



Invited Review Article

Mineral requirements for mitochondrial function: A connection to redox balance and cellular differentiation ☆

David W. Killilea ^a  , Alison N. Killilea ^b

[Show more](#) ▾

 Outline |  Share  Cite

<https://doi.org/10.1016/j.freeradbiomed.2022.02.022> ↗

[Get rights and content](#) ↗

Under a Creative Commons [license](#) ↗

PDF

Help

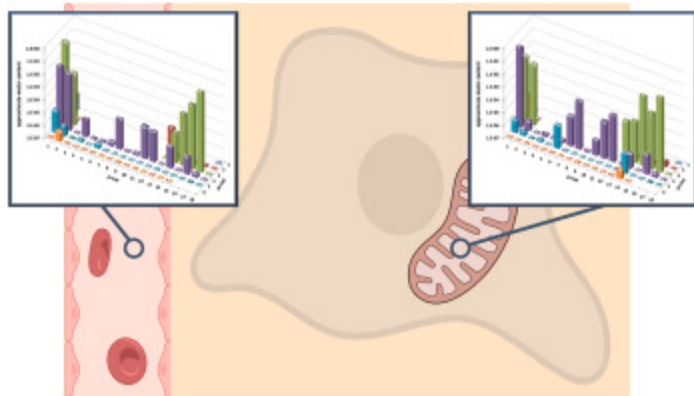
Highlights

- Eleven of the 12 minerals essential for human health have important roles within mitochondrial metabolism.
- Increased oxidative stress within the mitochondria is a common consequence of aberrant mineral homeostasis.
- Low oxidative stress is key for long-lived cell lineages, so optimizing mineral levels is important for cellular potency.
- Tuning up mineral metabolism should strengthen mitochondrial physiology, and thereby improve cellular health.

Abstract

Professor Bruce Ames demonstrated that nutritional recommendations should be adjusted in order to ‘tune-up’ metabolism and reduce mitochondria decay, a hallmark of aging and many disease processes. A major subset of tunable nutrients are the minerals, which despite being integral to every aspect of metabolism are often deficient in the typical Western diet. Mitochondria are particularly rich in minerals, where they function as essential cofactors for mitochondrial physiology and overall cellular health. Yet substantial knowledge gaps remain in our understanding of the form and function of these minerals needed for metabolic harmony. Some of the minerals have known activities in the mitochondria but with incomplete regulatory detail, whereas other minerals have no established mitochondrial function at all. A comprehensive metallome of the mitochondria is needed to fully understand the patterns and relationships of minerals within metabolic processes and cellular development. This brief overview serves to highlight the current progress towards understanding mineral homeostasis in the mitochondria and to encourage more research activity in key areas. Future work may likely reveal that adjusting the amounts of specific nutritional minerals has longevity benefits for human health.

Graphical abstract



[Download: Download high-res image \(261KB\)](#)

[Download: Download full-size image](#)

[<](#) Previous

Next [>](#)

Keywords

Mitochondria; Minerals; Metals; Redox; Differentiation

Abbreviations

MFRN1, mitoferrin 1; MFRN2, mitoferrin 2; mitoBK_{Ca}, mitochondrial calcium-activated potassium channel; mitoK_{ATP}, mitochondrial ATP-regulated potassium channel; mitoKv1.3, mitochondrial voltage-gated Kv1.3 potassium channel; mitoTASK, mitochondrial TASK-3 potassium channel; Moco, molybdenum cofactor; MTM1, manganese trafficking factor for mitochondrial SOD2; NCE, mitochondrial sodium/calcium exchanger; NCX, cellular sodium/calcium exchange; NHE, mitochondrial sodium/proton exchanger; ROS, reactive oxygen species; TCA, tricarboxylic acid cycle

1. Introduction

In a series of papers beginning 20 years ago, Professor Bruce Ames proposed that the recommended intake for essential micronutrients should be reset to promote optimal health rather than just avoiding acute disease – a campaign he called ‘tuning up metabolism’ [[1], [2], [3]]. Central to this proposal is boosting the metabolism of the mitochondria [4], which Professor Ames and others postulated was the fulcrum for the pathological processes that drive aging and senescence [[5], [6], [7], [8]]. Of the 40–50 micronutrients required for human physiology, most can be found within the mitochondria and many have recognized functions within mitochondrial metabolism. There is strong evidence that deficiencies in many of these micronutrients can result in increased production of reactive oxygen species (ROS), redox imbalance, and mitochondrial decay [4,9]. Conversely, supplementation with micronutrients that are needed by the mitochondria has proven very successful in improving health and restoring vitality in many different models [[10], [11], [12], [13], [14]].

The nutritional minerals form a unique subset within the essential micronutrients, and they are integrated into all aspects of systemic metabolism. In the literature on mineral homeostasis, mitochondria are repeatedly referred to as ‘hubs’ for mineral processing – particularly for calcium and iron – given the many cellular pathways utilizing minerals that converge within this organelle. Yet a hub is a device that functions as a passive node within an existing network; instead, mitochondria actively manage, buffer, utilize, and route the minerals in order to maintain homeostatic balance and minimize stress levels throughout the cell. Interruptions in mitochondrial mineral handling are associated with metabolic dysfunction, which if prolonged results in increased cellular stress and consequential disease. Thus a more fitting metaphor for mitochondria is that they function as central

processing units for minerals within the cell. This active control of mineral balance by mitochondria is a key part of an optimized metabolism and cellular health.

The first step in optimizing mineral metabolism is to define all the substrates, reactions, and interactions of the minerals within the cell. It is also essential to understand how metabolic profiles change during development to match physiological needs. Professor Ames addressed this latter point in a more recent article in which he proposed the existence of longevity vitamins that support processes connected to long-term health instead of immediate survival or reproduction, and thus would not be under the same evolutionary pressure to preserve cofactor availability [15]. It is likely that some minerals also fit the definition of longevity micronutrients, but detailed examination of this topic has not yet been reported. In order to understand how some minerals may support long-term health functions during aging and development, it is necessary to characterize all the minerals and determine how they are interconnected, especially within the mitochondria.

Interest in mitochondrial mineral homeostasis continues to grow. There have been several excellent reviews that explore the metallome and the cognate metalloproteome of the mitochondria [[16], [17], [18], [19]], though to date our understanding of mitochondrial mineral homeostasis still remains incomplete. The purpose of this review is to illustrate specific knowledge gaps and highlight details about mitochondrial minerals that deserve more research attention. The minerals are divided into sections based on periodic table assignment that share chemical and physical properties. Mitochondrial regulation of the minerals will be addressed when known, as well as the impact on redox balance and cellular differentiation when mineral balance is altered. A central theme in mitochondrial mineral homeostasis is that imbalances often lead to increased ROS production and redox stress [4]. The term ROS refers to widely different molecular species, including lipid and nitrogenous radicals, that have distinct pharmacodynamic properties and chemical reactivities [20]. The terms redox or oxidative stress refer to conditions when the levels of ROS increase relative to the antioxidant or defense capacity within the biological system. Prolonged redox or oxidative stress can result in mitochondrial decay, altered metabolism, cellular dysfunction, and eventually systemic disease.

2. Mitochondrial metallome

Alkali Metals: The alkali metals are exclusively monovalent cations that exhibit only +1 oxidation state as ions under physiological conditions. These alkali metals do not have inherent redox capability and are mostly found free or in weakly labile complexes with

cellular macromolecules. These two common alkali metals sodium and potassium have major roles in mitochondrial physiology.

Sodium: Sodium is the most abundant cation in the extracellular space, and has a reciprocal relationship with potassium. Circulating levels of sodium in the serum/plasma are between 136 and 145 mM in healthy adults, while the intracellular levels are kept at 8–15 mM [21,22,23]. The potassium-sodium gradient between intracellular and extracellular spaces powers many cellular processes to maintain homeostasis of other metabolites. The concentration of sodium within the resting mitochondria has been reported between 5 and 50 mM [22,24,25]; this wide range likely reflects biological variability in different cell types with different excitation potentials. Excitation can cause substantial increases in mitochondria sodium levels, with 10-fold elevation observed during conditions such as ischemia [22,26,27]. Entry of sodium into the mitochondria mainly occurs through the mitochondrial sodium/calcium exchanger (NCE) in many cell types, with a supporting role from the sodium/proton exchanger (NHE), calcium uniporter, and cellular sodium/calcium exchange (NCX) in reverse mode. The regulation of these transporters is complex, coordinated by mitochondrial membrane potential, oxygen tension, pH, and concentrations of sodium, calcium, magnesium, and protons in the mitochondria and cytosol. The role of sodium within the mitochondria is similar to its role in the cytosol, namely the maintenance of volume, pH, membrane polarization, and macromolecular charge balance. However, mitochondrial sodium also has the unique role as a major driver of calcium currents into the mitochondria, thereby influencing the overall mitochondrial calcium content [23,28]. Additionally, mitochondrial sodium levels reciprocally influence the movement of protons across the inner mitochondrial membrane, thereby contributing to changes in mitochondrial energy charge and redox capacity [23].

Potassium: Potassium is the most abundant cation inside the cell, and has a reciprocal relationship with sodium. Circulating levels of potassium in the serum/plasma are 3.5–5.1 mM in healthy adults, while the intracellular concentration is approximately 100–150 mM [21,29]. The potassium-sodium gradient between intracellular and extracellular spaces powers many cellular processes that maintain homeostasis of other metabolites. The concentration of potassium within the mitochondria has been reported to range between 150 and 180 mM, which is higher than cytosolic potassium and indicates that potassium can be sequestered within the organelle [30,31]. Transport of potassium into the mitochondria occurs through multiple channels, including the ATP-regulated potassium channel (mitoK_{ATP}), calcium-activated potassium channel (mitoBK_{Ca}), voltage-gated Kv1.3 potassium channel (mitoKv1.3), and TASK-3 potassium channel (mitoTASK) [32]. The regulation of these channels is also complex, and driven by changes in mitochondrial membrane

potential, redox balance, oxygen tension, and concentrations of potassium and protons in the mitochondria and cytosol. Potassium has numerous roles in mitochondrial physiology, including maintenance volume, pH, membrane polarization, and macromolecular charge balance [33]. Many proteins require binding of monovalent cations like potassium for optimal function, including mitochondrial pyruvate kinase [34]. Mitochondrial potassium levels also are key to the regulation of mitochondrial volume, membrane potential, calcium homeostasis, and oxidative stress levels, especially through the mitoBK_{Ca} channel that is functionally and structurally coupled to cytochrome c oxidase in the electron transport chain [32,33]. It seems clear that mitochondrial potassium balance is a major mechanism that controls mitochondrial energy charge and redox capacity. Evidence suggests that alterations in mitochondrial potassium currents can play important roles in the pathogenesis of cardiovascular and neurodegenerative diseases [32].

Other Alkali Metals: The other alkali metals are found at trace levels in the body, but have not been shown to be essential minerals for human health. *Lithium:* The average circulating lithium levels (not including lithium-treated patients) in the serum/plasma ranges from 1 to 4 μM in healthy adults [35]. Low levels of lithium in the diet are proposed to have health benefits. Studies specifically removing lithium from the diet of animals resulted in anemia and alterations in glucose, cholesterol, phospholipid, triglyceride, bile acid, and iron homeostasis [36]. Lithium has been shown to enter the mitochondria utilizing the sodium/calcium exchanger within the inner mitochondria membrane [37]. Once in the mitochondria matrix, lithium could inhibit various kinases and ion channels, including mitochondria glycogen synthase kinase [38] and Complex V of the electron transport chain [39]. Lithium may also indirectly promote mitochondrial energy charge by reducing the transcription factor LSD1 [40], which normally inhibits the expression of genes for mitochondrial oxidative phosphorylation [41]. *Rubidium:* The average circulating rubidium levels in the serum/plasma ranges from 0.5 to 7 μM in healthy adults [35]. Rubidium has been proposed to have health benefits, though it is difficult to discriminate specific activities of this metal given its similarity to potassium. Studies specifically removing rubidium from the diet of animal models resulted in decreased growth, fecundity, and life expectancy [36]. Yet while multiple investigations have shown that rubidium enters the mitochondria, this metal is usually studied as a tracer of potassium flux and not for determining any unique roles of rubidium per se [42,43]. Little information on the effects of rubidium on mitochondrial energy charge or redox capacity are known. The remaining members of this elemental group are too rare or unstable to have physiological relevance, so there is no information about content or function within the mitochondria.

