. (2010). Behaviour of acetyl-L-carnitine injections (Nicetile® fiale) with different drugs used for combined therapy. *Advances in Therapy*, Vol.27, No.8, (August 2010), pp. 547-554, ISSN 0741-238X.
- Spina M.B., & Cohen, G. (1988). Exposure of striatal synaptosomes to L-dopa increases levels of oxidized glutathione. *Journal of Pharmacology and Experimental Therapeutics*, Vol.247, No.2, (November 1988), pp. 502-507, ISSN 1521-0103.
- Stephenson, J. (2000). Exposure to home pesticides linked to Parkinson's disease. Journal of the American Medical Association. Vol.283, No.23, (June 2000), pp. 3055-3056, ISSN 1538-3598.
- Stoof, J.C., Booij, J. & Drukarch, B. (1992). Amantadine as a N-methyl-D-aspartic acid receptor antagonist: new possibilities for therapeutic application? *Clinical Neurology* and Neurosurgery, Vol.94, No.S1, pp. S4–S6, ISSN 0303-8467.
- Storch, A., Jost, W.H., Vieregge, P., Spiegel, J., Greulich, W., Durner, J., Muller, T., Kupsch, A., Henningsen, H., Oertel, W.H., Fuchs, G., Kuhn, W., Niklowitz, P., Koch, R., Herting, B. & Reichmann, H. (2007). Randomized, double-blind, placebo-controlled trial on symptomatic effects on coenzyme Q(10) in Parkinson disease. *Archives of Neurology*, Vol.64, No.7, (July 2007), pp. 938-944, ISSN 0375-8540.
- Swerdlow, R.H., Parks, J.K., Miller, S.W., Tuttle, J.B., Trimmer, P.A., Sheenam, J.P., Bennett, J.P., Davis, R.E. & Parker, W.D. (1996). Origin and functional consequences of the complex I defect in Parkinson's disease. *Annals of Neurology*, Vol.40, No.4, (October 1996), pp. 663-671, ISSN 0364-5134.
- Thal, L.J., Carta, A., Clarke, W.R., Ferris, S.H., Friedland, R.P., Petersen, R.C., Pettegrew, J.W., Pfeiffer, E., Raskind, M.A., Sano, M., Tuszynski, M.H. & Wodson, R.F. (1996). A 1-year multicenter placebo-controlled study of acetyl-L-carnitine in patients with Alzheimer's disease. *Neurology*, Vol.47, No.3, (September 2007), pp. 705-711, ISSN 0028-3878.
- Thapliyal, R. & Maru, G.B. (2001). Inhibition of P450 isoenzymes by curcumin *in vitro* and *in vivo*. *Food and Chemical Toxicolology*, Vol.39, No.6, (June 2001), pp. 541-547, ISSN 0278-6915.

- Virmani, A. & Binienda, Z. (2004). Role of carnitine esters in brain neuropathology. Molecular Aspects of Medicine, Vol.25, No.5-6, (October-December 2004), pp. 533-549, ISSN 0098-2997.
- Weisskopf, M.G., O'Reilly, E., Chen, H., Schwarzschild, M.A. & Ascherio, A. (2007). Plasma urate and risk of Parkinson' disease. *American Journal of Epidemiology*, Vol.166, No.5, (September 2007), pp. 561-567, ISSN 1476-6256.
- Winquist, A., Steenland, K. & Shankar, A. (2010). Higher serum uric acid associated with decreased Parkinson's disease prevalence in a large community-based survey. *Movement disorders*, Vol.25, No.7, (May 2010), pp. 932-936, ISSN 0885-3185.
- Wood-Kaczamar, A., Gandhi, S. & Wood, N.W. (2006). Understanding the molecular causes of Parkinson's disease. *Trends in Molecular Medicine*, Vol.12, No.11, (November 2006), pp. 521–528, ISSN 1471-4914.
- Yenari, M.A., Giffard, R.G., Sapolosky, R.M. & Steinberg, G.K. (1999). The neuroprotective potential of heat shock protein 70 (HSP70). *Molecular Medicine Today*, Vol.5, No.12, (Dicember 1999), pp. 525-531, ISSN 1357-4310.
- Youdim, M.B., Ben Schachar, D. & Riederer, P. (1989). Is Parkinson's disease a progressive siderosis of substantia nigra resulting in iron and melanin induced neurodegeneration? *Acta Neurologica Scandinavica*, Vol.80, No.S126, (November 1989), pp. 47-54, ISSN 1600-0404.
- Youdim, M.B.H, Ben-Schachar, D. & Riederer, P. (1990). The role of monoamine oxidase, iron-melanin interaction, and intracellular calcium in Parkinson's disease. *Journal of Neural Transmission*, Vol.32, pp. S239-S248, ISSN 0300-9564.
- Zhang, K. & Kaufman, R.J. (2006). The unfolded protein response: a stress signaling pathway critical for health and disease. *Neurology*, Vol.66, No.2, (January 2006), pp. S102-S109, ISSN 0028-3878.