Alkali Earth Metals: The alkali earth metals are exclusively divalent cations that exhibit only +2 oxidation state as ions under physiological conditions. These alkali earth metals do not have inherent redox capability and are found both free or in complexes with cellular macromolecules, including proteins, lipid membranes, and nucleic acid polymers. Two common alkali metals calcium and magnesium have major roles in mitochondrial physiology.

Calcium: Calcium is the most abundant metal in the human body, and has a reciprocal relationship with magnesium. Only 1% of whole-body calcium is found in the soft tissues and extracellular fluids, but the circulating levels in serum/plasma are still maintained at 2.1–2.6mM in healthy adults [21]. While the total intracellular concentration of calcium may be similar to circulating levels, the free/labile calcium in the cytosol is held to low nanomolar levels through a high-fidelity calcium sequestering system involving the mitochondria and endoplasmic reticulum [[44], [45], [46]]. Low cytosolic calcium is key to cellular function so that calcium oscillations can act as second messengers for stimuli ranging from proliferation to apoptosis. Calcium plays a critical role in mitochondrial physiology, and is arguably the most well studied of all the metals in this organelle. Resting concentrations for mitochondrial calcium are reported between 100 and 200nM [47], but these levels can quickly rise upon cellular excitation, with some microdomains peaking at 10–20 μ M [45]. Transport of calcium into the mitochondria is complex and depends on coordinate systems including calcium-selective voltage-dependent anion channels across the outer mitochondrial membrane and a calcium uniporter across the inner mitochondrial membrane, with transport through mitochondria-associated membrane microdomains also playing a role [46]. The release of calcium from the mitochondria is also important to overall homeostasis and driven mainly by the sodium-calcium-lithium exchanger, and thus contributing to an entangled relationship between those ions. Calcium has a wide range of functions within the mitochondria. Several enzymes within the tricarboxylic acid cycle (TCA) cycle and electron transport chain require calcium ions for activity, including the rate-limiting isocitrate dehydrogenase [45,48,49]. Calcium also stimulates ATP synthesis and adenylate transport in mitochondria, with a corresponding increase in mitochondrial membrane potential [[50], [51], [52]]. Mitochondrial dynamics are altered by calcium transients in the mitochondria, including altered fission/fusion dynamics and distribution within the cell [[53], [54], [55]]. With sustained elevated calcium, the mitochondrial permeability transition pore is activated, triggering apoptosis [46]. There is extensive literature on the regulation of mitochondria bioenergetics and redox state by calcium [56,57].

Magnesium: Magnesium is the second-most abundant cation within the cell, and has a reciprocal relationship with calcium. Circulating levels in serum/plasma are buffered to 0.7–1.1 mM in healthy adults, with a higher total cell concentration reported between 15 and 25 mM and free/labile concentration in the cytosol estimated between 0.5 and 1 mM [21,58]. Free/labile concentration of magnesium within the mitochondria is estimated between 0.4 and 0.7 mM, but total concentration should be higher when including magnesium ions tightly bound to mitochondrial macromolecules [59,60]. Magnesium has many functions within the cell, but the most prominent is being bound to ATP, as the magnesium-ATP complex is the recognized form of the cofactor required for binding by hundreds of enzymes [61]. Binding of magnesium to membranes, proteins, nucleic acids, and other small organic molecules is also important for structural balance and charge neutralization considerations [62]. Deficiencies in magnesium lead to increased oxidative stress, accelerated senescence, and mitochondrial dysfunction in human cells [[62], [63], [64], [65]]. Magnesium plays a major role in mitochondrial function as well, with a third or more of the total magnesium in a cell being located in this organelle [66]. Mitochondria concentrate magnesium ions especially through Mrs2, a magnesium-selective transporter expressed in the mitochondrial inner membrane [67]. Several enzymes within the TCA cycle and electron transport chain require or are regulated by magnesium, including 2-oxoglutarate dehydrogenase (rate-limiting step of the TCA cycle), hexokinase, phosphofructokinase, pyruvate kinase, and several subunits of the electron transport chain including acting as a direct activator of the mitochondrial Complex V [68,69]. Consequently, disruption of magnesium homeostasis in the mitochondria decreases ATP production, disrupts mitochondrial membrane potential, and increases oxidative stress [70]. Several studies have added to a comprehensive understanding of how magnesium levels regulate mitochondrial transmembrane potential, bioenergetics, and redox state [70,71]. These authors suggest that magnesium may be a central regulator of mitochondrial function and metabolism in mammalian cells.

Other Alkali Earth Metals: The other alkali earth metals are found at trace levels in the body, but have not been shown to be essential minerals for human health. *Barium:* The average circulating barium levels in the serum/plasma ranges from 0.2 to 0.6 μM [35]. The mitochondria were found to be the principal site of barium accumulation when cells were exposed to barium salts [72], but the physiological relevance is unknown. *Strontium:* The average circulating strontium levels in the serum/plasma ranges from 0.3 to 0.5 μM [35]. Strontium is taken up by cells and mitochondria in a manner similar to calcium such that strontium is often used as a tracer to map calcium currents [73], but a unique role for strontium per se has not been reported. Little information on the effects of barium or strontium on mitochondrial energy charge or redox capacity are known. The remaining

members of this elemental group are too rare or unstable to have physiological relevance, so there is no information about content or function within the mitochondria.

Transition (*d*-block) Metals: Most of the transition metals located in the *d*-block section of the periodic table are redox-active and can have a range of oxidation states from +1 to +7 as ions under physiological conditions. The exceptions are the metals from Group 3 (scandium and yttrium) and Group 12 (zinc and cadmium), which only have a single oxidation state under physiological conditions. Group 12 metals have a complete *d* shell, but will still be included here since they are more commonly listed in this elementary category. The *d*-block transition metals are usually not found free, but instead in complexes with cellular macromolecules, especially proteins where the range of oxidation states provides catalytic power for enzymatic activities. Many of these metals have essential roles in mitochondrial physiology.

Iron: Iron is the most abundant transition metal within the body and has interconnected relationships with other trace metals, especially copper and zinc. Circulating iron levels in whole blood are as high as 10mM due to erythrocyte content [35], but serum/plasma iron levels are normally between 9 and 31 μ M in healthy adults [21]. The most common form of iron in mammalian biology is within porphyrin ligands collectively called heme, in which iron ions bind at a ratio of 1:1 (metal:porphyrin). The second-most common form of iron is within iron-sulfur clusters, in which iron ions bind to sulfur ions at different ratios ranging 2:2 to 4:4 (metal:sulfur). Systemically, iron is a component of hundreds of proteins involved in oxygen transport, oxygen sensing, energy production, stress defense, hormone production, and nucleic acid synthesis. In the mitochondria, the concentration of iron is substantial, with studies reporting levels between 0.5 and 1 mM [17,74]. Depending on the cell type, the mitochondria can contain up to 50% of all iron in the cell [75]. Iron enters the mitochondria through the iron-selective transporters known as mitoferrins, specifically MFRN1 (SLC25A37) and MFRN2 (SLC25A28), although there is evidence that other transporters may be involved [75,76]. Mitochondria are the starting and ending location for the heme biosynthetic pathway, and also the major site for iron-sulfur cluster synthesis, so maintaining adequate iron levels in the mitochondria is critical to match the iron cofactors requirements of the cell [77]. Iron cofactors are required by several subunits in the electron transport chain proteins, succinate dehydrogenase, ferrochetalase, and several other mitochondrial proteins. Mitochondrial iron that is not immediately needed for cofactor synthesis can be stored in proteins, including mitochondrial ferritin [78,79]. Other chaperone proteins can bind iron, including frataxin which plays an important role in oxidative phosphorylation and mitochondrial energy production [80]. If not stored properly, iron can interact with other molecules in the mitochondria to promote the formation of ROS

[81]. Extensive literature is available that describes the many roles of iron in mitochondria bioenergetics and redox state [79,82].

Zinc: Zinc is the second-most abundant transition metal within the body and has interconnected relationships with other trace metals, especially iron and copper. Circulating zinc levels in whole blood are approximately 100 μ M due to erythrocyte content [35], but serum/plasma levels are normally between 11 and 18 μ M in healthy adults [21]. Unlike most other essential transition metals, zinc is not redox-active and normally exhibits only one oxidation state [83]. Zinc is a cofactor in numerous proteins, providing catalytic, structural, and/or regulatory activity. In the human proteome, several hundred transcription factors require zinc for proper folding and over 50 enzymes utilize zinc for catalytic functions. Systemically, zinc is involved in immune function, neuronal function, vision, reproduction, bone health, growth, and development [84]. The transport of zinc into the mitochondria occurs mainly through ZnT2 (SLC30A2), a member of the zinc efflux transporter family that moves zinc out of the cytoplasm (and in this case into intracellular organelles) [85]. The mitochondrial concentration of zinc is substantial, with studies reporting levels between 167 and 300 μ M, with some evidence of microdomains with elevated zinc content [17,74]. It is clear that mitochondria can sequester zinc, which serves as a zinc reserve for later cellular need [86]. Mitochondrial zinc that is not immediately needed as protein cofactors can be stored in the protein metallothionein, which although normally thought of in the cytoplasm has also been localized to the mitochondria [87]. Zinc is required as a cofactor for several mitochondrial enzymes and for multiple subunits of the electron transport chain [69]. Additionally, a fraction of zinc-containing superoxide dismutase 1 (normally found in the cytoplasm) localizes to the inner mitochondrial space and provides protection from oxidative stress [88]. Thus, low levels of mitochondrial zinc can promote increased sensitivity to oxidative stress [89]. On the other hand, high zinc levels in the mitochondria have been shown to cause toxicity by inducing loss of mitochondrial membrane potential, which is a known cause of increased ROS production [[90], [91], [92]]. There is additional literature describing the importance of mitochondrial zinc for control of mitochondria bioenergetics and redox state [87,91].

Copper: Copper is another important transition metal within the body and has interconnected relationships with trace metals, especially iron and zinc. Circulating copper in whole blood and serum/plasma are similar to iron with a range of 11–30 μ M. Copper may be concentrated in the mitochondria, with studies reporting levels between 71 and 115 μ M [17,74]. The transport of copper into the mitochondria occurs mainly through what was first known as the mitochondrial phosphate carrier protein (SLC25A3), but more recently shown to also transport copper and required for activity of mitochondrial copper proteins [93].

Copper is utilized as a cofactor in over 50 enzymes within the human proteome, including enzymes involved in energy production, connective tissue formation, iron homeostasis, neurotransmitter synthesis, and myelin formation [94]. Several of these enzymes are localized to the mitochondria, including subunits 1 & 2 of the cytochrome c oxidase and protein deglycase DJ-1, which functions as a sensor for oxidative stress and redox-sensitive chaperone proteins. Other important copper-containing chaperones and transporters include Cox11, Cox17, and cytochrome c oxidase assembly proteins SCO1 and SCO2, all of which have been identified in the mitochondria. Additionally, the copper proteins prion protein PrP and acetylcholinesterase-associated protein CutA have isoforms found in the mitochondria, but their function is unknown. Like other transition metals, excess copper can interact with other molecules in the mitochondria to promote the formation of ROS [95,96]. This is seen in Wilson disease, in which elevated increased mitochondrial copper levels leads directly to mitochondrial ultrastructure and function [97]. Thus maintaining balance in copper levels within the mitochondria is important for mitochondrial bioenergetics and redox balance [98,99].

Manganese: Manganese is a low abundant transition metal with circulating levels in serum/plasma at 8–18nM in healthy adults [21]. Manganese may be concentrated in the mitochondria, with studies reporting levels between 3 and 16 μ M [17,73]. The mechanisms for manganese entry into the mitochondria are not fully understood though, as both MTM1 (manganese trafficking factor for mitochondrial SOD2) and the mitochondrial calcium uniporter appear to be involved [18,100]. Manganese is required as a cofactor in a number of enzymes within the human proteome, including kinases, hydrolases, transferases, and decarboxylases [101]. Several of these enzymes are localized to the mitochondria, including arginase II, pyruvate carboxylase, and superoxide dismutase 2. Arginase is required for the urea cycle. Pyruvate carboxylase is essential for proper carbohydrate metabolism and anaplerotic reactions in the mitochondria. Superoxide dismutase 2 is critical to the protection of mitochondria from oxidative stress, and expression of this enzyme even correlates with lifespan in mammals [102]. Superoxide dismutase 2 and other enzymes systems in the mitochondria are also proposed to control the redox balance for the whole cell and to generate ROS transients that synchronize metabolism to changes in cellular physiology [103]. Like other transition metals, excess manganese can cause mitochondrial toxicity by inactivating oxidative phosphorylation, inducing the loss of mitochondrial membrane potential, and increasing ROS production [101,104,105]. Thus maintaining a balance in manganese levels within the mitochondria is also important for mitochondrial bioenergetics and redox balance [[101], [102], [103]].

Molybdenum: Molybdenum is another low abundant transition metal with circulating levels in serum/plasma at 1–31 nM in healthy adults [21]. The most common form of molybdenum in human biology is within pterin ligands collectively called molybdenum cofactors (Moco), in which molybdenum ions bind at a ratio of 1:1 (metal:pterin) [106]. Moco is required for at least 4 proteins in the human proteome, including xanthine oxidase, aldehyde oxidase, sulfite oxidase, and amidoxime reducing component [107]. The synthesis of Moco starts in the mitochondria, but finishes with metal addition to the pterin structure in the cytoplasm, however it is still unclear how the Moco cofactor is transported back into the mitochondria [18,108]. Mitochondria must take up molybdenum, with estimates of mitochondrial concentration between 1 and 6 μM ¹⁷. Some of this molybdenum must be the Moco form, as isoforms of sulfite oxidase and amidoxime reducing component are localized in the mitochondria and require Moco for function. The mitochondrial amidoxime reducing component protein is involved with nitric oxide metabolism. Molybdenum can also form thiomolybdate compounds, including tetrathiomolybdate, that influence copper metabolism and handling. Both nitric oxide and copper levels are critical to mitochondrial redox balance and function, and disruption of Moco synthesis is known to cause loss of mitochondrial energy charge and oxidative stress [108]. Yet the degree to which molybdenum regulates mitochondrial function and redox state is still under investigation.

Cobalt: The only known form of cobalt in human biology is within corrin ligands collectively called cobalamins or vitamin B12, in which cobalt ions bind at ratio of 1:1 (metal:corrin). The circulating levels in serum/plasma for cobalt are reported as 2–8 nM; interestingly, this is higher than the circulating levels listed for vitamin B12 at 0.1–0.6 nM, suggesting either error in existing measurements or unknown other role for cobalt [21]. Estimates for the mitochondrial concentration of cobalt range from 50 to 90 nM, which if all corrin-bound would represent about 15% of the cellular vitamin B12 [17,109]. There are at least 2 proteins identified in the human proteome that require vitamin B12, including methionine synthase and L-methylmalonyl-coenzyme A mutase, with the latter localized in the mitochondria and required for synthesis of the key metabolite succinyl-CoA and the degradation of specific amino acids and fatty acids [110]. Vitamin B12 is also involved in the synthesis of glutathione, an important antioxidant in the cytosol and mitochondria [111]. It is unclear how the vitamin B12 cofactor is transported or regulated within the mitochondria, and little is known about the pathophysiology of vitamin B12 imbalances in the mitochondria. Yet cobalt as vitamin B12 clearly plays an important role in the mitochondrial metabolism and redox state, so further investigation is needed.

Chromium: The activity of chromium within the human body is not well understood. The best described physiological form of chromium is within an oligopeptide called

chromodulin, in which chromium ions bind at a ratio of 4:1 (metal:peptide) [112]. Circulating levels of chromium in serum/plasma are low and listed as 1–31 nM in healthy adults [21]. Chromodulin was shown to potentiate glucose responses, potentially by increasing the inherent kinase activity within the insulin receptor complex, though this mechanism is not fully elucidated [113,114]. To date, there seem to be no reports of chromodulin activity within the mitochondria. Other activities of chromium have also been proposed including attenuating oxidative stress and inflammatory tone [114]. The cellular regulation and localization of chromium and chromodulin pools are still under investigation, and the role of chromium in mitochondrial function and redox balance is not known.

Other Transition Metals: The other *d*-block transition metals are found at trace levels in the body, but have not been shown to be essential minerals for human health. *Nickel:* The normal circulating nickel levels in the serum/plasma ranges from 2 to 17 nM²¹. Studies removing nickel from the diet of animal models resulted in decreased growth and alterations in glucose, folic acid, vitamin B12, calcium, iron, and zinc homeostasis in multiple mammalian species [36]. Nickel has been directly detected within the mitochondria of human cells when treated with nickel salts [115,116], but specific roles in mitochondrial metabolism are unknown. *Titanium:* The average circulating titanium levels in the serum/plasma ranges from 1.1 to 2.5 μM [35]. Most reports on the biological activity of titanium focus on nanoparticle and not ionic species. One study in mice showed that labeled titanium nanoparticles were taken up by mitochondria in multiple tissues [117], although the physiological relevance is unknown. *Vanadium:* The normal circulating vanadium levels in the serum/plasma ranges from 0.3 to 9 nM²¹. Studies removing vanadium from the diet of animal models resulted in decreased lifespan and alterations in bone and thyroid development in multiple mammalian species [36]. Treatment of multiple animals models with vanadium compounds resulted in vanadium being localized within the mitochondria, indicating that mitochondria can take up this metal but a specific role in mitochondrial metabolism is not known [118,119]. Little information on the effects of nickel, titanium, or vanadium on mitochondrial energy charge or redox capacity are known. The remaining members of this elemental group are too rare or unstable to have physiological relevance, so there is no information about content or function within the mitochondria.

Post-Transition Metals & Metalloids: The post-transition metals and metalloids are a heterogeneous grouping of elements in the periodic table that generally fill the *p*-block section of the periodic table, with some exceptions. These elements are often described as having partially non-metal character and exhibit a mix of single or multiple oxidation state potentials under physiologic conditions. Most of these elements are not often found free,

but instead exist in stable oxides or in complexes with cellular macromolecules. Only the element selenium in this group has been proven to have an essential role in human metabolism. However, two additional metalloids boron and silicon have strong evidence for beneficial roles in physiology.

Selenium: Selenium is the only metalloid with confirmed essentiality for humans. The normal circulating level of selenium in serum/plasma is between 0.6 and 1.8 μM in healthy adults [21]. The mitochondrial levels of selenium have been reported at 1.1–1.8 $\mu\text{g/g}$ dry weight, representing about 25% of total cellular selenium in human liver tissue [120]. The major function of selenium in the cell is to be incorporated into the amino acids selenocysteine and selenomethionine, which are used to synthesize the 25 known selenoproteins in the human proteome [121]. These proteins have roles in redox regulation, protein folding, thyroid hormone function, calcium homeostasis, and antioxidant enzyme systems. Several of these selenoproteins affect or are located to the mitochondria, including thioredoxin reductase 2, selenoprotein O, and glutathione peroxidase 4 [122]. In particular, thioredoxin reductase and glutathione peroxidase play a key role in protecting mitochondria from the harmful effects of ROS, utilizing the endogenous antioxidants thioredoxin and glutathione [123]. Small molecular weight selenium compounds may also directly function as antioxidants and signaling molecules within the circulation and the cells, but little information is available for activity within the mitochondria [124]. It is clear that selenium plays an important role in supporting mitochondrial function and reduction of redox stress, but further investigation is needed to reveal the detail.

Boron: Boron is an essential metalloid for plants, but to date no biological role has been established for humans. Still, the evidence for boron as a beneficial mineral for humans is strong, and studies suggest that boron might have a role in hormonal regulation and in the metabolism of carbohydrates and lipids [[125], [126], [127]]. Additionally, boron deprivation in multiple animal models resulted in altered homeostasis of calcium, phosphorus, magnesium, potassium [36]. The circulating levels of boron in plasma/serum are reported between 3 and 11 μM [35,128]. Boron supplementation in human subjects decreased serum glucose, creatinine, and calcitonin, while it increased serum triglycerides, ceruloplasmin, and erythrocyte superoxide dismutase [36]. Boron administration had positive effects on mitochondrial membrane potential and function in multiple species, but entry into mitochondria was not confirmed [129,130]. The available evidence suggest that mitochondria may benefit from the availability of boron, which may promote metabolism and reduce redox stress.

Silicon: There is also growing evidence that the metalloid silicon may be beneficial for human physiology, especially for biomineralized tissue. The circulating levels of silicon in the serum/plasma are reported between 89 and 356 μM ³⁵. Studies removing silicon from the diet of multiple animal models resulted in altered bone structure, amino acid balance, and homeostasis of calcium, copper, magnesium, manganese, and phosphorus [36]. Possible roles for silicon include a direct structural presence within collagen or cartilage effects on expression and calcification of connective tissues. Multiple forms of silicon can enter the mitochondria [131], but the mechanisms of entry, concentrations, and effects are not resolved.

Other Post-Transition Metals & Metalloids: The other post-transition metals and metalloids are found at trace levels in the body, but have not been shown to be essential minerals for human health. *Aluminum:* The circulating aluminum levels in the serum/plasma range from 0.2 to 0.6 μM ²¹. A few studies removing aluminum from the diet of animal models resulted in decreased growth and life expectancy [36]. Several studies show that high levels of aluminum can be toxic to mitochondria by increasing ROS production and decreasing mitochondrial membrane potential [132], but little work is available on any role for aluminum at normal exposure levels. *Antimony:* The circulating antimony levels in the serum/plasma range from 3 to 6 nM²¹, but concentrations in the mitochondria are not known. Antimony intoxication in rats did result in mitochondria swelling, suggesting entry into the organelle [133]. *Arsenic:* The circulating arsenic levels in the serum/plasma range from 0.02 to 0.2 μM ²¹. Arsenic is normally thought of as a toxic metalloid, but some reports claim that low levels of arsenic have health benefits. Studies removing arsenic from the diet of animal models resulted in decreased growth and changes in the balance of polyamines, taurine, glutathione, and zinc [36]. Prolonged arsenic deprivation in goats resulted in mitochondrial abnormalities in heart tissue, but mitochondrial concentrations were not measured [134]. Arsenic intoxication results in mitochondria depolarization and permeability transition, but entry into mitochondria was not confirmed [135]. *Bismuth:* The circulating bismuth levels in the serum/plasma range from 0.5 to 17 nM and is generally regarded as non-toxic despite being a heavy metal [35]. Humans consume bismuth in common antacids and other medicinal preparations. Treatment of cells with bismuth compounds resulted in localization of the metal within the mitochondria in one study [136]. *Germanium:* The circulating germanium levels in the serum/plasma are estimated to range within 2.6–4.0 μM [35,137]. A few reports have claimed that germanium stimulates the immune system [138], but corroborating evidence is needed. Germanium intoxication did result in mitochondria abnormalities including decreased cytochrome c oxidase activity, but entry into mitochondria was not confirmed [139]. *Indium:* The circulating indium levels in the serum/plasma are not widely known, but are estimated to range within 9–90 nM [140,

141]. Treatment of rats with indium compounds did result in indium uptake into cells and into the mitochondria [142,143]. *Lead*: Lead is normally thought of as a toxic metalloid, but some reports claim that low levels of lead have health benefits. Studies removing lead from the diet of animal models resulted in anemia and alterations in glucose, cholesterol, triglycerides, and iron homeostasis [36]. The circulating lead levels in the serum/plasma range from 0.06 to 0.9 μM ³⁵, but no beneficial effects in humans are known. Lead intoxication inhibits the mitochondrial enzyme ferrochelatase and eventually causes mitophagy [144]. *Tin*: The circulating tin levels in the serum/plasma range from 3 to 5 nM²¹. Organic tin compounds are widely studied as anti-cancer compounds that promote mitochondria-dependent apoptosis [145], but the effects of physiological levels of tin on mitochondria are unknown. Little information on the effects of these other post-transition and metalloid elements on mitochondrial energy charge or redox capacity are known. The remaining members of this elemental group are too rare or unstable to have physiological relevance, so there is no information about content or function within the mitochondria.

3. Role of mitochondrial minerals in differentiation

Numerous studies have demonstrated how aberrant mineral handling in the mitochondria can cause metabolic imbalance and result in disease [23,46,75,82,146], but there are fewer reports on the roles that mitochondrial minerals have in the normal development of the cell. This is an important topic since mitochondria have been recognized as a key driver of cellular differentiation and programming [147,148]. Earlier studies revealed that mitochondria from pluripotent cells generally have reduced mass, immature ultrastructure, and low metabolic activity [149,150]. This intracellular configuration is now known to favor glycolysis and reduce ROS released during oxidative phosphorylation. Reductions in redox burden and oxidative stress are key to promoting cellular longevity required for long-lived pluripotent cell lineages. In addition to decreased ROS levels, stem cells have elevated antioxidant defenses, including increased superoxide dismutase, catalase, and glutathione peroxidase activity [151]. Also, the levels of the key antioxidant glutathione can be 3–4 fold higher in pluripotent cell types compared to differentiated somatic cells [152]. Once pluripotent cells are stimulated to differentiate, the up-regulation of mitochondrial bioenergetics and metabolism seems to be an early and necessary step. In fact, elevated ROS production by mitochondria is known to be a trigger stimulus for differentiation and reduced regenerative potential in human mesenchymal stem cells [153].

Given the changes to mitochondrial physiology during pluripotent cell maintenance or differentiation, it can be inferred that adjustments to the mitochondrial metallome are also needed. Calcium was reported to be reduced within the mitochondria of resting pluripotent

cells [154], which might be expected since calcium stimulates mitochondrial bioenergetics and increases ROS production. Other than calcium, only a few studies have evaluated mitochondrial mineral balance in the context of cellular differentiation. *Alkali Metals*: Sodium currents directly increase mitochondrial calcium content, so it is assumed that mitochondrial sodium levels are also low in pluripotent cells, but this has yet to be shown directly. Interestingly, lithium was shown to increase mitochondrial respiration in human neural precursor cells [155], yet the physiological relevance is not known. *Alkali Earth Metals*: Magnesium levels were shown to regulate neural stem cell proliferation *ex vivo*, although changes in mitochondrial magnesium content were not reported [156]. *Transition Metals*: Zinc levels were shown to regulate neural stem cell proliferation *ex vivo*, with increased ROS during zinc deficient conditions [[157], [158], [159]]. Also, exogenous zinc treatment of mouse embryonic stem cells resulted in increased expression of genes that maintain pluripotency and downregulated several genes involved in differentiation [160]. Another report showed that copper levels were low in resting hematopoietic stem cells, perhaps since copper can stimulate mitochondria metabolism [96,161]. Artificially increasing copper levels in hematopoietic progenitor cells resulted in accelerated differentiation [162]. Elevated manganese was shown to be toxic to neural stem cells [163], but that is true for most cell types; no specific role is known in pluripotent cells. *Post-Transition Metals & Metalloids*: Selenium content is known to influence cancer stem cell development due to its effects on mitochondrial redox balance [164], but we are unaware of studies investigating the role of selenium in maintenance of pluripotency or differentiation. Given the strong regulatory influence that minerals have on mitochondrial metabolism and redox balance, it is likely that other important relationships between minerals and cellular differentiation will be identified. A better understanding of the roles of mitochondrial metals will be important as we move closer to widespread application of pluripotent cells in clinical therapies.

4. Final comments

This brief survey reveals an incomplete understanding of content and function for the minerals within the mitochondria (Table 1). For essential minerals such as calcium and iron, there is a substantial amount of information on their metabolic roles and impact on mitochondrial physiology. But for other essential minerals such as chromium and selenium, the content and function of these minerals within the mitochondria is incomplete. For minerals such as boron and silicon, the details of their role with human metabolism are still not elucidated, so there is little to no information on mitochondrial context. Clearly there is much work to be done.

Table 1. Metals and metalloids known in the mammalian mitochondria.

Element	Serum/plasma concentration	Entry into in mitochondria	Mitochondrial concentration	Sequestration	Metabolic role(s)
Confirmed essential:					
Ca	2.1–2.6mM	yes	0.1–20 μ M	yes	yes
Co	2–8nM	yes	50–90nM	yes	yes
Cr	1–31 nM	–	–	–	–
Cu	11–30 μ M	yes	71–115 μ M	yes	yes
Fe	9–31 μ M	yes	0.5–1.1 mM	yes	yes
K	3.5–5.1 mM	yes	150–180mM	yes	yes
Mg	0.7–1.1 mM	yes	0.4–0.7mM	yes	yes
Mn	8–18 nM	yes	3–16 μ M	yes	yes
Mo	1–31 nM	yes	1–6 μ M	yes	yes
Na	136–145mM	yes	5–50mM	yes	yes
Se	0.6–1.8 μ M	yes	–	–	yes
Zn	11–18 μ M	yes	167–300 μ M	yes	yes
Possibly beneficial:					
Al	200–600nM	yes	–	–	–
As	20–200nM	–	–	–	–
B	3–11 μ M	–	–	–	–
Ba	200–600nM	yes	–	–	–
Bi	0.5–17nM	yes	–	–	–
Ge	2.6–4.0 μ M	–	–	–	–
In	9–90nM	yes	–	–	–
Li	1–4 μ M	yes	–	–	–
Ni	2–17nM	yes	–	–	–

Element	Serum/plasma concentration	Entry into in mitochondria	Mitochondrial concentration	Sequestration	Metabolic role(s)
Pb	60–900nM	yes	–	–	–
Rb	0.5–7μM	yes	–	–	–
Sb	3–6nM	–	–	–	–
Si	89–356μM	yes	–	–	–
Sn	3–5nM	–	–	–	–
Sr	300–500nM	yes	–	–	–
Ti	1.1–2.5μM	yes	–	–	–
V	0.3–9nM	yes	–	–	–

Notes: See text for references. Many of the concentrations for mitochondria derive from disparate cell/tissue types and from limited number of references, so values are best viewed as estimates for mitochondrial metal and metalloid content. Table does not include data from reports listing elemental content per mitochondrial protein content or per dry weight. The symbol ‘–’ indicates that a concentration or value could not be found in the available literature.

Many questions remain about mitochondrial mineral homeostasis. First, how are mitochondrial mineral pools sensed and maintained? Most proteins destined for the mitochondria are imported in as unfolded peptides, then assembled and metal-bound once inside the mitochondrial matrix. This presumes that an adequate amount of the required minerals are already maintained inside the mitochondrial matrix, although how that pool is maintained is not yet resolved^{16,18,19}. Second, once the metal content of the mitochondria is fully identified, how will the binding proteins and cognate small molecules be determined? Development of new technologies to effectively measure labile metals and weak metal-protein binding dynamics is becoming more accessible [17,18]. Also, there has been progress in the development of new bioinformatics tools that can predict mineral binding sites in metalloproteins or metallochaperones [165]. Thirdly, mineral binding sites are never absolutely selective, so substitute minerals do sometimes replace primary minerals in binding sites, which could change the activity or behavior of the metalloproteins or metallochaperones. Determining this ‘second dimension’ of mineral complexes will be important to appreciate mineral activity during changing physiological conditions. Finally, how do the mineral profiles of the mitochondria change with differentiation and developmental stages? Many studies on mitochondrial minerals focus at a single timepoint,

but it is essential to monitor how mineral homeostasis in the mitochondria changes over time and during development to better optimize mineral requirements to metabolic need.

Once we have a complete catalog for the metallome of the mitochondria, more practical approaches to optimizing nutrition inputs become available. We can then better determine if there are longevity minerals that might provide countermeasures against the pathophysiologic processes of aging and senescence.

OSM

No.

Funding sources

This manuscript was not supported with external funding

Declaration of competing interest

David W. Killilea, PhD no conflicts of interest.




Alison N. Killilea, PhD no conflicts of interest.

Acknowledgements

In response to questions about his involvement in so many different fields, Professor Bruce Ames often responded that he was ‘incurably distractable.’ This is polite modesty from a polymath who is always curious, deeply thoughtful, and has exceptional forest-for-the-trees vision. Bruce's trainees and colleagues continue to be inspired by his success across such a wide spectrum of biochemistry and nutrition fields. We greatly appreciate Bruce's investment in D.W.K. as a post-doctoral fellow in his laboratory (2000–2003), and continued collaboration together at Children's Hospital Oakland Research Institute. D.W.K. acknowledges his fellow collaborators in projects on minerals and mitochondria with Drs. Patrick Grant, Jiankang Liu, Mark Shigenaga, and Patrick Walter – all former Ames Lab members as well. We also acknowledge the writings of Dr. Forrest Nielsen which inspired us to learn about the lesser-known nutritional minerals.

[Special issue articles](#) [Recommended articles](#)

References

- [1] B.N. Ames, P. Wakimoto
Are vitamin and mineral deficiencies a major cancer risk?
Nat. Rev. Cancer, 2 (2002), pp. 694-704
[Crossref ↗](#) [View in Scopus ↗](#) [Google Scholar ↗](#)
- [2] B.N. Ames
The metabolic tune-up: metabolic harmony and disease prevention
J. Nutr., 133 (2003), pp. 1544S-1548S
 [View PDF](#) [View article](#) [Google Scholar ↗](#)
- [3] B.N. Ames
A role for supplements in optimizing health: the metabolic tune-up
Arch. Biochem. Biophys., 423 (2004), pp. 227-234
 [View PDF](#) [View article](#) [View in Scopus ↗](#) [Google Scholar ↗](#)
- [4] B.N. Ames, H. Atamna, D.W. Killilea
Mineral and vitamin deficiencies can accelerate the mitochondrial decay of aging
Mol. Aspect. Med., 26 (2005), pp. 363-378
 [View PDF](#) [View article](#) [View in Scopus ↗](#) [Google Scholar ↗](#)
- [5] D. Harman
The biologic clock: the mitochondria?
J. Am. Geriatr. Soc., 20 (1972), pp. 145-147
[Crossref ↗](#) [View in Scopus ↗](#) [Google Scholar ↗](#)
- [6] M.K. Shigenaga, T.M. Hagen, B.N. Ames
Oxidative damage and mitochondrial decay in aging
Proc. Natl. Acad. Sci. Unit. States Am., 91 (1994), pp. 10771-10778
[Crossref ↗](#) [View in Scopus ↗](#) [Google Scholar ↗](#)
- [7] T.M. Hagen, *et al.*
Mitochondrial decay in hepatocytes from old rats: membrane potential declines, heterogeneity and oxidants increase
Proc. Natl. Acad. Sci. Unit. States Am., 94 (1997), pp. 3064-3069
[View in Scopus ↗](#) [Google Scholar ↗](#)
- [8] K.B. Beckman, B.N. Ames
The free radical theory of aging matures
Physiol. Rev., 78 (1998), pp. 547-581

[Crossref ↗](#) [View in Scopus ↗](#) [Google Scholar ↗](#)

- [9] B.N. Ames
Low micronutrient intake may accelerate the degenerative diseases of aging through allocation of scarce micronutrients by triage
Proc. Natl. Acad. Sci. Unit. States Am., 103 (2006), pp. 17589-17594

[Crossref ↗](#) [View in Scopus ↗](#) [Google Scholar ↗](#)

- [10] T.M. Hagen, *et al.*
Feeding acetyl-L-carnitine and lipoic acid to old rats significantly improves metabolic function while decreasing oxidative stress
Proc. Natl. Acad. Sci. Unit. States Am., 99 (2002), pp. 1870-1875

[View in Scopus ↗](#) [Google Scholar ↗](#)

- [11] J. Liu, H. Atamna, H. Kuratsune, B.N. Ames
Delaying brain mitochondrial decay and aging with mitochondrial antioxidants and metabolites
Ann. N. Y. Acad. Sci., 959 (2002), pp. 133-166

[Crossref ↗](#) [View in Scopus ↗](#) [Google Scholar ↗](#)

- [12] J. Liu, D.W. Killilea, B.N. Ames
Age-associated mitochondrial oxidative decay: improvement of carnitine acetyltransferase substrate-binding affinity and activity in brain by feeding old rats acetyl-L- carnitine and/or R- -lipoic acid
Proc. Natl. Acad. Sci. Unit. States Am., 99 (2002), pp. 1876-1881

[View in Scopus ↗](#) [Google Scholar ↗](#)

- [13] J. Liu, *et al.*
Memory loss in old rats is associated with brain mitochondrial decay and RNA/DNA oxidation: partial reversal by feeding acetyl-L-carnitine and/or R- -lipoic acid
Proc. Natl. Acad. Sci. Unit. States Am., 99 (2002), pp. 2356-2361

[View in Scopus ↗](#) [Google Scholar ↗](#)

- [14] E. Wesselink, W.A.C. Koekkoek, S. Grefte, R.F. Witkamp, A.R.H. van Zanten
Feeding mitochondria: potential role of nutritional components to improve critical illness convalescence
Clin. Nutr., 38 (2019), pp. 982-995

 [View PDF](#) [View article](#) [View in Scopus ↗](#) [Google Scholar ↗](#)

- [15] B.N. Ames
Prolonging healthy aging: longevity vitamins and proteins
Proc. Natl. Acad. Sci. Unit. States Am., 115 (2018), pp. 10836-10844
[Crossref ↗](#) [View in Scopus ↗](#) [Google Scholar ↗](#)
- [16] A. Atkinson, D.R. Winge
Metal acquisition and availability in the mitochondria
Chem. Rev., 109 (2009), pp. 4708-4721
[Crossref ↗](#) [View in Scopus ↗](#) [Google Scholar ↗](#)
- [17] S.P. McCormick, M.J. Moore, P.A. Lindahl
Detection of labile low-molecular-mass transition metal complexes in mitochondria
Biochemistry, 54 (2015), pp. 3442-3453
[Crossref ↗](#) [View in Scopus ↗](#) [Google Scholar ↗](#)
- [18] P.A. Lindahl, M.J. Moore
Labile low-molecular-mass metal complexes in mitochondria: trials and tribulations of a burgeoning field
Biochemistry, 55 (2016), pp. 4140-4153
[Crossref ↗](#) [View in Scopus ↗](#) [Google Scholar ↗](#)
- [19] X. Hu, Y.-M. Go, D.P. Jones
Omics integration for mitochondria systems biology
Antioxidants Redox Signal., 32 (2020), pp. 853-872
[Crossref ↗](#) [View in Scopus ↗](#) [Google Scholar ↗](#)
- [20] Y.R. Li, M. Trush
Defining ROS in biology and medicine
React. Oxyg. Species, 1 (2016)
[Google Scholar ↗](#)
- [21] Tietz, N. & Wu, A. *Tietz Clinical Guide to Laboratory Tests* - Elsevier eBook on VitalSource, fourth ed..
[Google Scholar ↗](#)
- [22] M.M. Pike, M. Kitakaze, E. Marban
²³Na-NMR measurements of intracellular sodium in intact perfused ferret hearts during ischemia and reperfusion
Am. J. Physiol. Heart Circ. Physiol., 259 (1990), pp. H1767-H1773

[Crossref ↗](#) [View in Scopus ↗](#) [Google Scholar ↗](#)

- [23] E. Murphy, D.A. Eisner
Regulation of intracellular and mitochondrial sodium in health and disease
Circ. Res., 104 (2009), pp. 292-303

[View in Scopus ↗](#) [Google Scholar ↗](#)

- [24] P. Donoso, J.G. Mill, S.C. O'Neill, D.A. Eisner
Fluorescence measurements of cytoplasmic and mitochondrial sodium concentration in rat ventricular myocytes
J. Physiol., 448 (1992), pp. 493-509

[Crossref ↗](#) [View in Scopus ↗](#) [Google Scholar ↗](#)

- [25] S. Baron, *et al.*
Role of mitochondrial Na⁺ concentration, measured by CoroNa red, in the protection of metabolically inhibited MDCK cells
J. Am. Soc. Nephrol., 16 (2005), pp. 3490-3497

[View in Scopus ↗](#) [Google Scholar ↗](#)

- [26] E. Murphy, M. Perlman, R.E. London, C. Steenbergen
Amiloride delays the ischemia-induced rise in cytosolic free calcium
Circ. Res., 68 (1991), pp. 1250-1258

[View in Scopus ↗](#) [Google Scholar ↗](#)

- [27] S.E. Anderson, C.Z. Dickinson, H. Liu, P.M. Cala
Effects of Na-K-2Cl cotransport inhibition on myocardial Na and Ca during ischemia and reperfusion
Am. J. Physiol. Cell Physiol., 270 (1996), pp. C608-C618





[View in Scopus ↗](#) [Google Scholar ↗](#)

- [28] M. Sedova, L.A. Blatter
Intracellular sodium modulates mitochondrial calcium signaling in vascular endothelial cells
J. Biol. Chem., 275 (2000), pp. 35402-35407

 [View PDF](#) [View article](#) [View in Scopus ↗](#) [Google Scholar ↗](#)


- [29] W. Walz
Mechanism of rapid K⁺-induced swelling of mouse astrocytes
Neurosci. Lett., 135 (1992), pp. 243-246

 [View PDF](#) [View article](#) [View in Scopus ↗](#) [Google Scholar ↗](#)

- [30] K.D. Garlid
Cation transport in mitochondria – the potassium cycle
Biochim. Biophys. Acta BBA - Bioenerg., 1275 (1996), pp. 123-126
 [View PDF](#) [View article](#) [View in Scopus ↗](#) [Google Scholar ↗](#)
- [31] M.G. Kozoriz, J. Church, M.A. Ozog, C.C. Naus, C. Krebs
Temporary sequestration of potassium by mitochondria in astrocytes
J. Biol. Chem., 285 (2010), pp. 31107-31119
 [View PDF](#) [View article](#) [View in Scopus ↗](#) [Google Scholar ↗](#)
- [32] M. Laskowski, *et al.*
What do we not know about mitochondrial potassium channels?
Biochim. Biophys. Acta BBA - Bioenerg., 1857 (2016), pp. 1247-1257
 [View PDF](#) [View article](#) [View in Scopus ↗](#) [Google Scholar ↗](#)
- [33] K.D. Garlid, P. Paucek
Mitochondrial potassium transport: the K⁺ cycle
Biochim. Biophys. Acta BBA - Bioenerg., 1606 (2003), pp. 23-41
 [View PDF](#) [View article](#) [View in Scopus ↗](#) [Google Scholar ↗](#)
- [34] T.M. Larsen, L.T. Laughlin, H.M. Holden, I. Rayment, G.H. Reed
Structure of rabbit muscle pyruvate kinase complexed with Mn²⁺, K⁺, and pyruvate
Biochemistry, 33 (1994), pp. 6301-6309
[Crossref ↗](#) [View in Scopus ↗](#) [Google Scholar ↗](#)
- [35] G.V. Iyengar, W.E. Kollmer, H.J.M. Bowen, H.J.M. Bowen
The Elemental Composition of Human Tissues and Body Fluids: a Compilation of Values for Adults
Verlag Chemie (1978)
[Google Scholar ↗](#)
- [36] F.H. Nielsen
Ultratrace elements in nutrition: current knowledge and speculation
J. Trace Elem. Exp. Med., 11 (1998), pp. 251-274
[View in Scopus ↗](#) [Google Scholar ↗](#)
- [37] S. Roy, K. Dey, M. Hershinkel, E. Ohana, I. Sekler
Identification of residues that control Li⁺ versus Na⁺ dependent Ca²⁺ exchange at the transport site of the mitochondrial NCLX

Biochim. Biophys. Acta BBA - Mol. Cell Res., 1864 (2017), pp. 997-1008

 [View PDF](#) [View article](#) [View in Scopus ↗](#) [Google Scholar ↗](#)

- [38] G.N. Bijur, R.S. Jope
Glycogen synthase kinase-3 β is highly activated in nuclei and mitochondria
Neuroreport, 14 (2003), pp. 2415-2419
[View in Scopus ↗](#) [Google Scholar ↗](#)
- [39] J. Rizak
The inhibition of ATP production by lithium: a preliminary study in whole mitochondria from rat brain and a putative Model for Bipolar Disorder, 2 (2014), p. 1014
[Google Scholar ↗](#)
- [40] G. McColl, *et al.*
Pharmacogenetic analysis of lithium-induced delayed aging in *Caenorhabditis elegans*
J. Biol. Chem., 283 (2008), pp. 350-357
 [View PDF](#) [View article](#) [View in Scopus ↗](#) [Google Scholar ↗](#)
- [41] A. Sakamoto, *et al.*
Lysine demethylase LSD1 coordinates glycolytic and mitochondrial metabolism in hepatocellular carcinoma cells
Cancer Res., 75 (2015), pp. 1445-1456
[View in Scopus ↗](#) [Google Scholar ↗](#)
- [42] E.J. Lehning, C.L. Gaughan, J. Eichberg, R.M. LoPachin
Rubidium uptake and accumulation in peripheral myelinated internodal axons and Schwann cells
J. Neurochem., 69 (2002), pp. 968-977
[Google Scholar ↗](#)
- [43] M.L.H. Gruwel, B. Kuzio, R. Deslauriers, V.V. Kupriyanov
Measurements of mitochondrial K⁺ fluxes in whole rat hearts using ⁸⁷Rb-NMR
Am. J. Physiol. Cell Physiol., 276 (1999), pp. C193-C200
[View in Scopus ↗](#) [Google Scholar ↗](#)
- [44] J.H. Schreur, *et al.*
Cytosolic and mitochondrial [Ca²⁺] in whole hearts using indo-1 acetoxymethyl ester: effects of high extracellular Ca²⁺

Biophys. J., 70 (1996), pp. 2571-2580

 [View PDF](#) [View article](#) [View in Scopus ↗](#) [Google Scholar ↗](#)

[45] L. Contreras, I. Drago, E. Zampese, T. Pozzan

Mitochondria: the calcium connection

Biochim. Biophys. Acta BBA - Bioenerg., 1797 (2010), pp. 607-618

 [View PDF](#) [View article](#) [View in Scopus ↗](#) [Google Scholar ↗](#)

[46] S. Romero-Garcia, H. Prado-Garcia

Mitochondrial calcium: transport and modulation of cellular processes in homeostasis and cancer (Review)

Int. J. Oncol. (2019), [10.3892/ijo.2019.4696 ↗](#)

[Google Scholar ↗](#)

[47] A.A. Gerencser, V. Adam-Vizi

Mitochondrial Ca²⁺ dynamics reveals limited intramitochondrial Ca²⁺ diffusion

Biophys. J., 88 (2005), pp. 698-714

 [View PDF](#) [View article](#) [View in Scopus ↗](#) [Google Scholar ↗](#)

[48] J. Satrústegui, B. Pardo, A. del Arco

Mitochondrial transporters as novel targets for intracellular calcium signaling

Physiol. Rev., 87 (2007), pp. 29-67

[Crossref ↗](#) [View in Scopus ↗](#) [Google Scholar ↗](#)

[49] R.M. Denton

Regulation of mitochondrial dehydrogenases by calcium ions

Biochim. Biophys. Acta BBA - Bioenerg., 1787 (2009), pp. 1309-1316

 [View PDF](#) [View article](#) [View in Scopus ↗](#) [Google Scholar ↗](#)

[50] G.A. Rutter, *et al.*

Subcellular imaging of intramitochondrial Ca²⁺ with recombinant targeted aequorin: significance for the regulation of pyruvate dehydrogenase activity

Proc. Natl. Acad. Sci. Unit. States Am., 93 (1996), pp. 5489-5494

[View in Scopus ↗](#) [Google Scholar ↗](#)

[51] H.J. Kennedy, *et al.*

Glucose generates sub-plasma membrane ATP microdomains in single islet β -cells

J. Biol. Chem., 274 (1999), pp. 13281-13291

 [View PDF](#) [View article](#) [View in Scopus ↗](#) [Google Scholar ↗](#)

- [52] P.R. Territo, V.K. Mootha, S.A. French, R.S. Balaban
Ca²⁺ activation of heart mitochondrial oxidative phosphorylation: role of the F₀/F₁-ATPase

Am. J. Physiol. Cell Physiol., 278 (2000), pp. C423-C435

[View in Scopus ↗](#) [Google Scholar ↗](#)

- [53] X. Wang, T.L. Schwarz
The mechanism of Ca²⁺-dependent regulation of kinesin-mediated mitochondrial motility

Cell, 136 (2009), pp. 163-174

 [View PDF](#) [View article](#) [View in Scopus ↗](#) [Google Scholar ↗](#)

- [54] W. Ji, A.L. Hatch, R.A. Merrill, S. Strack, H.N. Higgs
Actin filaments target the oligomeric maturation of the dynamin GTPase Drp1 to mitochondrial fission sites

Elife, 4 (2015), Article e11553

[View in Scopus ↗](#) [Google Scholar ↗](#)

- [55] Y. Ohshima, *et al.*
Disrupting mitochondrial Ca²⁺ homeostasis causes tumor-selective TRAIL sensitization through mitochondrial network abnormalities

Int. J. Oncol., 51 (2017), pp. 1146-1158

[Crossref ↗](#) [View in Scopus ↗](#) [Google Scholar ↗](#)

- [56] C. Delierneux, *et al.*
Mitochondrial calcium regulation of redox signaling in cancer

Cells, 9 (2020), p. 432

[Crossref ↗](#) [Google Scholar ↗](#)

- [57] C.S. Gibhardt, D. Ezeriņa, H.-M. Sung, J. Messens, I. Bogeski
Redox regulation of the mitochondrial calcium transport machinery

Curr. Opin. Physiol., 17 (2020), pp. 138-148

 [View PDF](#) [View article](#) [View in Scopus ↗](#) [Google Scholar ↗](#)

- [58] S.L. Volpe
Magnesium in disease prevention and overall health

Adv. Nutr., 4 (2013), pp. 378S-383S

 [View PDF](#) [View article](#) [Crossref ↗](#) [Google Scholar ↗](#)

- [59] D.W. Jung, E. Panzeter, K. Baysal, G.P. Brierley
On the relationship between matrix free Mg²⁺ concentration and total Mg²⁺ in heart mitochondria

Biochim. Biophys. Acta BBA - Bioenerg., 1320 (1997), pp. 310-320

 [View PDF](#) [View article](#) [View in Scopus ↗](#) [Google Scholar ↗](#)

- [60] B.E. Corkey, J. Duszynski, T.L. Rich, B. Matschinsky, J.R. Williamson
Regulation of free and bound magnesium in rat hepatocytes and isolated mitochondria

J. Biol. Chem., 261 (1986), pp. 2567-2574

 [View PDF](#) [View article](#) [View in Scopus ↗](#) [Google Scholar ↗](#)

- [61] Dietary Reference Intakes for Calcium, Phosphorus, Magnesium, Vitamin D, and Fluoride, National Academies Press (1997), p. 5776, [10.17226/5776 ↗](#)

[Google Scholar ↗](#)

- [62] F. Wolf
Cell physiology of magnesium

Mol. Aspect. Med., 24 (2003), pp. 11-26

 [View PDF](#) [View article](#) [View in Scopus ↗](#) [Google Scholar ↗](#)

- [63] D.W. Killilea, B.N. Ames
Magnesium deficiency accelerates cellular senescence in cultured human fibroblasts

Proc. Natl. Acad. Sci. Unit. States Am., 105 (2008), pp. 5768-5773

[Crossref ↗](#) [View in Scopus ↗](#) [Google Scholar ↗](#)

- [64] B.F. Dickens, W.B. Weglicki, Y.-S. Li, I.T. Mak
Magnesium deficiency in vitro enhances free radical-induced intracellular oxidation and cytotoxicity in endothelial cells

FEBS Lett., 311 (1992), pp. 187-191

 [View PDF](#) [View article](#) [Crossref ↗](#) [View in Scopus ↗](#) [Google Scholar ↗](#)

- [65] E. Rock, *et al.*
Dietary magnesium deficiency in rats enhances free radical production in skeletal muscle

J. Nutr., 125 (1995), pp. 1205-1210

 [View PDF](#) [View article](#) [View in Scopus ↗](#) [Google Scholar ↗](#)

- [66] A. Romani, C. Marfella, A. Scarpa
Cell magnesium transport and homeostasis: role of intracellular compartments
Miner. Electrolyte Metab., 19 (1993), pp. 282-289
[View in Scopus ↗](#) [Google Scholar ↗](#)
- [67] G. Zsurka, J. Gregáň, R.J. Schweyen
The human mitochondrial Mrs2 protein functionally substitutes for its yeast homologue, A candidate magnesium transporter
Genomics, 72 (2001), pp. 158-168
 [View PDF](#) [View article](#) [View in Scopus ↗](#) [Google Scholar ↗](#)
- [68] L. Garfinkel, D. Garfinkel
Magnesium regulation of the glycolytic pathway and the enzymes involved
Magnesium, 4 (1985), pp. 60-72
[View in Scopus ↗](#) [Google Scholar ↗](#)
- [69] T. Soulimane, G. Buse
Integral cytochrome-c oxidase. Preparation and progress towards a three-dimensional crystallization
Eur. J. Biochem., 227 (1995), pp. 88-595
[Google Scholar ↗](#)
- [70] R. Yamanaka, *et al.*
Mitochondrial Mg²⁺ homeostasis decides cellular energy metabolism and vulnerability to stress
Sci. Rep., 6 (2016), p. 30027
[View in Scopus ↗](#) [Google Scholar ↗](#)
- [71] I. Pilchova, K. Klacanova, Z. Tatarkova, P. Kaplan, P. Racay
The involvement of Mg²⁺ in regulation of cellular and mitochondrial functions
Oxid. Med. Cell. Longev., 2017 (2017), pp. 1-8
[Crossref ↗](#) [Google Scholar ↗](#)
- [72] S.L. Howell, M. Tyhurst
Barium accumulation in rat pancreatic B cells
J. Cell Sci., 22 (1976), pp. 455-465
[Crossref ↗](#) [View in Scopus ↗](#) [Google Scholar ↗](#)

- [73] A.V. Somlyo, A.P. Somlyo
Strontium accumulation by sarcoplasmic reticulum and mitochondria in vascular smooth muscle
Science, 174 (1971), pp. 955-958
[Crossref ↗](#) [View in Scopus ↗](#) [Google Scholar ↗](#)
- [74] N.D. Jhurry, M. Chakrabarti, S.P. McCormick, G.P. Holmes-Hampton, P.A. Lindahl
Biophysical investigation of the ironome of human jurkat cells and mitochondria
Biochemistry, 51 (2012), pp. 5276-5284
[Crossref ↗](#) [View in Scopus ↗](#) [Google Scholar ↗](#)
- [75] D.M. Ward, S.M. Cloonan
Mitochondrial iron in human health and disease
Annu. Rev. Physiol., 81 (2019), pp. 453-482
[Crossref ↗](#) [View in Scopus ↗](#) [Google Scholar ↗](#)
- [76] H. Yoon, *et al.*
Rim2, a pyrimidine nucleotide exchanger, is needed for iron utilization in mitochondria
Biochem. J., 440 (2011), pp. 137-146
[View in Scopus ↗](#) [Google Scholar ↗](#)
- [77] D.R. Richardson, *et al.*
Mitochondrial iron trafficking and the integration of iron metabolism between the mitochondrion and cytosol
Proc. Natl. Acad. Sci. U. S. A., 107 (2010), pp. 10775-10782
[Crossref ↗](#) [View in Scopus ↗](#) [Google Scholar ↗](#)
- [78] S. Levi, *et al.*
A human mitochondrial ferritin encoded by an intronless gene
J. Biol. Chem., 276 (2001), pp. 24437-24440
 [View PDF](#) [View article](#) [View in Scopus ↗](#) [Google Scholar ↗](#)
- [79] B.T. Paul, D.H. Manz, F.M. Torti, S.V. Torti
Mitochondria and Iron: current questions
Expet Rev. Hematol., 10 (2017), pp. 65-79
[Crossref ↗](#) [View in Scopus ↗](#) [Google Scholar ↗](#)
- [80] M. Ristow, *et al.*

Frataxin activates mitochondrial energy conversion and oxidative phosphorylation

Proc. Natl. Acad. Sci. Unit. States Am., 97 (2000), pp. 12239-12243

[View in Scopus ↗](#) [Google Scholar ↗](#)

[81] D. Galaris, A. Barbouti, K. Pantopoulos

Iron homeostasis and oxidative stress: an intimate relationship

Biochim. Biophys. Acta BBA - Mol. Cell Res., 1866 (2019), p. 118535

 [View PDF](#) [View article](#) [View in Scopus ↗](#) [Google Scholar ↗](#)

[82] W. Xu, T. Barrientos, N.C. Andrews

Iron and copper in mitochondrial diseases

Cell Metabol., 17 (2013), pp. 319-328

 [View PDF](#) [View article](#) [View in Scopus ↗](#) [Google Scholar ↗](#)

[83] B.L. Vallee, K.H. Falchuk

The biochemical basis of zinc physiology

Physiol. Rev., 73 (1993), pp. 79-118

[Crossref ↗](#) [View in Scopus ↗](#) [Google Scholar ↗](#)

[84] Modern Nutrition in Health and Disease, Wolters Kluwer Health/Lippincott Williams & Wilkins (2014)

[Google Scholar ↗](#)

[85] Y.A. Seo, V. Lopez, S.L. Kelleher

A histidine-rich motif mediates mitochondrial localization of ZnT2 to modulate mitochondrial function

Am. J. Physiol. Cell Physiol., 300 (2011), pp. C1479-C1489

[Crossref ↗](#) [View in Scopus ↗](#) [Google Scholar ↗](#)

[86] M. Yamaguchi, M. Kura, S. Okada

Zinc accumulation and succinate dehydrogenase activation in hepatic mitochondria of rats orally administered zinc sulfate

Chem. Pharm. Bull. (Tokyo), 29 (1981), pp. 2370-2374

[Crossref ↗](#) [View in Scopus ↗](#) [Google Scholar ↗](#)

[87] B. Ye, W. Maret, B.L. Vallee

Zinc metallothionein imported into liver mitochondria modulates respiration

Proc. Natl. Acad. Sci. Unit. States Am., 98 (2001), pp. 2317-2322

[View in Scopus ↗](#) [Google Scholar ↗](#)

- [88] A. Okado-Matsumoto, I. Fridovich
Subcellular distribution of superoxide dismutases (SOD) in rat liver
J. Biol. Chem., 276 (2001), pp. 38388-38393

 [View PDF](#) [View article](#) [View in Scopus ↗](#) [Google Scholar ↗](#)

- [89] D. Marreiro, *et al.*
Zinc and oxidative stress: current mechanisms
Antioxidants, 6 (2017), p. 24

[Crossref ↗](#) [View in Scopus ↗](#) [Google Scholar ↗](#)

- [90] J. Wudarczyk, G. Dębska, E. Lenartowicz
Zinc as an inducer of the membrane permeability transition in rat liver mitochondria
Arch. Biochem. Biophys., 363 (1999), pp. 1-8

 [View PDF](#) [View article](#) [View in Scopus ↗](#) [Google Scholar ↗](#)

- [91] H.Y. Liu, J.R. Gale, I.J. Reynolds, J.H. Weiss, E. Aizenman
The multifaceted roles of zinc in neuronal mitochondrial dysfunction
Biomedicines, 9 (2021), p. 489

[Google Scholar ↗](#)

- [92] S.L. Sensi, *et al.*
Modulation of mitochondrial function by endogenous Zn²⁺ pools
Proc. Natl. Acad. Sci. Unit. States Am., 100 (2003), pp. 6157-6162

[View in Scopus ↗](#) [Google Scholar ↗](#)

- [93] A. Boulet, *et al.*
The mammalian phosphate carrier SLC25A3 is a mitochondrial copper transporter required for cytochrome c oxidase biogenesis
J. Biol. Chem., 293 (2018), pp. 1887-1896

 [View PDF](#) [View article](#) [View in Scopus ↗](#) [Google Scholar ↗](#)

- [94] S. Blockhuys, *et al.*
Defining the human copper proteome and analysis of its expression variation in cancers
Metallomics, 9 (2017), pp. 112-123

[View in Scopus ↗](#) [Google Scholar ↗](#)

- [95] L. Gaetke

Copper toxicity, oxidative stress, and antioxidant nutrients

Toxicology, 189 (2003), pp. 147-163

 [View PDF](#) [View article](#) [View in Scopus ↗](#) [Google Scholar ↗](#)

[96]

L.M. Ruiz, A. Libedinsky, A.A. Elorza

Role of copper on mitochondrial function and metabolism

Front. Mol. Biosci., 8 (2021), p. 711227

[View in Scopus ↗](#) [Google Scholar ↗](#)

[97]

H. Zischka, J. Lichtmannegger

Pathological mitochondrial copper overload in livers of Wilson's disease patients and related animal models: mitochondrial copper in WD

Ann. N. Y. Acad. Sci., 1315 (2014), pp. 6-15

[Crossref ↗](#) [View in Scopus ↗](#) [Google Scholar ↗](#)

[98]

R.I. Bustos, *et al.*

Copper deficiency alters cell bioenergetics and induces mitochondrial fusion through up-regulation of MFN2 and OPA1 in erythropoietic cells

Biochem. Biophys. Res. Commun., 437 (2013), pp. 426-432

 [View PDF](#) [View article](#) [View in Scopus ↗](#) [Google Scholar ↗](#)

[99]

Z.N. Baker, P.A. Cobine, S.C. Leary

The mitochondrion: a central architect of copper homeostasis

Metallomics, 9 (2017), pp. 1501-1512

[View in Scopus ↗](#) [Google Scholar ↗](#)

[100]

B. Chance, L. Mela

Calcium and manganese interactions in mitochondrial ion accumulation

Biochemistry, 5 (1966), pp. 3220-3223

[Crossref ↗](#) [View in Scopus ↗](#) [Google Scholar ↗](#)

[101]

M.R. Smith, J. Fernandes, Y.-M. Go, D.P. Jones

Redox dynamics of manganese as a mitochondrial life-death switch

Biochem. Biophys. Res. Commun., 482 (2017), pp. 388-398

 [View PDF](#) [View article](#) [View in Scopus ↗](#) [Google Scholar ↗](#)

[102]

D. Hu, *et al.*

Hippocampal long-term potentiation, memory, and longevity in mice that overexpress mitochondrial superoxide dismutase

Neurobiol. Learn. Mem., 87 (2007), pp. 372-384

 [View PDF](#) [View article](#) [View in Scopus ↗](#) [Google Scholar ↗](#)

[103] X. Zou, *et al.*

Manganese superoxide dismutase (SOD2): is there a center in the universe of mitochondrial redox signaling?

J. Bioenerg. Biomembr., 49 (2017), pp. 325-333

[Crossref ↗](#) [View in Scopus ↗](#) [Google Scholar ↗](#)

[104] S.F. Ali, H.M. Duhart, G.D. Newport, G.W. Lipe, W. Slikker

Manganese-induced reactive oxygen species: comparison between Mn⁺² and Mn⁺³

Neurodegeneration, 4 (1995), pp. 329-334

 [View PDF](#) [View article](#) [View in Scopus ↗](#) [Google Scholar ↗](#)

[105] L. Li, X. Yang

The essential element manganese, oxidative stress, and metabolic diseases: links and interactions

Oxid. Med. Cell. Longev., 2018 (2018), pp. 1-11

[Google Scholar ↗](#)

[106] R.R. Mendel

The molybdenum cofactor

J. Biol. Chem., 288 (2013), pp. 13165-13172

 [View PDF](#) [View article](#) [Crossref ↗](#) [View in Scopus ↗](#) [Google Scholar ↗](#)

[107] G. Schwarz, R.R. Mendel, M.W. Ribbe

Molybdenum cofactors, enzymes and pathways

Nature, 460 (2009), pp. 839-847

[Crossref ↗](#) [View in Scopus ↗](#) [Google Scholar ↗](#)

[108] M. Grings, *et al.*

ETHE1 and MOCS1 deficiencies: disruption of mitochondrial bioenergetics, dynamics, redox homeostasis and endoplasmic reticulum-mitochondria crosstalk in patient fibroblasts

Sci. Rep., 9 (2019), p. 12651

[View in Scopus ↗](#) [Google Scholar ↗](#)

[109] H. Zhao, K. Ruberu, H. Li, B. Garner

Analysis of subcellular [⁵⁷Co] cobalamin distribution in SH-SY5Y neurons and brain tissue

J. Neurosci. Methods, 217 (2013), pp. 67-74

 [View PDF](#) [View article](#) [View in Scopus ↗](#) [Google Scholar ↗](#)

- [110] J.J.E. Janssen, S. Grefte, J. Keijer, V.C.J. de Boer
Mito-nuclear communication by mitochondrial metabolites and its regulation by B-vitamins

Front. Physiol., 10 (2019), p. 78

[View in Scopus ↗](#) [Google Scholar ↗](#)

- [111] A. Pastore, *et al.*
Glutathione metabolism in cobalamin deficiency type C (cb1C)

J. Inherit. Metab. Dis., 37 (2014), pp. 125-129

[Crossref ↗](#) [View in Scopus ↗](#) [Google Scholar ↗](#)

- [112] J.B. Vincent
Elucidating a biological role for chromium at a molecular level

Acc. Chem. Res., 33 (2000), pp. 503-510

[View in Scopus ↗](#) [Google Scholar ↗](#)

- [113] H. Wang, A. Kruszewski, D.L. Brautigan
Cellular chromium enhances activation of insulin receptor kinase

Biochemistry, 44 (2005), pp. 8167-8175

[Crossref ↗](#) [View in Scopus ↗](#) [Google Scholar ↗](#)

- [114] Y. Hua, S. Clark, J. Ren, N. Sreejayan
Molecular mechanisms of chromium in alleviating insulin resistance

J. Nutr. Biochem., 23 (2012), pp. 313-319

 [View PDF](#) [View article](#) [View in Scopus ↗](#) [Google Scholar ↗](#)

- [115] T. Yu, *et al.*
A reversible and highly selective fluorescence “on-off-on” probe for detecting nickel ion in the mitochondria of living cells

Biosens. Bioelectron., 82 (2016), pp. 93-98

 [View PDF](#) [View article](#) [View in Scopus ↗](#) [Google Scholar ↗](#)

- [116] S. Ghazali, J. Fan, J. Du, X. Peng
Mito-targeted “turn-on” fluorescent probe for nickel (II) detection

Methods, 168 (2019), pp. 24-28

 [View PDF](#) [View article](#) [View in Scopus ↗](#) [Google Scholar ↗](#)

- [117] C.E. Nichols, *et al.*

Reactive oxygen species damage drives cardiac and mitochondrial dysfunction following acute nano-titanium dioxide inhalation exposure

Nanotoxicology, 12 (2018), pp. 32-48

[Crossref ↗](#) [View in Scopus ↗](#) [Google Scholar ↗](#)

[118] R.P. Sharma, S.G. Oberg, R.D.R. Parker

Vanadium retention in rat tissues following acute exposures to different dose levels

J. Toxicol. Environ. Health, 6 (1980), pp. 45-54

[Crossref ↗](#) [Google Scholar ↗](#)

[119] M. Aureliano, C.A. Ohlin

Decavanadate in vitro and in vivo effects: facts and opinions

J. Inorg. Biochem., 137 (2014), pp. 123-130

 [View PDF](#) [View article](#) [View in Scopus ↗](#) [Google Scholar ↗](#)

[120] C. Chen

Subcellular distribution of selenium and Se-containing proteins in human liver

Biochim. Biophys. Acta BBA - Gen. Subj., 1427 (1999), pp. 205-215

 [View PDF](#) [View article](#) [View in Scopus ↗](#) [Google Scholar ↗](#)

[121] E. Mangiapane, A. Pessione, E. Pessione

Selenium and selenoproteins: an overview on different biological systems

Curr. Protein Pept. Sci., 15 (2014), pp. 598-607

[Crossref ↗](#) [View in Scopus ↗](#) [Google Scholar ↗](#)

[122] D.L. Hatfield, P.A. Tsuji, B.A. Carlson, V.N. Gladyshev

Selenium and selenocysteine: roles in cancer, health, and development

Trends Biochem. Sci., 39 (2014), pp. 112-120

 [View PDF](#) [View article](#) [View in Scopus ↗](#) [Google Scholar ↗](#)

[123] S.L. Mehta, S. Kumari, N. Mendelev, P.A. Li

Selenium preserves mitochondrial function, stimulates mitochondrial biogenesis, and reduces infarct volume after focal cerebral ischemia

BMC Neurosci., 13 (2012), p. 79

[View in Scopus ↗](#) [Google Scholar ↗](#)

[124] H.J. Thompson

Role of low molecular weight, selenium-containing compounds in human health

D.L. Hatfield (Ed.), Selenium, Springer US (2001), pp. 283-297, [10.1007/978-1-4615-1609-5_23](https://doi.org/10.1007/978-1-4615-1609-5_23) ↗

[Google Scholar](#) ↗

- [125] F.H. Nielsen, T.R. Shuler
Studies of the interaction between boron and calcium, and its modification by magnesium and potassium, in rats: effects on growth, blood variables, and bone mineral composition

Biol. Trace Elem. Res., 35 (1992), pp. 225-237

[View in Scopus](#) ↗ [Google Scholar](#) ↗

- [126] F.H. Nielsen
Biochemical and physiologic consequences of boron deprivation in humans
Environ. Health Perspect., 102 (1994), pp. 59-63

[View in Scopus](#) ↗ [Google Scholar](#) ↗

- [127] Dietary reference intakes for vitamin C, vitamin E, selenium, and carotenoids: a report of the panel on dietary antioxidants and related compounds, subcommittees on upper reference levels of nutrients and of interpretation and use of dietary reference intakes, and the standing committee on the scientific evaluation of dietary reference intakes

Food and Nutrition Board, Institute of Medicine, National Academy Press (2000)

[Google Scholar](#) ↗

- [128] F.H. Nielsen
Manganese, molybdenum, boron, chromium, and other trace elements
J.W. Erdman, S.H. Zeisel (Eds.), Present Knowledge in Nutrition, Wiley-Blackwell (2012), pp. 586-607

[Crossref](#) ↗ [View in Scopus](#) ↗ [Google Scholar](#) ↗

- [129] A. Basoglu, N. Baspinar, A.S. Ozturk, P.P. Akalin
Effects of long-term boron administrations on high-energy diet-induced obesity in rabbits: NMR-based metabonomic evaluation

J. Anim. Vet. Adv., 10 (2011), pp. 1512-1515

[View in Scopus](#) ↗ [Google Scholar](#) ↗

- [130] H. Khaliq, Z. Juming, P. Ke-Mei
The physiological role of boron on health

Biol. Trace Elem. Res., 186 (2018), pp. 31-51

[Crossref ↗](#) [View in Scopus ↗](#) [Google Scholar ↗](#)

- [131] R.N. Johnson, B.E. Volcani
The uptake of silicic acid by rat liver mitochondria
Biochem. J., 172 (1978), pp. 557-568

[Crossref ↗](#) [View in Scopus ↗](#) [Google Scholar ↗](#)

- [132] P.Y. Niu, *et al.*
Aluminum impairs rat neural cell mitochondria *in vitro*
Int. J. Immunopathol. Pharmacol., 18 (2005), pp. 683-689

[Crossref ↗](#) [View in Scopus ↗](#) [Google Scholar ↗](#)

- [133] A.H. Rashedy, A.A. Solimany, A.K. Ismail, M.H. Wahdan, K.A. Saban
Histopathological and functional effects of antimony on the renal cortex of growing albino rat
Int. J. Clin. Exp. Pathol., 6 (2013), pp. 1467-1480

[View in Scopus ↗](#) [Google Scholar ↗](#)

- [134] F.H. Nielsen
Nutritional significance of the ultratrace elements
Nutr. Rev., 46 (2009), pp. 337-341

[Crossref ↗](#) [Google Scholar ↗](#)

- [135] M.A. Peraza, D.W. Cromey, B. Carolus, D.E. Carter, A.J. Gandolfi
Morphological and functional alterations in human proximal tubular cell line induced by low level inorganic arsenic: evidence for targeting of mitochondria and initiated apoptosis
J. Appl. Toxicol., 26 (2006), pp. 356-367

[Crossref ↗](#) [View in Scopus ↗](#) [Google Scholar ↗](#)

- [136] X. Gao, *et al.*
An *in vitro* study on the cytotoxicity of bismuth oxychloride nanosheets in human HaCaT keratinocytes
Food Chem. Toxicol., 80 (2015), pp. 52-61

 [View PDF](#) [View article](#) [View in Scopus ↗](#) [Google Scholar ↗](#)

- [137] L. Li, *et al.*
Analysis of blood concentrations of zinc, germanium, and lead and relevant environmental factors in a population sample from Shandong province, China

[138] J.M. Cho, *et al.*

Immune activation of Bio-Germanium in a randomized, double-blind, placebo-controlled clinical trial with 130 human subjects: therapeutic opportunities from new insights

PLoS One, 15 (2020), Article e0240358

[Crossref ↗](#) [View in Scopus ↗](#) [Google Scholar ↗](#)

[139] I. Higuchi, *et al.*

Germanium myopathy: clinical and experimental pathological studies

Acta Neuropathol., 79 (1989), pp. 300-304

[View in Scopus ↗](#) [Google Scholar ↗](#)

[140] R.R. Harvey, M.A. Virji, N.T. Edwards, K.J. Cummings

Comparing plasma, serum and whole blood indium concentrations from workers at an indium-tin oxide (ITO) production facility

Occup. Environ. Med., 73 (2016), pp. 864-867, [10.1136/oemed-2016-103685 ↗](#)

[Google Scholar ↗](#)

[141] N. Liu, Y. Guan, B. Li, S. Yao

Biomonitorization of concentrations of 28 elements in serum and urine among workers exposed to indium compounds

PLoS One, 16 (2021), Article e0246943

[Crossref ↗](#) [View in Scopus ↗](#) [Google Scholar ↗](#)

[142] M. Van Hulle, K. De Cremer, R. Cornelis, N. Lameire

In vivo distribution and speciation of [^{114m}In]InCl₃ in the Wistar rat

J. Environ. Monit., 3 (2001), pp. 86-90

[View in Scopus ↗](#) [Google Scholar ↗](#)

[143] M. Samira, A. Ahlem, B.A. Aouatef, J.M. Habib, T. Leila

Histological and ultrastructural study of the intracellular behavior of indium in the testicular tissues

Microsc. Res. Tech., 74 (2011), pp. 546-550

[Crossref ↗](#) [View in Scopus ↗](#) [Google Scholar ↗](#)

[144] K. Gao, *et al.*

The role of endoplasmic reticulum stress in lead (Pb)-induced mitophagy of HEK293 cells

Toxicol. Ind. Health, 36 (2020), pp. 1002-1009

[Crossref ↗](#) [View in Scopus ↗](#) [Google Scholar ↗](#)

[145] S. Fani, *et al.*

Anticancer activity of a monobenzyltin complex C1 against MDA-MB-231 cells through induction of Apoptosis and inhibition of breast cancer stem cells

Sci. Rep., 6 (2016), p. 38992

[View in Scopus ↗](#) [Google Scholar ↗](#)

[146] M. Calvo-Rodriguez, *et al.*

Increased mitochondrial calcium levels associated with neuronal death in a mouse model of Alzheimer's disease

Nat. Commun., 11 (2020), p. 2146

[View in Scopus ↗](#) [Google Scholar ↗](#)

[147] C.D. Folmes, H. Ma, S. Mitalipov, A. Terzic

Mitochondria in pluripotent stem cells: stemness regulators and disease targets

Curr. Opin. Genet. Dev., 38 (2016), pp. 1-7



[View PDF](#) [View article](#) [View in Scopus ↗](#) [Google Scholar ↗](#)

[148] R.P. Chakrabarty, N.S. Chandel

Mitochondria as signaling organelles control mammalian stem cell fate

Cell Stem Cell, 28 (2021), pp. 394-408



[View PDF](#) [View article](#) [View in Scopus ↗](#) [Google Scholar ↗](#)

[149] A. Prigione, B. Fauler, R. Lurz, H. Lehrach, J. Adjaye

The senescence-related mitochondrial/oxidative stress pathway is repressed in human induced pluripotent stem cells

Stem Cell., 28 (2010), pp. 721-733



[Crossref ↗](#) [View in Scopus ↗](#) [Google Scholar ↗](#)

[150] H. Kondoh, *et al.*

A high glycolytic flux supports the proliferative potential of murine embryonic stem cells

Antioxidants Redox Signal., 9 (2007), pp. 293-299

[Crossref ↗](#) [View in Scopus ↗](#) [Google Scholar ↗](#)

- [151] E. Dernbach, *et al.*
Antioxidative stress–associated genes in circulating progenitor cells: evidence for enhanced resistance against oxidative stress
Blood, 104 (2004), pp. 3591-3597
 [View PDF](#) [View article](#) [View in Scopus ↗](#) [Google Scholar ↗](#)
- [152] B. Dannenmann, S. Lehle, F. Essmann, K. Schulze-Osthoff
Genome surveillance in pluripotent stem cells: low apoptosis threshold and efficient antioxidant defense
Mol. Cell. Oncol., 3 (2016), Article e1052183
[Crossref ↗](#) [View in Scopus ↗](#) [Google Scholar ↗](#)
- [153] Y. Cao, *et al.*
ROS functions as an upstream trigger for autophagy to drive hematopoietic stem cell differentiation
Hematology, 21 (2016), pp. 613-618
[Crossref ↗](#) [View in Scopus ↗](#) [Google Scholar ↗](#)
- [154] L.L. Luchsinger, *et al.*
Harnessing hematopoietic stem cell low intracellular calcium improves their maintenance in vitro
Cell Stem Cell, 25 (2019), pp. 225-240.e7
 [View PDF](#) [View article](#) [View in Scopus ↗](#) [Google Scholar ↗](#)
- [155] J.R. Osete, *et al.*
Lithium increases mitochondrial respiration in iPSC-derived neural precursor cells from lithium responders
Mol. Psychiatr. (2021), [10.1038/s41380-021-01164-4 ↗](#)
[Google Scholar ↗](#)
- [156] S. Jia, C. Mou, Y. Ma, R. Han, X. Li
Magnesium regulates neural stem cell proliferation in the mouse hippocampus by altering mitochondrial function: magnesium regulates neural stem cell proliferation
Cell Biol. Int., 40 (2016), pp. 465-471
[Crossref ↗](#) [View in Scopus ↗](#) [Google Scholar ↗](#)
- [157] R.S. Corniola, N.M. Tassabehji, J. Hare, G. Sharma, C.W. Levenson

Zinc deficiency impairs neuronal precursor cell proliferation and induces apoptosis via p53-mediated mechanisms

Brain Res., 1237 (2008), pp. 52-61



[View PDF](#) [View article](#) [View in Scopus ↗](#) [Google Scholar ↗](#)

[158] S.W. Suh, *et al.*

Decreased brain zinc availability reduces hippocampal neurogenesis in mice and rats

J. Cerebr. Blood Flow Metabol., 29 (2009), pp. 1579-1588

[Crossref ↗](#) [View in Scopus ↗](#) [Google Scholar ↗](#)

[159] R. Seth, *et al.*

Zinc deficiency induces apoptosis via mitochondrial p53- and caspase-dependent pathways in human neuronal precursor cells

J. Trace Elem. Med. Biol., 30 (2015), pp. 59-65



[View PDF](#) [View article](#) [View in Scopus ↗](#) [Google Scholar ↗](#)

[160] J. Hu, *et al.*

Zinc chloride transiently maintains mouse embryonic stem cell pluripotency by activating Stat3 signaling

PLoS One, 11 (2016), Article e0148994

[Crossref ↗](#) [View in Scopus ↗](#) [Google Scholar ↗](#)

[161] L.M. Ruiz, *et al.*

Non-cytotoxic copper overload boosts mitochondrial energy metabolism to modulate cell proliferation and differentiation in the human erythroleukemic cell line K562

Mitochondrion, 29 (2016), pp. 18-30



[View PDF](#) [View article](#) [View in Scopus ↗](#) [Google Scholar ↗](#)

[162] T. Peled, E. Landau, E. Prus, A.J. Treves, E. Fibach

Cellular copper content modulates differentiation and self-renewal in cultures of cord blood-derived CD34⁺ cells: copper Modulates Differentiation/Self-renewal of CD34 Cells

Br. J. Haematol., 116 (2002), pp. 655-661

[Google Scholar ↗](#)

[163] C. Tamm, F. Sabri, S. Ceccatelli

Mitochondrial-Mediated apoptosis in neural stem cells exposed to manganese

Toxicol. Sci., 101 (2008), pp. 310-320

[Crossref ↗](#) [View in Scopus ↗](#) [Google Scholar ↗](#)

[164] G. Murdolo, *et al.*

Selenium and cancer stem cells

Advances in Cancer Research, Elsevier (2017), pp. 136 235-257

[Google Scholar ↗](#)

[165] Y. Zhang, Zheng

J. Bioinformatics of metalloproteins and metalloproteomes

Molecules, 25 (2020), p. 3366

[Google Scholar ↗](#)

Cited by (19)

[Osteogenesis and angiogenesis promoting bioactive ceramics](#)

2024, Materials Science and Engineering R: Reports

[Show abstract](#) ✓

[Development and characterization of antibacterial 3D porous hydroxyapatite-gelatin-PVA scaffolds containing zinc oxide nanoparticles](#)

2024, Materials Chemistry and Physics

[Show abstract](#) ✓

[Associations of essential trace elements with epigenetic aging indicators and the potential mediating role of inflammation](#)

2023, Redox Biology

[Show abstract](#) ✓

[A big picture of the mitochondria-mediated signals: From mitochondria to organism](#)

2023, Biochemical and Biophysical Research Communications

[Show abstract](#) ✓

Impacts of iron on ultrastructural features of NCI-H295R cell line related to steroidogenesis

2023, Acta Histochemica

[Show abstract](#) ✓

Nutraceuticals and food supplements: Basic concepts and regulatory aspects ↗

2024, Nutraceuticals: A Holistic Approach to Disease Prevention



[View all citing articles on Scopus](#) ↗

- ☆ This article is a contribution to the special issue entitled “Mitochondrial Redox Signaling in Aging-Related Diseases: In Honor of Bruce N. Ames: A Pioneer in Mitochondrial Redox Signaling in Aging Research” Guest Edited by Prof. Jiankang Liu & Prof. Douglas C. Wallace.

© 2022 The Authors. Published by Elsevier Inc.



All content on this site: Copyright © 2024 Elsevier B.V., its licensors, and contributors. All rights are reserved, including those for text and data mining, AI training, and similar technologies. For all open access content, the Creative Commons licensing terms apply.

